

Traditional Chinese Medicine

Plant distribution and pharmacological activity of flavonoids

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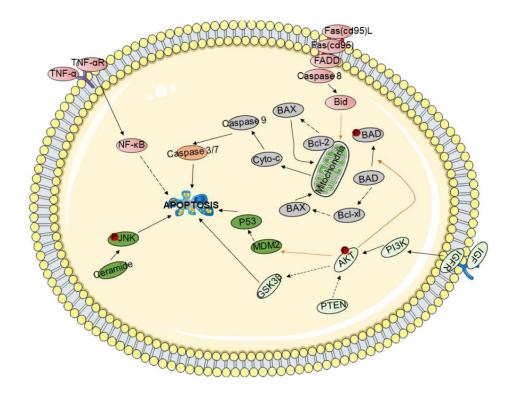
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Highlights

This review covers the plant distribution and pharmacological activities of flavonoids, stressing the importance of identifying such valuable flavonoids in another genus or family while providing a basis for fully exploiting the therapeutic potential of flavonoids.

Traditionality

Flavonoids are found in some traditional Chinese medicines that function to clear heat and dampness, some pathological products resulted from diseases. The most representative drugs among them are Huangqin (Scutellaria baicalensis), Chuanhuangbai (Phellodendri Chinensis Cortex), and Kushen (Sophora flavescens). As early as the Donghan dynasty of China, these three herbs were recorded in an ancient book of Chinese medicine called Shennong Bencao Jing.





Abstract

Flavonoids are natural organic compounds that are widely found in nature, their structural types are complex, and they mainly include flavonoids, flavonols, dihydroflavonols, isoflavones, dihydroisoflavones, chalcones, orange ketones, flavanoids, anthocyanidins, and biflavonoids. This review covers the plant distribution and pharmacological activities of flavonoids. Flavonoids are mainly distributed in angiosperms and gymnosperms, and they are abundant in plants such as Rutaceae, Labiatae, Zingiberaceae, Scrophulariaceae, and Leguminosae. Because of their wide distribution and variety, researchers have found that flavonoids have diverse biological activities, mainly focusing on anti-inflammatory, antibacterial and antitumor activities. Mechanistically, the anti-inflammatory effects are mainly related to the NF-κB and MAPK (mitogen-activated protein kinase) signaling pathway and then the inhibition of the production of inflammatory cytokines and mediators. The antibacterial activity is mainly manifested as inhibitory effects on many strains, including Escherichia coli, Cryptococcus neoformans, and Pseudomonas aeruginosa, via destroying the stability of the microbial membrane, inhibiting the invasion of virulent bacteria into host cells, promoting the apoptosis of bacteria, inhibiting bacterial fatty acid synthesis, etc. The antitumor activity of flavonoids is related to their inhibition of cell proliferation and induction of apoptosis via the mitochondria-mediated, endoplasmic reticulum-mediated, and death factor and its receptor-mediated signal transduction pathways. Understanding the plant distribution and pharmacological activity of flavonoids not only reveals the importance of identifying such valuable flavonoids in another genus or family but also provides a basis for fully exploiting the therapeutic potential of flavonoids.

Keywords: Flavonoids, Plant distribution, Pharmacological activity, Antitumor, Anti-inflammatory, Antibacterial.

Acknowledgments:

This study was financially supported by the Autonomous Foundation of Key Laboratory of Ethnomedicine (Minzu University of China), Ministry of Education (No. KLEM-ZZ201902), the National Natural Science Foundation of China Grants (No. 81973977), and the National Training Programs of Innovation and Entrepreneurship for Undergraduates (No. GCYS2018110001).

Abbreviations:

ROS, Reactive oxygen species; Bcl-2, B-cell lymphoma 2; BAX, Bcl-2 associated X protein; PERK, Protein kinase R-like ER kinase; ATF, Activated transcription factor; IRE1, Inositol-requiring enzyme-1; XBP1, X-box binding protein 1; MMP-2: Matrix metalloproteinase-2, MMP-9: Matrix metalloproteinase-2; PARP: Poly (ADP-ribose) polymerase; Akt: Protein kinase B; DPPH, 1,1-Diphenyl-2-picrylhydrazyl; TLR: Toll-like receptor; RIG, Retinoic acid-inducible gene; AQP, Aquaporin; iNOS, Inducible nitric oxide synthase; COX, Cyclooxygenase; IL, Interleukin; AFC, Alveolar fluid clearance; Nrf2: Nuclear factor erythroid-2-related factor 2

Competing interests:

The authors declare that they have no conflict of interest.

Citation:

Shao-Hui Wang, Yan-Lan Hu, Tong-Xiang Liu. Plant distribution and pharmacological activity of flavonoids. Traditional Medicine Research 2019, 4 (5): 269-287.

Executive Editor: Nuo-Xi Pi.

Submitted: 2 August 2019, Accepted: 26 August 2019, Online: 5 September 2019.

Background

Flavonoids are an important class of natural organic compounds that includes more than 4,000 polyphenolic compounds that are widely found in various natural plants [1]. Flavonoids are also present in some traditional Chinese medicines. The most representative drugs among them are Huangqin (*Scutellaria baicalensis*), Chuanhuangbai (*Phellodendron chinense Schneid*), and Kushen (*Sophora flavescens*). As early as the Donghan dynasty of China, these three herbs were recorded in an ancient book of Chinese medicine called *Shennong Bencao Jing*.

The C₆-C₃-C₆ skeleton is formed in plants by the condensation of three malonyl residues with hydroxy cinnamic acid [2], labeled as rings A, B, and C. Currently, flavonoids are roughly classified into 10 categories based on their structure, including flavonoids, flavonols, dihydroflavonols, isoflavones, dihydroisoflavones, chalcones, orange ketones, flavanoids, anthocyanidins, and biflavonoids. In addition, the 5 and 7 positions of ring A; 3', 4', and 5'

positions of ring B; 3 and 2 positions of ring C of flavonoids are often replaced by hydroxyl groups, these readily form glycosides with a variety of five- or six-carbon sugars through a β glycoside bond, most of which are oxyglycosides, such as 7-o-glycosides in flavones, flavanones, and isoflavones. Compared with oxygen glycosides, which have been widely studied, there are relatively few studies on carbon glycosides in the literature [3]. Carbon glycosides can be directly connected to the skeleton through acid-resistant C-C bonds, and they mainly occur at the C₆ and C₈ positions. The basic structures and glycosylation sites of common flavonoids are summarized in Figure 1.

Because of their wide distribution and variety, flavonoids have been found to possess diverse biological activities, such as antioxidation, antibacterial. antihypertensive, protection, liver antitumor, and neuroprotection effects. In this review, the plant distribution and pharmacological activities of flavonoids are systematically summarized, and the results not only reveal the importance of identifying such valuable flavonoids in another genus or family but also provide a basis for fully exploiting the therapeutic potential of flavonoids.

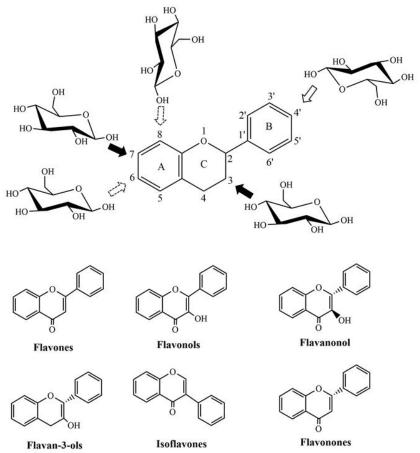


Figure 1 Basic structure of the common classes of flavonoids and the common points of glycosylation Common glycosylation points are C_3 and C_7 (black arrows); B ring glycosylation is also observed in some plants (hollow arrow); C-glycosides are the rarest in plants (arrow with dotted lines); C-glycosylation mostly occurs at the C_6 and C_8 positions.



Plant distribution of flavonoids

A search in *Phenol-Explorer* (http://phenol-explorer.eu/) produced 492 different flavonoids and isoflavone components in 400 types of food; thus, there are even more flavonoids distributed in plants. By sorting through the plant sources of flavonoids, we found that the distribution of flavonoids

was the highest in the families of *Rutaceae*, *Zingiberaceae*, *Scrophulariaceae*, *Leguminosae*, *Ranunculaceae*, *Labiatae*, and *Myrtaceae* (Table 1). Kaempferol, quercetin, apigenin, luteolin, isorhamnetin, and quercetin are the most common chemical constituents. In addition, many new compounds that have been isolated, purified, and identified were all modified from the existing flavonoids such as kaempferol and luteolin.

Table 1 Plant sources of flavonoids

Families	Plants/Sources	Components	References	
Selaginellaceae	Juanbai (Selaginella tamariscina), Cuiyuncao (Selaginella uncinate)	Apigenin, Amentoflavone, Hinokiflavone, Isocrytomerin, Arnentoflavone, Unciflavones A-F	[4, 5]	
Cumuassaaaa	Cobaixa (Platualadus aviantalis)		[6]	
Cupressaceae	Cebaiye (<i>Platycladus orientalis</i>)	Acacetin	[6]	
Gramineae	Oryza sativa	Apigenin, Luteolin, Isovitexin-2'-O-(6'-(E)-ferric-glucopyranosi de, Quercetin, Kaempferol	[7]	
Sinopteridaceae	Libingjinfenjue (Onychium lucidum)	Luteolin-7-glucoside, 2,7-Dimethoxyquercetin, Quercetin-3-glucoside, Onychin	[4]	
Acrostichaceae	Acrostichum aureum	Quercetin-3-O-β-D-glucoside, Quercetin-3-O-β-D-glucosyl-(6→1)-α-L-rha mnoside, Quercetin-3-O-α-L-rhamnoside, Quercetin-3-O-α-L-rhamnosyl-7-O-β-D-glu coside, Kaempferol	[8]	
Polypodiaceae	Youbingshiwei (Pyrrosia petiolosa)	Kaempferol-3-O-β-D-glucopyranoside-7-O- α-L-arabinofuranoside	[9, 10]	
Cycadaceae	Cycas revoluta	Ginkgetin, Bilobetin, Isoginkgetin	[4]	
Ginkgoaceae	Yinxing (Ginkgo biloba)	Quercetin, Kaempferol, Isorhamnetin, Genkwanin, Sciadopitysin, Ginkgetin, Isoginkgetin, Apigenin, Luteolin, Naringenin, Diosmetin	[4, 11, 12]	
Typhaceae	Typha orientalis	Catechin, Isorhamnetin-3-O-neohesperidoside, Isorhamnetin-3-O-β-D-glucoside, Naringenin, Kaempferol, Isorhamnetin	[13]	
Ephedraceae	Mahuang (Ephedra)	Luteolin-7-O-glucuronide flavone, Myricetin 3-rhamnoside	[14]	
Salicaceae	Populus X canadensis, Populus tomentosa	Pinostrobin, Pinocembrin, Chrysin, Galangal-3-methoxy, Quercetin-3,7-dimethoxy, Rhamnetin, Kaempferol-3-methoxy, Apigenin	[4, 15]	
Moraceae	Morus alba, Pijiuhua (Humulus lupulus), Artocarpus lingnanensis	Mul-berin, Morin Hydrate, Cyclomorusin, Cyclomorin, Procyanidin dimer, Quercetin, Kaempferol glycoside, Rhamnoside, 2-Hydroxynaringin 4'-O-β-D-glucopyranoside, 2-Hydroxynaringenin, Anthocyanin-3-O-glucoside, Rutin, Isoquercetin, Anthocyanin-3-O-rutinoside, Pelargonin-3-O-glucoside, Kaempferol	[3, 4, 16-18]	
Polygonaceae	Batiansuanmo (<i>Rumex patientia</i>), Juanjingliao (<i>Fallopia convolvulus</i>), Bianxu (<i>Polygonum aviculare</i>)	Kaempferol, Quercetin-3-O-β-D-glucoside, Isorhamnetin, Kaempferol-3-O-β-D-glucoside	[4, 19]	

Caryophyllaceae	Maicailan (<i>Vaccaria segetalis</i>), <i>Paronychia argentea</i>	Apigenin-6-C-arabinose-glucoside, Apigenin-6-C-diglucoside, Vaccarin	[4, 20]
Ranunculaceae	Jian'eloudoucai (Aquilegia oxysepala), Tiankui (Semiaquilegia adoxoides), Mudanhua (Paeonia suffruticosa), Wutou (Aconitum carmichaelii)	Genkwanin, Luteolin, Apigenin, Sweroside, Tiliroside, Semiaquilinoside, Apigenin-7-O-D-glucoside, Apigenin-7-O-D-neohesperidin, Kaempferol-7-O-D-glucopyranoside, Kaempferol-3-O-D-glucopyranosyl-7-O-D-Pyranoglucoside, Dihydrokaempferol	[4, 21-24]
Berberidaceae	Yinyanghuo (<i>Epimedium</i>)	Baohuoside I/II, Icariin A/B/C/D/E/F/G/H, Icariin, Quercetin, Sagittatoside A/B/C, New icariin, Chuanhuoside, Dehydrated icariin, Epimedium B/C/I, Korean icariin I/II	[4, 25-30]
Geraniaceae	Tianzhukui (Pelargonium hortorum)	Kaempferol, Quercetin	[31]
Crassulaceae	Chuipencao (Sedum sarmentosum Bunge), Hongjingtian (Rhodiola rosea), Fojiacao (Sedum lineare), Aoyejingtian (Sedum emarginatum), Gaoshanhongjingtian (Rhodiola cretinii)	Tricin, Luteolin, Glycyrrhizin, Citrin, Quercetin, Kaempferol, Rutin, Rhodiola rutin, Kaempferol-7-rhamnoside	[4, 32, 33]
Lauraceae	Cryptocar-ya chinensis	Cryochinones A/B/C/D/E/F	[34]
Gentianaceae	Longdancao (<i>Gentiana scabra</i>), Qinjiao (<i>Gentiana macrophylla</i>)	Isophorin-4'-O-glucoside, Isophorin, Isovacilin	[35, 36]
Rosaceae	Rosa davurica, Rosa sericea, Shanzha (Crataegus pinnatifida), Pipa (Eriobotrya japonica)	Quercetin, Kaempferol, Hypericin, Rutin, Hesperidin, Cephalosporin, Cephalosporin, Tiliarin, Vitexin	[4, 37]
Leguminosae	Zhilihuangqi (Astragalus Pall.), Kummerowia striata, Sophora davidii, Huluba (Trigonella foenum-graecum), Oxytropis deflexa, Houguojixueteng (Millettia pachycarpa), Huzhizi (Lespedeza bicolor), Caohongteng (Sargentodoxa cuneata), Gancao (Glycyrrhiza uralensis), Eriosema chinense, Luohuasheng (Arachis Linn.), Oxytropis psamocharis, Kushen (Sophora flavescens), Mihuadou (Spatholobus suberectus), Maoguoyuteng (Derris eriocarpa), Dahongpao (Vigna (Willd.) Ohwi et Ohashi), Buguzhi (Psoralea corylifolia), Ge (Pueraria lobata), Oxytropis falcata, Jixueteng (Millettia reticulata)	3'7-Dihydroxy-2'4'-dimethoxyisoxane, Zapotinin, Astragalin, Isoquercitrin, Rutin, Apigenin, Apigenin-7-O-β-D-glucoside, Kaempferol-3-O-β-D-glucoside, Apigenin-7-O-neohesperidin, Diosmetin, Naringenin, Quercetin, Tricin, Kaempferol, Pachycarin, Orientin, Isoorientin, Isovitexin, Myricetin, Dihydromyricetin, Liquiritin, Isoliquiritin, Liquritigenin, Isoliquiritigenin, Formononetin, Licoricone, Rhamnose isoliquiritin, Rhamnosin-3-O-[(S)-3-hydroxy-3-methyl-g lutaryl (1→6)]-β-D-glucopyranoside, Rhamnoside-3-O-[(S)-3-hydroxy-3-methyl glutaryl (1→6)]-β-D-glucopyranoside, Isorhamnetin-3-O-[(S)-3-hydroxy-3- Methylglutaryl (1→6), Kurarinol, Kurarinol K/H, Trifolirhizin, Kuraridine, Isobavachalcone, Rhamnetin, 2',4'-Dihydroxychalcone, 2',4',β-Trihydroxydihydrochalcone, Bavachinin, (-)- Epicatechin, 5,7,3',5'-Tetrahydroxyflavanone, Genistein	[4, 38-48]
Zygophyllaceae	Duolieluotoupeng (Peganum multisectum)	Acacetin, Deacetyl peganetin, Peganetin, 7,4'-Dihydroxy-3'-methoxyflavone-5-O-ruti noside	[4]
Saururaceae	Yuxingcao (Houttuynia cordata)	Quercetin, Rutin, Hyperoside, Isoquercitrin, Houttuynoid G/H/I/J	[49, 50]
Verbenaceae	Vitex negundo)	Isophorin, Luteolin-7-O-β-D-glucopyranoside, Luteolin-3'-O-β-D-glucopyranoside,	[51]
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Rutaceae	Chuanhuangbai (Phellodendri Chinensis Cortex), Citrus aurantium, Citrus maxima, Citrus sinensis, Huangpi (Clausena lansium), Wuzhuyu (Evodia rutaecarpa), Qianlixiang (Murraya paniculata), Huajiao (Zanthoxylum bungeanum)	Apigenin-7-O-β-D-glucoside Phellamurin, Amuresin, Hyperoside, Phellochinin A, Nobiletin, Xanthoxylin, Vitexin	[4, 52]
Meliaceae	Xiangchun (Toona sinensis)	5,7-Dihydroxy-8-methoxyflavone, Kaempferol	[4]
Rhamnaceae	Poncirus trifoliata, Xizangmaoru (Rhamnella gilgitica), Shuli (Rhamnus davurica)	Dihydrokaempferol, Quercetin, (+)-3,3',5',5,7'-Pentahydroxydihydroflavone, (+)-Dihydromyricetin, Methadone, Aromadendrin, Naringenin, Kaempferol, Quercetin, Taxifoliol	[2, 4]
Lamiaceae	Huaqizhu (Mosla grosseserrata), Huoxiang (Agastache), Huangqin (Scutellaria baicalensis), Xunyicao (Lavandula angustifolia), Jingjie (Nepeta cataria), Shixiangru (Mosla chinensis), Zisu (Perilla frutescens), Xiakucao (Prunella vulgaris), Baihuazhizihua (Dracocephalum heterophyllum)	5-Hydroxy-6,7-dimethylflavone, 5,7-Dihydroxy-4'-methoxyflavone, Carotene, Rhamnolidin, Morin-7-O-β-D-glucoside, Kaempferol-3-O-β-D-glucoside, Acacetin, Baicalein, Baicalin, Apigenin, Luteolin, Quercetin, Naringenin, Myricetin, Wogonoside, Wogonin, Luteolin-7-O-Glucoside, Apigenin-7-O-Glucoside, Chrysoeriol, Orohylin-A, Rutin, Kaempferol, Kaempferol-3-O-glucoside, Anthocyanins, Delphinidin, Trimethyl-delphin-3,5-diglucoside, Acacetin-7-O-β-D-glucopyranoside, Isoorientin, Quercetin-3-O-β-D-rhamnoside,	[4, 53-65]
Scrophulariaceae	Liuchuanyu (<i>Linaria vulgaris</i>), Paotongguo (<i>Paulownia fortune</i>), Maxianhao (<i>Pedicularis resupinata</i>)	Hesperidin, Norwogonin, Diosmetin Pectolinarin, Acacetin, Pectolinarigenin, Hispiduline acetyllinarin, Taxifolin-7-O-α-rhamnoside, Baicalein, 3'-O-methyl-5'-hydroxydiacetone, 3'-O-methyl-5'-O-methyldipyrone, 3'-O-methyldipropanediol, 3'-O-methyl dipyridone, Quercetin, Quercetin-4'-O-D-galactoside, Quercetin-7-O-D-galactoside, Luteolin-5-O-β-D-glucoside, Luteolin-7-O-glucoside, Luteolin, 4'- O-β-D-glucoside, Apigenin, Apigenin-7-O-glucoside,	[4, 66, 67]
Asteraceae	Duoshefeipeng (Erigeron multiradiatus), Beiyeju (Dendranthema indicum), Duantingfeipeng (Erigeron breviscapus), Tianshanxuelian (Saussurea involucrata), Shuifeiji (Silybum marianum), Xuetuzi (Saussurea gossypiphora), Huanghuahao (Artemisia annua), Caiji (Cynara scolymus), Hanliancao (Eclipta prostrata), Liulengju (Laggera alata), Guizhencao (Bidens	Apigenin-7-O-β-glucopyranoside Apigenin, Quercetin, Luteolin, Apigenin-7-O-glucuronide, Isoquercetin, Scutellarin, Baicalein-7-β-D-glucoside, Apigenin, Acacetin-7-O-α-L-rhamnosin (1→6)-β-D-glucoside, Silymarin, Silymarin II, Luteolin-7-glucoside, Myricetin, Apigenin-7-O-glucuronide	[4, 68-73]
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Aquifoliaceae	Ilex centrochinensis, Maodongqing (Ilex pubescens)	Huazhong Holly Flavonoids, Hesperidin, Isosakuranetin	[4, 74]
Myrtaceae	Eucalyptus camaldulensis, Cleistocalyx operculatus, Taojinniang	Quercetin, Rutin, Myricetin, Dihydromyricetin, Kaempferol,	[4, 75, 76]
Liliaceae	(Rhodomyrtus tomentosa) Allium fistulosum, Allium cepa, Aletris spicata, Tufuling (Smilax glabra), Xuejie (Daemonorops draco)	Quercetin-7,4'-diglucoside, Vitexin Alliumoside A, Kaempferol, Quercetin 4'-O-β-D-glucopyranoside, Quercetin-4'-glucoside, Quercetin-3,4'-diglucoside, 5-Hydroxy-7,8,4'-trimethoxyflavone, Amentoflavone, Apigenin, Isoengelitin, Isoastilbin, Taxifolin, Astilbin, Engeletin,	[4, 77-80]
Campanulaceae	Dangshen (Codonopsis pilosula),	Neoastilbin, 2-(3,4-Dihydroxyphenyl)-5,7-dihydroxy-3-(3,4,5-trihydroxy-6-methyl-oxa n-2-yl)oxy-chroman-4-one, (-)-Epicatechin, Naringenin, Dihydrokaempferol, Kukulkanin B, Myricetin, Luteolin, Apigenin, Kaempferol, Wogonin,	[81]
	Banbianlian (Lobelia chinensis)	Quercetin, 5-Hydroxy-4',6,7-trimethoxyflavone, 5-Hydroxy-4',7-dimethoxyflavone, Apigenin-7-O-β-D-glucopyranoside, Codonopilodiynoside C/D/E/F/G, Cordifolioidyne B, Tangshenyne A/B, Buddleoside, Luteolin-3',4'-dimethyl ether-7-O-β-D-glucoside	[]
Zingiberaceae	Shanjiang (Alpinia japonica), Caodoukou (Katsumada galangal), Gaoliangjiang (Alpinia officinarum), Jianghuang (Curcuma longa)	Izalpinin, Alpinone, Rhamnocitrin, Kumatahenin, Alpinetin, Cardamonin, Quercetin, Kaempferol, Quercetin-3-methyl ether, Isorhamnetin, 4'-Methyl kaempferin, Galangin, Galangin-3-methyl ether, Turmerone	[4]
Capparaceae Passifloraceae	Malvastrum coromandelianum Xifanlian (Passiflora coerulea)	Methoxy-trihydroxyflavone Isophorin, 7-O-β-glucosyl-6-C-β-glucosyl luteolin, orientin, 7-O-α-rhamnosyl 6-C-β-glucosyl luteolin	[4] [82]
Ebenaceae Flacourtiaceae	Shiye (Diospyros kaki) Oncoba spinosa	Quercetin, Kaempferol Kaempferol, Quercetin, Apigenin-7-O-β-D-glucuronic acid pyranoside, Quercetin-3-O-β-D-galactopyranoside	[83, 84] [85]
Solanaceae	Suanjiang (Physalis alkekengi), Jindenglong (Calyx seu Fructus Physalis)	Luteolin, Luteolin-7-O-glucopyranoside, Quercetin, Kaempferol, Apigenin-O-β-D-glucopyranoside, Diphosphate-O-β-D-diglucopyranoside, 3',4'-Dimethoxyartem, 5,4',5'-Trihydroxy-7,3'-dimethoxyflavonol, Luteolin-7-O-β-D-glucopyranoside, Luteolin-4'-O-β -D-glucopyranoside, Luteolin-7,4'-dioxy-β-D-glucopyranoside, Luteolin-7,3'-dioxy-β-D-glucopyranoside, 3',7-Dimethylquercetin, 3',4',7-Trimethylquercetin, 3',4',7-Trimethylquercetin, Quercetin-3-O-β-D-glucopyranoside, Quercetin-3,7-dioxo-β-D-glucopyranoside,	[86-88]



		Kaempferol-3-O-β-D-glucose, 3,7-Dioxo-α-L-rhamylacetyl, Kaempferol, Diosmetin-O-β-D-glucopyranoside, 5,4',5'-Trihydroxy-7,3'-dimethoxyflavonol, Chrysoeriol	
Brassicaceae Cynomoriaceae	Arabidopsis thaliana Shuoyang (Cynomorium songaricum)	Kaempferol, Quercetin Epicatechin, Cis-5-deoxypentanoic acid-γ-lactone, (+)-Catechin, Rutin, Naringenin, Isoquercitrin, (-)- Epicatechin-3-O-gallate, Isoquercetin, Luteolin-7-O-glucoside, Naringenin-4'-O-β-glucopyranose, Phloridzin, (-)- Epicatechin, Procyanidin B3/B1/B6, Epicatechin-(4β-8)-epicatechin-(4β-8)-catechin, Catechin-(6'-6)-catechin, Catechin-(6'-8)-catechin, Epicatechin- (4β-6)-epicatechin-(4β-8)-catechin, Cyanidin 3-O-glucoside, Cyanidin 3-O-(6-O-rhamnosylglucoside)	[89] [90, 91]
Loganiaceae	Zuiyucao (Buddleja lindleyana)	Acacetin, Apigenin, Luteolin, Luteolin-7-O-rutinoside, Luteolin-7-O-glucoside	[92, 93]
Oleaceae	Nvzhen (<i>Ligustrum lucidum</i>), Lianqiao (<i>Forsythia suspensa</i>)	Apigenin, Apigenin-7-O-acetyl-β-D-glucoside, Apigenin-7-O-β-D-rutin, Luteolin, Luteolin-7-O-β-D-glucopyranoside, Quercetin, Rutin, Baicalin	[94-96]
Convolvulaceae	Tusizi (Cuscuta chinensis)	Quercetin 3-O-β-D-galactoside-7-O-β-D-glucoside, Quercetin 3-O-β-D-ribofuranosyl-(1→2)-β-D -galactoside, Hyperoside, Quercetin, Kaempferol	[97]
Thymelaeaceae	Daphne giraldii, Legewang (Wikstroemia indica), Langdu (Stellera chamaejasme)	Luteolin-7-methylether-5-β-D-glucoside, 4',5-Dihydroxy-3',7-dimethoxyflavone, Luteolin-3',7-dimethyl Ether-5-β-D-glucoside, 5-Methoxy-7-β-D-glucopyranosyl-(-)-afzele chin, Rutin	[98-100]
Caprifoliaceae	Badongrendong (Lonicera henryi), Jinyinhua (Lonicera japonica).	Luteolin, Apigenin, Luteolin-O-hexosides, Luteolin-7-O-β-glucoside, Luteolin-O-rhamnosyl-hexoside, Kaempferol-O-rhamnosyl-hexoside	[101, 102]
Orchidaceae	Jinxianlian (Anoectochilus roxburghii)	Isorhamnetin-3-O-β-D-rutinoside, Isorhamnetin-3-O-β-D-glucopyranoside	[103]
Violaceae	Zihuadiding (Viola philippica), Viola prionantha, Sansejin (Viola tricolor)	Delphinidin-3-O-rutinoside, Cyanidin-3-O-rutinoside, Delphinidin-3-O-rhamnosylglucoside-5-O-Glucoside	[104]
Saxifragaceae	Ganhuangcao (Penthorum chinense), Luojingjinyao (Chrysosplenium nudicaule)	Pinusin-7-O-β-D-glucopyranoside, Pinocin, 5-Methoxyxypin-7-O-β-D-glucoside, Kaempferol, Quercetin, Quercetin-3-O-β-D-xyloside, Apigenin, Luteolin, Catechin, 6,7,3'-Trimethoxy-3,5,4'-trihydroxyflavone, 5,4'-Dihydroxy-3,6,3'-trimethoxyflavone-7-O-β-D-glucoside	[105, 106]



Malvaceae	Qingma (Abutilon theophrasti)	Quercetin-7-O-β-glucoside, Rutin,	[107]
		Kaempferol-3-O-α-rhamnosyl	
		$(1\rightarrow 6)$ -β-D-glucopyranoside, Luteolin,	
		Apigenin -7-O-β-diglucoside	
Anacardiaceae	Renmianzi (Dracontomelon	Luteolin, L-Epicatechin, Quercetin,	[108]
	duperreanum)	Catechin	
Celastraceae	Leigongteng (Tripterygium wilfordii)	(\pm) -5, 4'-Dihydroxy-2'-methoxy-6',	[109]
		6'-dimethyl-(2', 3', 7, 8)-6-methyl flavanone	

Pharmacological Activity of Flavonoids

Anti-inflammatory activity

Inflammation is not only a defensive protective response but also a stimulus to various injury factors. It is dedicated to the defense and elimination of pathogenic factors in vivo and in vitro. However, persistent and intense inflammation can cause damage to human tissues and cells. Macrophages are important immunoregulatory cells that secrete different inflammatory factors, stimulate inflammatory responses, clear pathogens, and maintain the balance of the internal environment. LPS stimulates the RAW monocyte/macrophage-like 264.7 cell commonly used inflammatory model, and many flavonoids provide good resistance to this stimulation. It was reported that luteolin blocked the activation of NF-κB and inhibited the expression of NO, PGE2 (prostaglandin E2), inducible nitric oxide synthase (iNOS), cyclooxygenase (COX)2, TNF- α , and interleukin (IL)-6 in a dose-dependent manner under LPS stimulation in a RAW264.7 cell inflammation model [87]. Alpinetin can block the phosphorylation of IκBa protein, p65, p38, and ERK (extracellular signal-regulated kinase) and significantly inhibit the production of TNF-α, IL-6, and IL-1β [96]. Baicalein was shown to upregulate the expression of estrogen receptor ERα/ERβ in an LPS-induced RAW264.7 cell inflammation model, downregulate TNF-α, iNOS, and COX2 mRNA, inhibit NO and cytokine production in cells, and ultimately regulate the NF-kB pathway and activity to inhibit LPS-induced estrogen-like inflammatory cytokine production, thereby preventing inflammation-related diseases [110]. It can also play an anticomplement role through classical and alternative approaches [64]. Total flavonoids of Sangshen (Fructus Mori) can inhibit the expression of IL-6, induce iNOS, phosphorylate p65 and IkB, increase the expression of IL-10, and exert anti-inflammatory and analgesic effects by inhibiting the pro-inflammatory cytokines iNOS and NF-κB [18]. Also acting on the classical NF-κB inflammatory signaling pathway, the total flavonoids of Qingma (Abutilon theophrasti) can reduce the content of COX2, NO, IL-1β, IL-6, and TNF-α, increase the concentration of IL-10, inhibit the mRNA expression and phosphorylation of p65, and regulate the expression of inflammation-related factors Submit a manuscript: https://www.tmrjournals.com/tmr

through the NF- κ B and MAPK (mitogen-activated protein kinase) signaling pathways, in turn affecting the process of inflammation [107].

Acacetin inhibits 5-lipoxygenase activity in a concentration-dependent manner, which exerts an anti-inflammatory effect through inhibiting the activity of 5-lipoxygenase and preventing the production of leukotrienes, and it also inhibits the biosynthesis of TNF- α and NO [6]. In addition, some scholars have found in vivo anti-inflammatory experiments that luteolin and the ethyl acetate fraction of Suanjiang (Physalis alkekengi) can significantly alleviate edema in a carrageenan-induced acute rat inflammation model [87]. In the chronic rat inflammatory model, luteolin was also found to inhibit cotton ball-induced granuloma formation at 110 mg/kg and 50 mg/kg, and its anti-inflammatory effect was comparable to that of 50 mg/kg indomethacin [87]. Compared with 2 mg/kg aspirin, 5 mg/kg and 10 mg/kg of the ethanol extract of Youbingshiwei (Pyrrosia petiolosa) were shown to exert significant anti-inflammatory activity against xylene-induced ear swelling in mice, and the maximum inhibition rate was as high as 67% [10]. Apigenin, quercetin, kaempferol, and their analogues have similar anti-inflammatory activities [87].

Antibacterial activity

Since Fleming discovered penicillin, antibiotics have been used extensively in the clinic to reduce the prevalence and spread of a large number of diseases, but the struggle between humans and infectious diseases is endless. Despite the tremendous progress we have made, the emergence of various drug-resistant germs due to the abuse of antibiotics has made it difficult to treat certain infectious diseases. Faced with such a dilemma, researchers have turned their attention to the natural chemical ingredient flavonoids. Studies have found that the flavonoids from Oncoba spinosa can bind to soluble proteins outside the bacteria, destroying the stability of the microbial membrane [85]. Baicalin increases alkaline phosphatase activity and attaches to and penetrates the cell membrane of bacteria, causing the surface of the membrane to sag, changing the permeability, and increasing susceptibility to various antibiotics [111]. Another study reported that baicalein could penetrate bacterial biofilm, promote the hydrolysis of the quorum sensing regulatory protein TraR protein, interfere with cell signaling, and inhibit bacterial quorum sensing [112].

In addition, baicalein can inhibit bacterial virulence by inhibiting Salmonella typhimurium invasion into host cells by covalently binding to the protein substrates SipA/B/C/D and SopB of Salmonella typhimurium SPI-1 T3SS [60]. It has been shown that the flavonoids from Paotongguo (Paulownia fortune) can activate the pathway of Leishmania apoptotic methicillin-resistant Staphylococcus aureus and have synergistic effects with oxacillin and tetracycline [66]. Some scholars have reported that luteolin can inhibit the activity of enoyl-ACP reductase and inhibit bacterial fatty acid synthesis in a non-competitive manner and then exert its antibacterial effect [108, 113]. Interestingly, 1 mg/mL quercetin reduced the F-ATPase activity of Streptococcus mutans by 47.37% and 0.5 mg/mL kaempferol reduced it by 49.66%; they significantly inhibited the production of acid by Streptococcus mutans, suggesting that quercetin and kaempferol may have potential for the prevention and treatment of dental caries [114]. Apigenin alone or in combination with LysGH15, a lysin from phage GH15, which exhibits a highly efficient and extensive cleavage profile for MRSA (methicillin-resistant Staphylococcus aureus), was found to reduce rabbit erythrocyte lysis, protect lung tissue from mice with S. aureus pneumonia, and reduce the number of the bacteria in the lungs and blood, and TNF-α, IL-1β, and IL-6 levels were very similar to those in healthy mice [115]. Other studies have found that flavonoids have antimicrobial activity, but the mechanisms of their antimicrobial activity have not been thoroughly studied. For example, the total flavonoids of Sangshen (Fructus Mori) [18], the total flavonoids of Qingma (Abutilon theophrasti) [107], catechin [108], quercetin [108, 114], L-epicatechin [108], the ethyl acetate fraction of the ethanol extract of Leigongteng (Tripterygium wilfordii) [109], and kaempferol [114] were all found to have inhibitory effects on many strains of Escherichia coli, Cryptococcus neoformans, Pseudomonas aeruginosa, Staphylococcus Streptococcus aureus, mutans, Streptococcus sobrinus, Streptococcus Actinomyces naeslundii and Lactobacillus rhamnosus (Table 2).

Antiviral effects

In one study, 120-360 mg/kg of the total flavonoids from Shiqizhu (*Mosla scabra*) was used to treat influenza A virus-infected mice for 5 days, and the results indicated that the total flavonoids from Shiqizhu (*Mosla scabra*) upregulated the expression of *Toll-like receptor (TLR)-7*, retinoic acid-inducible gene (RIG)-1, TRAF6 (TNF receptor associated factor 6), BCL-2, BAX, VIPR1 (Vasoactive intestinal polypeptide receptor 1), PKCα (Protein kinase C alpha), and Aquaporin (AQP)-5 mRNAs and downregulated the expression of Caspase-3, NF-κB, and p65 proteins. In addition, the total flavonoids from Shiqizhu (*Mosla scabra*) reduced the levels of IL-6, TNF-α, and IL-1β

in serum and increased the expression of IFN-α. These results clearly demonstrate that the total flavonoids from Dendrobium can significantly alleviate lung inflammation, apoptosis, and water transport abnormalities induced by influenza A virus, which may be achieved by regulating the TLR7, RIG-1, and AQP5 signaling pathways [116].

Immunomodulatory effects

The total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) were found to downregulate the expression of FASL and TNFR1 mRNAs, upregulate the level of BCL-2 mRNA, and decrease the apoptosis rate of T cells; moreover, the activities of caspase-8 and caspase-3 in T cells of corticosterone rats were significantly inhibited, and the number of T cells was maintained [117]. Another study found that the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) significantly enhanced phagocytosis bv monocyte-macrophage system in normal mice, increased the level of serum hemolysin antibody formation, and antagonized the inhibition of monocyte-macrophage phagocytosis cyclophosphamide, reducing serum hemolysin antibody levels and delayed-type hypersensitivity [30].

Antioxidant activity

There is a complete oxidation system in cells, including ROS/RNS (reactive nitrogen species), O2, HO, HO₂, H₂, NO, NO+, and N₂O₃. Endogenous products mainly originate from mitochondria, peroxisomes, NADPH oxidase, cytochrome P450, and the xanthine redox enzyme system, and exogenous products are mainly related to ultraviolet and ionizing radiation. Low levels of ROS provide a stable microenvironment for maintaining the normal function of various intracellular biological macromolecules. On the contrary, high levels of free radicals can destroy the redox environment in cells and cause cell damage and death, and a persistent imbalance will eventually lead to aging and related diseases. Many flavonoids are excellent free radical scavengers because they can be used as hydrogen or electron donors. HORAC (Hydroxyl radical antioxidant capacity), TEAC [6-hydroxy-2, 5,7,8-tetramethylchroman-2-carboxylic acid (Trolox)-equivalent antioxidant capacity], ABTS [2, 2-azinobis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt], FCR (Folin-Ciocalteu), ORAC radical absorbance capacity), (oxygen 1,1-diphenyl-2-picrylhydrazyl (DPPH), FRAP (ferric reducing antioxidant power), POV (peroxide value), and others are commonly used indicators to detect antioxidant activity, especially DPPH (Table 3). Compared with the commonly used antioxidants such as vitamin C and ascorbic acid, quercetin [85] in Oncoba spinosa and flavonoids [21] in the methanol extract of peony have the same antioxidant activity as



vitamin C. At the same concentration, the total flavonoids of Taojinniang (*Rhodomyrtus tomentosa*) [76] and Maobaiyang (*Populus tomentosa*) stamen [15] were shown to be better than ascorbic acid at scavenging DPPH. The levels of SOD and GSH-Px (glutathione peroxidase) increased and that of MDA (malonic dialdehyde) were found to decrease in the plasma of Kunming mice after administration, which indicated that flavonoids could promote the expression of antioxidant enzymes and protect the body from peroxidation [76].

Inhibit the proliferation of tumor cells. The first step in cancer research is to study antiproliferation, which forms the basis for subsequent research. Both plant total flavonoid extracts, such as total flavonoids from Shuli (*Rhamnus davurica*), total flavonoids extracted from Shiye (*Diospyros kaki*), and the N-butanol extract of Shiwei (*Pyrrosia petiolosa*), and monomeric compounds, such as 6,7,3'-trimethoxy-3,5,4'-trihydroxyflavone, genistein, and rutin, all have good inhibitory effects on the growth of various tumor cell lines. Their sources and corresponding IC₅₀ values are shown in Table 4.

Antitumor effect

Table 2 The MICs values of flavonoids

	Table 2 The MICs values of flavonoids	
Compounds	Plants/Sources	MICs
Derrubon-5-methyl ether	Maoguoyuteng (Derris eriocarpa)	25-100 μg/mL [118]
MeOH extract ^a	The leaves of Oncoba spinosa	256-204 8 μg/mL [85]
Quercetin	The leaves of Oncoba spinosa, Fengfang (Nidus	64-128 μg/ml [85], 1-4 mg/mL
	vespae)	[114]
Kaempferol	Fengfang (Nidus vespae)	1-2 mg/mL [114]
Apigenin-7-O-β-D-glucuronopyranoside	The leaves of Oncoba spinosa	64-256 μg/mL [85]
Quercetin 3-O-β-D-galactopyranoside	The leaves of Oncoba spinosa	32-256 μg/mL [85]
2'-Hydroxyerythrin A	Soya beans	10.6-16.5 μg/mL [119]
Isoerythrinin A	Soya beans	15.2-22.6 μg/mL [119]
3'-O-methyldiplacol	Paotongguo (Paulownia fortune)	2-8 μg/mL [66]
Mimulone	Paotongguo (Paulownia fortune)	2-8 μg/mL [66]
Icariin	Yinyanghuo (Epimedium)	0.05%-0.23% [30]
Ethanol extract ^a	Youbingshiwei (Pyrrosia petiolosa)	1.25-10 mg/mL [10]
Ethyl acetate fraction ^a	Youbingshiwei (Pyrrosia petiolosa)	2.5 mg/mL [10]
Hydromethanolic extract of rhizome ^a	Ximalayadahuang (Rehum emodi)	25-125 μg/mL [120]
Petroleum ether fraction ^a	Youbingshiwei (Pyrrosia	2.5 mg/mL [10]
	petiolosa)	
N-butanol fraction ^a	Youbingshiwei (Pyrrosia	1.25-5 mg/mL [10]
	petiolosa)	
Ethyl acetate fraction ^a	Suanjiang (Physalis alkekengi)	500 μg/ml [88]
Aqueous fraction ^a	Youbingshiwei (<i>Pyrrosia petiolosa</i>)	2.5-5 mg/mL [10]
Ethanol extract ^a	Dingxiangye (Folium syringae)	0.65 mg/mL [121]
Hexane extracts ^a	Guizhencao (Bidens pilosa)	1.25-20 mg/mL [122]
Methanol extracts ^a	Guizhencao (Bidens pilosa)	2.5-20 mg/mL [122]
Ethanol extract a	Tufuling (Smilax glabra)	50-200 μg/ml [79]
Total flavonoids ^a	Qingma (Abutilon theophrasti)	0.06-1.02 g/mL [107]
Methanol extract ^a	Qinjiao (Gentiana macrophylla)	60-240 μg/mL [36]
Baicalin	Huangqin (Scutellaria baicalensis)	4 mg/ml [111], 1-2 mg/mL [64]
Baicalein	Huangqin (Scutellaria baicalensis)	0.125-0.5 mg/mL [64]
Wogonin	Huangqin (Scutellaria baicalensis)	0.5-2 mg/mL [64]
Oroxylin A	Huangqin (Scutellaria baicalensis)	0.125 mg/mL [64]
Norwogonin	Huangqin (Scutellaria baicalensis)	0.0625-0.25 mg/mL [64]
Nanoparticle-encapsulated ^a	Huangqin (Scutellaria baicalensis)	12.5 μg/mL [61]
Genistein	Unspecified	378 μg/mL [113]
Gentamicin ^b		0.5-2 μg/mL [107]
Metronidazole ^b		8 μg/mL [88]
Levofloxacin ^b		1.25-2.5 μg/mL [119]
Streptomycin ^b		$15 \pm 0.8 \mu \text{g/mL} [120]$

Minimum inhibitory concentration (MICs); a mixture; b positive control.

Table 3 Determination of the antioxidant capacity of flavonoids

Comment	DI 4 m/C	DPPH
Compounds	Plants/Sources	IC ₅₀ Value(μg/mL)
Quercetin	Xifanlian (Passiflora loefgrenii)	2.56 [82]
Total flavonoids a	Qingma (Abutilon theophrasti)	8.96 [107], 8.41 [107], 14.41 [107]
Ethanol extract ^a	Xifanlian (Passiflora loefgrenii)	350 [82]
Ethanol extract ^a	Mudanhua (Paeonia suffruticosa)	32.6 [21]
Dihydrokaempferol	Mudanhua (Paeonia suffruticosa)	24.6 [21]
Apigenin-7-O-β-D-glucoside	Mudanhua (Paeonia suffruticosa)	34.2 [21]
2'-Hydroxyerythrin A	Soya beans	28.5 ± 2.2 [119]
Daidzein-7-O-β-D-glycoside	Soya beans	> 50.0 [119]
7,4'-Dihydroxy-6-methoxyisoflavone	Soya beans	24.2 ± 1.5 [119]
Daidzein	Soya beans	31.3 ± 2.0 [119]
Genistein	Soya beans	13.6 ± 0.8 [119]
Apigenin-7-O-β-D-neohesperidoside	Mudanhua (Paeonia suffruticosa)	40.1 [21]
Kaempferol-7-O-β-D-glucopyranoside	Mudanhua (Paeonia suffruticosa)	35.3 [21]
Kaempferol-3-O-β-D-glucopyranosyl-7-O-β	Mudanhua (Paeonia suffruticosa)	20.9 [21]
-D-glucopyranoside		
Total flavonoids a	Yinyanghuo (Epimedium)	4.67 [30]
Trolox ^b		1.2 ± 0.1 [119]
2,6-Di-Tert-Butyl hydroxytoluene ^b		8.11 [107]

DPPH: 1,1-Diphenyl-2-picrylhydrazyl. ^a mixture; ^b positive control.

Table 4 IC₅₀ values of flavonoids that inhibit the proliferation of tumor cell lines

Cell lines	Compounds	Plants/Sources	IC ₅₀
SGC-7901	Total flavonoids ^a	Shuli (Rhamnus davurica)	89.53 ± 4.11 μg/mL [2]
	Cambodianal	Xuejie (Daemonorops draco)	5.0 μg/mL [80]
	Paclitaxel ^b		8.7 μg/mL [80]
	6,7,3'-Trimethoxy-3,5,4'-trihydroxy	Luojingjinyao (Chrysosplenium	8.33 μg/mL [106]
	flavone	nudicaule)	
HT-29	Total flavonoids a	Shuli (Rhamnus davurica)	$24.96 \pm 0.74 \ \mu g/mL \ [2]$
PC-3	Total flavonoids extracted	Shiye (Diospyros kakia)	$42.7 \pm 6.1 \mu \text{g/mL} [83]$
	Quercetin ^b		$48.0 \pm 5.4 \mu \text{g/mL}$ [83]
	Rutin ^b		$49.2 \pm 8.2 \mu \text{g/mL} [83]$
	Docetaxel b		$0.041 \pm 0.005 \mu \text{g/mL}$ [83]
K562	Cambodianal	Xuejie (Daemonorops draco)	1.5 μg/mL [80]
	Paclitaxel ^b		7.3 μg/mL [80]
SMMC-7721	Cambodianal	Xuejie (Daemonorops draco)	2.8 μg/mL [80]
	Paclitaxel ^b		2.1 μg/mL [80]
A549	N-Butanol extract ^a	Shiwei (Pyrrosia petiolosa)	0.33 mg/mL [10]
SK-Hep-1	Genistein	Jixueteng (Millettia reticulata)	16.23 μmol/L [48]
HepG2	Bavachin	Buguzhi (Psoralea corylifolia)	24.4 μmol/L [46]
L-O2	Total flavonoids ^a	Shuli (Rhamnus davurica)	$229.19 \pm 8.52 \mu \text{g/mL}}$ [2]

Promote apoptosis of tumor cells. Apoptosis, known as programmed cell death, is an autonomous and orderly procedure for death controlled by genes. Apoptosis not only participates in the regulation of the number of cells in the body but also can remove cells in the body that are not functional, cells that are harmful to the body, mutated cells, and cells that cannot survive after being damaged. Moreover, apoptosis plays an important role in the development and homeostasis of the body. Therefore, promoting the apoptosis of tumor cells can effectively inhibit the occurrence and development of tumors. There are three main apoptosis-related signal transduction pathways

(Figure 2).

Mitochondria-mediated signal transduction pathway. Some scholars have found that the total flavonoids extracted from Shiye (Diospyros kaki) [83], 7-hydroxy-2',2'-dimethyl-2'H,4H-3,6'-bichromen-4-one [46], Baicalein [123], quercetin, and resveratrol [124] can reduce mitochondrial membrane potential, change the mitochondrial morphology, induce the production of large amounts of reactive oxygen species (ROS), reduce the ratio of B-cell lymphoma-2 (Bcl-2)/ Bcl-2 associated X protein (BAX), promote the release of cytochrome C from mitochondria, and then induce the activation of caspase-9 and caspase-3, followed by

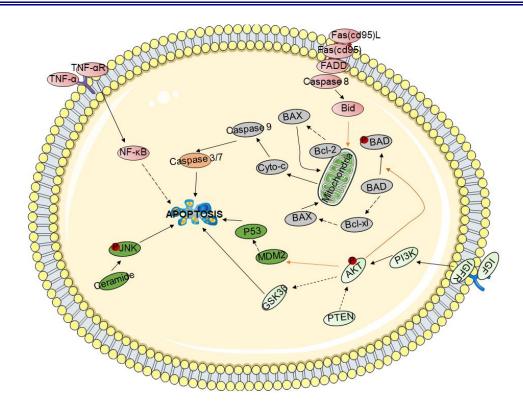


Figure 2 Schematic representation of the cell apoptosis signaling pathway

Shown are the extrinsic (receptor-mediated, red) and intrinsic (mitochondria-driven, gray) apoptosis pathways, which act in opposition to the survival proteins such as the PI3K/Akt signaling circuitry (light green). Other signaling loops (dark green) and executioner caspases (orange) are activated in both the extrinsic and intrinsic pathways; inter-talk between pathways (arrows) and the activation and suppression effects are indicated by solid arrows and dashed arrows, respectively.

cleavage of poly (ADP-ribose) polymerase (PARP), leads to DNA fragmentation and ultimately to tumor cell apoptosis. In addition, studies have reported that the combination of quercetin and resveratrol can affect the expression of mammalian target of rapamycin (mTOR), p-protein kinase B (Akt), p-mTOR, NF-κB, matrix metalloproteinase-2 (MMP-2), matrix metalloproteinase-2 (MMP-9), and TIMP-2 (tissue inhibitor of metalloproteinases-2) proteins, which may further enhance the antitumor activity of the drug by targeting mitochondria, Akt/mTOR, and NF-κB signaling pathways [124].

Endoplasmic reticulum-mediated signal transduction pathway. It has been shown in the literature that the treatment of HepG2 cells with 20 µmol/L Bavachin can activate activating transcription factor (ATF)4, transcription factors the CHOP(C/EBP-homologous protein) X-box binding protein1 (XBP1), and protein kinase R-like ER kinase (PERK)-ATF4, inositol-requiring enzyme1(IRE1)-XBP1s, and ATF6. Three pathways induce endoplasmic reticulum stress. Bavachin induces endoplasmic reticulum stress by inducing PERK-ATF4, IRE1-XBP1s, and ATF6; moreover, the depletion of Mfn2 (mitochondrial fusion 2) aggravates ER stress by phosphorylating Akt, activating apoptosis-responsive

proteins such as death receptors, and inducing tumor cell apoptosis [46].

Death factor and its receptor-mediated signal transduction pathway. Studies in the literature have shown that genistein can upregulate the expression of Fas, FasL, and p53 proteins in SK-Hep-1 cells, subsequently induced the activation of caspase-9 and caspase-3, which was followed by cleavage of PARP, leading to DNA fragmentation and ultimately to apoptosis [48]. Baicalin can block cells in G0/G1 phase; decrease mitochondrial membrane potential; activate caspase-3, caspase-8, and caspase-9; initiate FasL/Fas expression; and induce apoptosis significantly through internal and external sources. Moreover, the proto-oncogene C-MYC and its target gene, hTERT (human telomerase reverse transcriptase) are downregulated to inhibit telomerase activity, ultimately inhibiting the proliferation of tumor HL-60 cells [125]. Some scholars have found through miRNA microarray analysis that baicalin downregulates many oncomiRs such as miR-10a, miR-23a, miR-30c, miR-31, miR-151a, and miR-205 in vitro and in vivo, inhibiting C-MYC expression. It effectively promotes apoptosis in the HT-29 colon cancer cell line in doseand time-dependent manners and inhibits tumor growth in corresponding transplanted nude mice [126].



Antihypertensive effect

In one study, 26 mg/kg Epimedium flavonoids was injected into the lateral ventricle of mice, and epimedium flavonoids were found to increase the secretion of the amino acid neurotransmitter r-aminobutyric acid in the periventricular system and increase its affinity for receptors; strengthen the inhibition of the central sympathetic nervous system; expand coronary blood vessels, the femoral artery, and the cerebrovascular system; and reduce peripheral resistance, thus lowering blood pressure [30].

Reduce obesity

The occurrence and development of various diseases are related to obesity, and in recent years, the incidence of diabetes, hyperlipidemia, hypertension, and endocrine disorders in obese patients has increased significantly. Obesity has become an important cause of cardiovascular and cerebrovascular diseases, hypertension, and diabetes. Some scholars found that the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) increased the appetite and exercise volume of rats, but the weight of the rats was lower than that of untreated rats after 24 months. Moreover, the concentration of age-related metabolites returned to a level seen in younger animals; the amount of unsaturated fatty acids decreased; saturated fatty acids, deoxycholic acid, triglycerides, and total cholesterol increased; and their fur was smoother and more lustrous. These results suggest that the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) may have potential body weight controlling and anti-aging effects [117].

Role in liver protection

Dihydromyricetin can significantly improve the abnormal expression of cytochrome P450 2E1, Kelch-like ECH-binding protein 1, and heme oxygenase-1 in the liver of mice with alcoholic liver disease, improve the disordered nuclear localization of NF-κB and nuclear factor erythroid-2-related factor 2 (Nrf2), and play a role in liver protection. Further exploration of the mechanism revealed that dihydromyricetin may mediate the activation of Nrf2 via the autophagy pathway, upregulate p62 positive feedback Nrf2 activation, reduce inflammation in liver steatosis and the pathological progression of alcoholic liver disease, and partially restore liver pathological changes [127].

Neuroprotection

Some researchers have found that in mice, isobavachalcone can inhibit the excessive activation of microglia, reduce the expression of IL-6 and IL-1β in the brain of Parkinson's disease mice, prolong the residence time of mice on a Rota-rod, reduce neuronal necrosis, and effectively alleviate Parkinson's disease Submit a manuscript: https://www.tmrjournals.com/tmr

induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *In vitro* experiments showed that isobavachalcone inhibited NF-κB by blocking LPS-induced NF-κB subunit transfer from mouse microglia BV-2 cytoplasm to the nucleus, reduced the LPS-induced oxidative stress response to inflammatory cytokine expression, and protected nerves through antagonism [44].

Other effects

In one study, the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) (150 mg/kg) were administered to young rats by intraperitoneal injection for 7 days. It was found that the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) significantly increased the weight of the anterior pituitary, epididymis, and seminal vesicles in young rats; increased testosterone, estradiol, and luteinizing hormone levels; and promoted the secretion of testosterone by rat stromal cells [30]. In addition to promoting the male reproductive system and reproductive endocrine activity, the total flavonoids extracted from Yinyanghuo (Epimedium brevicornu Maxim) were shown to promote the proliferation of primary osteoblasts and osteoblast-like MAGR106 cells, enhance the extent of mineralized tuberculosis in vitro, and enhance the mRNA expression of BMP-2(human Bone morphogenetic protein 2), BMP-4 (human Bone morphogenetic protein 4), RUNX2 (runt-related transcription factor 2), β -catenin, and cyclin D1, as well as promote the osteogenic differentiation of human bone marrow mesenchymal stem cells [30]. Icariin can significantly increase the expression of eNOS (endothelial nitric oxide synthase) and cGMP in corpus cavernosum smooth muscle, inducing the relaxation of corpus cavernosum smooth muscle and penile erection [28]. Another study reported that baicalin increased the concentration of cAMP and AFC (alveolar fluid clearance) in a dose-dependent manner in rats with acute lung injury, and baicalin prevented the reduction of AFC by upregulating α-ENaC (α-epithelial Na+ channel) protein expression, which was activated by stimulating cAMP/PKA (cyclic adenosine monophosphate/protein kinase A) signaling pathway [128].

Prospective

The structure of flavonoids in plants is diverse, and many plant families contain them. The research topics mentioned in this review include flavonoid monomer components, different polar extraction sites, and total flavonoids. Flavonoids have a variety of pharmacological effects, including anti-oxidation, antitumor, weight loss, antibacterial, antihypertensive, liver protective, anti-inflammatory, and



neuroprotective activities. In short, plant-derived flavonoids are widely used as a bulk component, and whether the monomer is administered alone, monomers are administered in combination, or the total extract is administered directly, good effects have been obtained. However, the research on the mechanisms of the various activities of flavonoids is still in its infancy. Therefore, further studies are needed on the mechanisms by which flavonoids affect diseases and their long-term effects. Such studies will provide a sound scientific basis for the development and application of natural medicines, bringing relief to patients with various diseases.

References

- 1. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci 2016, 5: e47.
- Chen G, Li X, Saleri F, et al. Analysis of Flavonoids in Rhamnus davurica and Its Antiproliferative Activities. Molecules 2016, 21: 1275.
- Gonzales GB. In vitro bioavailability and cellular bioactivity studies of flavonoids and flavonoid-rich plant extracts: questions, considerations and future perspectives. Proc Nutr Soc 2017, 76: 175-181.
- 4. Ma YF, Shang FD. Distribution of flavonoid in medicinal plants. J Biol 2003, 20: 35-39.
- Zou H, Xu KP, Li FS, et al. Unciflavones A-F, six novel flavonoids from Selaginella uncinata (Desv.) Spring. Fitoterapia 2014, 99: 328-333.
- Fan SY, Zeng HW, Pei YH, et al. The anti-inflammatory activities of an extract and compounds isolated from Platycladus orientalis (Linnaeus) Franco in vitro and ex vivo. J Ethnopharmacol 2012, 141: 647-652.
- Ogo Y, Mori T, Nakabayashi R, et al. Transgenic rice seed expressing flavonoid biosynthetic genes accumulate glycosylated and/or acylated flavonoids in protein bodies. J Exp Bot. 2016, 67: 95-106.
- 8. Uddin SJ, Grice D, Tiralongo E. Evaluation of cytotoxic activity of patriscabratine, tetracosane and various flavonoids isolated from the Bangladeshi medicinal plant Acrostichum aureum. Pharm Biol 2012, 50: 1276-1280.
- 9. Wang N, Wang JH, Li X, *et al.* Flavonoids from Pyrrosia petiolosa (Christ) Ching. J Asian Nat Prod Res 2006, 8: 753-756.
- Cheng D, Zhang Y, Gao D, et al. Antibacterial and anti-inflammatory activities of extract and fractions from Pyrrosia petiolosa (Christ et Bar.) Ching. J Ethnopharmacol 2014, 155: 1300-1305.
- 11. Wang T, Xiao J, Hou H, *et al.* Development of an ultra-fast liquid chromatography-tandem mass spectrometry method for simultaneous determination of seven flavonoids in rat plasma:

- Application to a comparative pharmacokinetic investigation of Ginkgo biloba extract and single pure ginkgo flavonoids after oral administration. J Chromatogr B Analyt Technol Biomed Life Sci 2017, 1060: 173-181.
- 12. Chen S, Xing XH, Huang JJ, et al. Enzyme-assisted extraction of flavonoids from Ginkgo biloba leaves: improvement effect of flavonol transglycosylation catalyzed by Penicillium decumbens cellulase. Enzyme Microb Technol 2011, 48: 100-105.
- 13. Yu XA, Azietaku JT, Li J, *et al.* Simultaneous determination of eight flavonoids in plasma using LC-MS/MS and application to a pharmacokinetic study after oral administration of Pollen Typhae extract to rats. J Chromatogr B Analyt Technol Biomed Life Sci 2017, 1044: 158-165.
- 14. Al-Rimawi F, Abu-Lafi S, Abbadi J, *et al.* Analysis of phenolic and flavonoids of wild Ephedra alata plant extracts by LC/PDA and LC/MS and their antioxidant activity. Afr J Tradit Complement Altern Med 2017, 14: 130-141.
- 15. Wan P, Sheng Z, Han Q, et al. Enrichment and purification of total flavonoids from Flos Populi extracts with macroporous resins and evaluation of antioxidant activities in vitro. J Chromatogr B Analyt Technol Biomed Life Sci 2014, 945: 68-74.
- 16. Tao XY, Zhang DW, Chen RD, *et al.* Chemical constituents from cell cultures of Morus alba. Chin J Chin Mater Med 2012, 37: 3738-3742.
- 17. Ti H, Wu P, Lin L, *et al*. Stilbenes and flavonoids from Artocarpus nitidus subsp. lingnanensis. Fitoterapia 2011, 82: 662-665.
- 18. Chen H, Yu W, Chen G, *et al*. Antinociceptive and Antibacterial Properties of Anthocyanins and Flavonols from Fruits of Black and Non-Black Mulberries. Molecules 2017, 23: 4.
- 19. Zhang CF, Chen J, Zhao LQ, *et al.* Three new flavonoids from the active extract of Fallopia convolvulus. J Asian Nat Prod Res 2011, 13: 136-142.
- 20. Sait S, Hamri-Zeghichi S, Boulekbache-Makhlouf L, *et al.* HPLC-UV/DAD and ESI-MS(n) analysis of flavonoids and antioxidant activity of an Algerian medicinal plant: Paronychia argentea Lam. J Pharm Biomed Anal 2015, 111: 231-240.
- 21. Zhang H, Li X, Wu K, et al. Antioxidant Activities and Chemical Constituents of Flavonoids from the Flower of Paeonia ostii. Molecules 2017, 22: 5.
- 22. He C, Peng B, Dan Y, *et al.* Chemical taxonomy of tree peony species from China based on root cortex metabolic fingerprinting. Phytochemistry 2014, 107: 69-79.
- 23. Xu L, Zhang X, Lin LM, *et al*. Two new flavonol glycosides from the Tibetan medicinal plant Aconitum tanguticum. J Asian Nat Prod Res 2013,

15: 737-742.

- 24. Yu Y, Yi ZB, Liang YZ. Validate antibacterial mode and find main bioactive components of traditional Chinese medicine Aquilegia oxysepala. Bioorg Med Chem Lett 2007, 17: 1855-1859.
- 25. Wang R, Li YX, Quan QM. The contents changes of polysaccharides and total flavonoids during flower bud differentiation of Epimedium sagittatum. J Chin Med Master 2009, 32: 1511-1514.
- 26. Chen J, Xu Y, Wei G, *et al.* Chemotypic and genetic diversity in Epimedium sagittatum from different geographical regions of China. Phytochemistry 2015, 116: 180-187.
- 27. Li WK, Zhang RY, Xiao PG. Flavonoids from Epimedium wanshanense. Phytochemistry 1996, 43: 527-530.
- 28. Li C, Li Q, Mei Q, *et al.* Pharmacological effects and pharmacokinetic properties of icariin, the major bioactive component in Herba Epimedii. Life Sci 2015, 126: 57-68.
- 29. Peng YD, Huang WH, Guo BL. Research on quality of Epimedium extract in market. Chin J Chin Mater Med 2007, 32: 1858-1861.
- 30. Ma H, He X, Yang Y, *et al*. The genus Epimedium: an ethnopharmacological and phytochemical review. J Ethnopharmacol 2011, 134: 519-541.
- 31. Vidovic M, Morina F, Milic S, *et al.* Carbon allocation from source to sink leaf tissue in relation to flavonoid biosynthesis in variegated Pelargonium zonale under UV-B radiation and high PAR intensity. Plant Physiol Biochem 2015, 93: 44-55.
- Wang L, Mei Q, Wan D. Simultaneous determination by HPLC of quercetin and kaempferol in three Sedum medicinal plants harvested in different seasons. J Chromatogr Sci 2014, 52: 334-338.
- 33. Zhang S, Liu C, Bi H, *et al.* Extraction of flavonoids from Rhodiola sachlinesis A. Bor by UPE and the antioxidant activity of its extract. Natur Prod Res 2008, 22: 178-187.
- 34. Chou TH, Chen JJ, Lee SJ, *et al.* Cytotoxic flavonoids from the leaves of Cryptocarya chinensis. J Natur Prod 2010, 73: 1470-1475.
- 35. Olennikov DN, Kashchenko NI, Chirikova NK, *et al.* Iridoids and Flavonoids of Four Siberian Gentians: Chemical Profile and Gastric Stimulatory Effect. Molecules 2015, 20: 19172-19188.
- 36. Yin C, Xie L, Guo Y. Phytochem Anal and antibacterial activity of Gentiana macrophylla extract against bacteria isolated from burn wound infections. Microb Pathog 2018, 114: 25-28.
- 37. Wu SK, Zhang N, Shen XR, et al. Preparation of total flavonoids from loquat flower and its protective effect on acute alcohol-induced liver injury in mice. J Food Drug Anal 2015, 23:

136-143.

- 38. Thongnest S, Lhinhatrakool T, Wetprasit N, *et al.* Eriosema chinense: a rich source of antimicrobial and antioxidant flavonoids. Phytochemistry 2013, 96: 353-359
- 39. Huang M, Wang W, Wei S. Investigation on medicinal plant resources of Glycyrrhiza uralensis in China and chemical assessment of its underground part. Chin J Chin Mater Med 2010, 35: 947-952.
- 40. Lou H, Yuan H, Yamazaki Y, *et al.* Alkaloids and flavonoids from peanut skins. Planta medica 2001, 67: 345-349.
- 41. Song S, Zheng X, Liu W, *et al.* 3-Hydroxymethylglutaryl flavonol glycosides from a Mongolian and Tibetan medicine, Oxytropis racemosa. Chem Pharm Bull 2010, 58: 1587-1590.
- 42. Woo ER, Kwak JH, Kim HJ, *et al.* A new prenylated flavonol from the roots of Sophora flavescens. J Natur Prod 1998, 61: 1552-1554.
- 43. Tan Q, Zhang S, Shen Z. Flavonoids from the roots of Campylotropis hirtella. Planta medica 2011, 77: 1811-1817.
- 44. Jing H, Wang S, Wang M, *et al.* Isobavachalcone Attenuates MPTP-Induced Parkinson's Disease in Mice by Inhibition of Microglial Activation through NF-kappaB Pathway. PLoS One 2017, 12: e0169560.
- 45. Jiang H, Hu JR, Zhan WQ, *et al.* Screening for fractions of Oxytropis falcata Bunge with antibacterial activity. Natur Prod Res 2009, 23: 953-959.
- 46. Yang Y, Tang X, Hao F, *et al.* Bavachin Induces Apoptosis through Mitochondrial Regulated ER Stress Pathway in HepG2 Cells. Biol Pharm Bull 2018, 41: 198-207.
- 47. Zhang L, Qiang P, Yu J, et al. Identification of compound CA-5f as a novel late-stage autophagy inhibitor with potent anti-tumor effect against non-small cell lung cancer. Autophagy 2018: 1-16.
- 48. Fang SC, Hsu CL, Lin HT, *et al.* Anticancer effects of flavonoid derivatives isolated from Millettia reticulata Benth in SK-Hep-1 human hepatocellular carcinoma cells. J Agric Food Chem 2010, 58: 814-820.
- Ma L, Wu F, Chen RY. Advance of chemical constituents and bioactivity of Saururuaceae plants. Chin J Chin Mater Med 2003, 28: 196-198.
- 50. Chen SD, Li T, Gao H, *et al.* Anti HSV-1 flavonoid derivatives tethered with houttuynin from Houttuynia cordata. Planta medica 2013, 79: 1742-1748.
- 51. Zheng CJ, Li HQ, Ren SC, *et al.* Phytochemical and Pharmacological Profile of Vitex negundo. Phytother Res 2015, 29: 633-647.

- 52. Liu X, Luo Y, Wu H, *et al.* Systematic analysis of O-methyltransferase gene family and identification of potential members involved in the formation of O-methylated flavonoids in Citrus. Gene 2016, 575: 458-472.
- 53. Adaszynska-Skwirzynska M, Dzieciol M. Comparison of phenolic acids and flavonoids contents in various cultivars and parts of common lavender (Lavandula angustifolia) derived from Poland. Natur Prod Res 2017, 31: 2575-2580.
- 54. Yu C, Qu F, Mao Y, *et al.* Different extraction pretreatments significantly change the flavonoid contents of Scutellaria baicalensis. Pharm Biol 2013, 51: 1228-1235.
- 55. Huang ZH, Jiang DX, Lai XP. Study on the enrichment and purification of total flavonoids in Schizonepeta tenuifolia by macroporous adsorption resin. J Chin Med Master 2010, 33: 1476-1480.
- 56. Gao Z, Huang K, Yang X, *et al.* Free radical scavenging and antioxidant activities of flavonoids extracted from the radix of Scutellaria baicalensis Georgi. Biochimica et biophysica acta 1999, 1472: 643-650.
- 57. Hu HW, Xie XM, Zhang PZ, *et al.* Study on the flavonoids from Mosla chinensis "jiangxiangru". J Chin Med Master 2010, 33: 218-219.
- 58. Broncel M. Antiatherosclerotic properties of flavones from the roots of Scutellaria baicalensis Georgi. Wiad Lek 2007, 60: 294-297.
- 59. Guan Z, Li S, Lin Z, et al. Identification and quantitation of phenolic compounds from the seed and pomace of Perilla frutescens using HPLC/PDA and HPLC-ESI/QTOF/MS/MS. Phytochem Anal 2014, 25: 508-513.
- 60. Tsou LK, Lara-Tejero M, RoseFigura J, et al. Antibacterial Flavonoids from Medicinal Plants Covalently Inactivate Type III Protein Secretion Substrates. J Am Chem Soc 2016, 138: 2209-2218.
- 61. Leung KC, Seneviratne CJ, Li X, et al. Synergistic Antibacterial Effects of Nanoparticles Encapsulated with Scutellaria baicalensis and Pure Chlorhexidine on Oral Bacterial Biofilms. Nanomaterials 2016, 138: 2209-2218.
- 62. Zhu L, Zhao L, Wang H, *et al.* Oroxylin A reverses P-glycoprotein-mediated multidrug resistance of MCF7/ADR cells by G2/M arrest. Toxicol Lett 2013, 219: 107-115.
- 63. Bai Y, Xia B, Xie W, *et al.* Phytochemistry and pharmacological activities of the genus Prunella. Food Chem 2016, 204: 483-496.
- 64. Xing S, Wang M, Peng Y, et al. Simulated gastrointestinal tract metabolism and pharmacological activities of water extract of Scutellaria baicalensis roots. J Ethnopharmacol 2014, 152: 183-189.
- 65. Shi QQ, Dang J, Wen HX, *et al*. Anti-hepatitis, Submit a manuscript: https://www.tmrjournals.com/tmr

- antioxidant activities and bioactive compounds of Dracocephalum heterophyllum extracts. Bot Stud 2016, 57: 16.
- 66. Navratilova A, Nesuta O, Vancatova I, *et al.* C-Geranylated flavonoids from Paulownia tomentosa fruits with antimicrobial potential and synergistic activity with antibiotics. Pharm Biol 2016, 54: 1398-1407.
- 67. Li MX, He XR, Tao R, *et al.* Phytochemistry and pharmacology of the genus pedicularis used in traditional Chinese medicine. Am J Chin Med 2014, 42: 1071-1098.
- 68. Borgognone D, Cardarelli M, Rea E, *et al.* Salinity source-induced changes in yield, mineral composition, phenolic acids and flavonoids in leaves of artichoke and cardoon grown in floating system. J Sci Food Agric 2014, 94: 1231-1237.
- 69. Pereira C, Barros L, Carvalho AM, *et al.* Infusions of artichoke and milk thistle represent a good source of phenolic acids and flavonoids. Food Funct 2015, 6: 56-62.
- 70. Qiu J, Gao F, Shen G, *et al.* Metabolic engineering of the phenylpropanoid pathway enhances the antioxidant capacity of Saussurea involucrata. PLoS One 2013, 8: e70665.
- 71. Liu QM, Zhao HY, Zhong XK, *et al.* Eclipta prostrata L. phytochemicals: isolation, structure elucidation, and their antitumor activity. Food Chem Toxicol 2012, 50: 4016-4022.
- 72. Yang GE, Bao L, Zhang XQ, et al. Studies on flavonoids and their antioxidant activities of Artemisia annua. J Chin Med Master 2009, 32: 1683-1686.
- 73. Zhou CX, Wu DY, Li XP, *et al.* Research progress in Laggera medicinal plants. Chin J Chin Mater Med 2006, 31: 1133-1140.
- 74. Xing XD, Zhang Q, Feng F, *et al*. Chemical constituents from stems of Ilex pubescens. J Chin Med Master 2012, 35: 1429-1431.
- 75. Goodger JQ, Seneratne SL, Nicolle D, *et al.* Foliar Essential Oil Glands of Eucalyptus Subgenus Eucalyptus (Myrtaceae) Are a Rich Source of Flavonoids and Related Non-Volatile Constituents. PLoS One 2016, 11: e0151432.
- 76. Wu P, Ma G, Li N, *et al.* Investigation of in vitro and in vivo antioxidant activities of flavonoids rich extract from the berries of Rhodomyrtus tomentosa(Ait.) Hassk. Food Chem 2015, 173: 194-202.
- 77. Perez-Gregorio MR, Regueiro J, Simal-Gandara J, *et al.* Increasing the added-value of onions as a source of antioxidant flavonoids: a critical review. Crit Rev Food Sci Nutr 2014, 54: 1050-1062.
- 78. Porta C, Paglino C, Mosca A. Targeting PI3K/Akt/mTOR Signaling in Cancer. Front Oncol 2014, 4: 64.
- 79. Hua S, Zhang Y, Liu J, et al. Ethnomedicine, Phytochemistry and Pharmacology of Smilax

- glabra: An Important Traditional Chinese Medicine. Am J Chin Med 2018, 46: 261-297.
- 80. Wang H, Jiang HM, Li FX, *et al.* Flavonoids from artificially induced dragon's blood of Dracaena cambodiana. Fitoterapia 2017, 121: 1-5.
- 81. Gao SM, Liu JS, Wang M, *et al.* Traditional uses, phytochemistry, pharmacology and toxicology of Codonopsis: A review. J Ethnopharmacol 2018, 219: 50-70.
- 82. Argentieri MP, Levi M, Guzzo F, *et al.* Phytochem Anal of Passiflora loefgrenii Vitta, a rich source of luteolin-derived flavonoids with antioxidant properties. J Pharm Pharmacol 2015, 67: 1603-1612.
- 83. Ding Y, Ren K, Dong H, *et al.* Flavonoids from persimmon (Diospyros kaki L.) leaves inhibit proliferation and induce apoptosis in PC-3 cells by activation of oxidative stress and mitochondrial apoptosis. Chem Biol Interact 2017, 275: 210-217.
- 84. Bei WJ, Xu AL, Li CY, *et al.* Flavonoids from Diospyros kaki inhibit the adhesion between lymphocyte and dorsal root ganglion. J Chin Med Master 2009, 32: 740-744.
- 85. Djouossi MG, Tamokou JD, Ngnokam D, *et al.* Antimicrobial and antioxidant flavonoids from the leaves of Oncoba spinosa Forssk. (Salicaceae). BMC Complement Altern Med 2015, 15: 134.
- 86. Zhou XL, Shi T, Yang L, *et al.* Research on the effect and mechanism of total ginkgo flavones-glycoides on hepatocyte apoptosis in rats with nonalcoholic fatty liver disease. Chin J Integ Trad West Med Dig 2017.
- 87. Li AL, Chen BJ, Li GH, et al. Physalis alkekengi L. var. franchetii (Mast.) Makino: An ethnomedical, phytochemical and pharmacological review. J Ethnopharmacol 2018, 210: 260-274.
- 88. Wang Y, Wang SL, Zhang JY, et al. Anti-ulcer and anti-Helicobacter pylori potentials of the ethyl acetate fraction of Physalis alkekengi L. var. franchetii (Solanaceae) in rodent. J Ethnopharmacol 2018, 211: 197-206.
- 89. Kurepa J, Nakabayashi R, Paunesku T, *et al.* Direct isolation of flavonoids from plants using ultra-small anatase TiO(2) nanoparticles. The Plant Journal 2014, 77: 443-453.
- 90. Luan N, Li D. Study on supercritical CO₂ extraction of flavonoids from Cynomorium songaricum. J Chin Med Master 2010, 33: 1167-1171.
- 91. Cui Z, Guo Z, Miao J, et al. The genus Cynomorium in China: an ethnopharmacological and phytochemical review. J Ethnopharmacol 2013, 147: 1-15.
- 92. Li JS, Zhao YY, Wang B, et al. Separation and identification of the flavonoids from Buddleia officinalis Maxim. Acta Pharmaceutica Sinica

- 1996, 31: 849-854.
- 93. Peng XJ, Li C. Study on the flavanone constitutes of Buddleja davidii. J Chin Med Master 2011, 34: 1534-1537.
- 94. Xu XH, Yang NY, Qian SH, *et al.* Stduy on flavonoids in Ligustrum lucidum. J Chin Med Master 2007, 30: 538-540.
- 95. Jia J, Zhang F, Li Z, *et al.* Comparison of Fruits of Forsythia suspensa at Two Different Maturation Stages by NMR-Based Metabolomics. Molecules 2015, 20: 10065-10081.
- 96. Qu J, Yan X, Li C, *et al.* Comparative Evaluation of Raw and Ripe Fruits of Forsythia suspensa by HPLC-ESI-MS/MS Analysis and Anti-Microbial Assay. J Chromatogr Sci 2017, 55: 451-458.
- 97. Ye M, Li Y, Yan Y, *et al.* Determination of flavonoids in Semen Cuscutae by RP-HPLC. J Pharm Biomed Anal 2002, 28: 621-628.
- 98. Sun Q, Wang D, Li FF, *et al.* Cytotoxic prenylated flavones from the stem and root bark of Daphne giraldii. Bioorg Med Chem Lett 2016, 26: 3968-3972.
- 99. Lu CL, Zhu L, Piao JH, *et al.* Chemical compositions extracted from Wikstroemia indica and their multiple activities. Pharm Biol 2012, 50: 225-231.
- 100. Pan L, Hu H, Wang X, et al. Inhibitory effects of neochamaejasmin B on P-glycoprotein in MDCK-hMDR1 cells and molecular docking of NCB binding in P-glycoprotein. Molecules 2015, 20: 2931-2948.
- 101. Jaiswal R, Muller H, Muller A, *et al.* Identification and characterization of chlorogenic acids, chlorogenic acid glycosides and flavonoids from Lonicera henryi L. (Caprifoliaceae) leaves by LC-MSn. Phytochemistry 2014, 108: 252-263.
- 102. Yuan Y, Song L, Li M, *et al.* Genetic variation and metabolic pathway intricacy govern the active compound content and quality of the Chinese medicinal plant Lonicera japonica thunb. BMC genomics 2012, 13: 195.
- 103. Zhang FS, Lv YL, Zhao Y, *et al.* Promoting role of an endophyte on the growth and contents of kinsenosides and flavonoids of Anoectochilus formosanus Hayata, a rare and threatened medicinal Orchidaceae plant. J Zhejiang Univ Sci B 2013, 14: 785-792.
- 104. Zhang J, Wang LS, Gao JM, *et al.* Rapid separation and identification of anthocyanins from flowers of Viola yedoensis and V. prionantha by high-performance liquid chromatography-photodiode array detection-electrospray ionisation mass spectrometry. Phytochem Anal 2012, 23: 16-22.
- 105. Wang A, Lin L, Wang Y. Traditional Chinese Herbal Medicine Penthorum chinense Pursh: A Phytochemical and Pharmacological Review. Am J Chin Med 2015, 43: 601-620.

- 106. Luo Y, Yu H, Yang Y, *et al.* A flavonoid compound from Chrysosplenium nudicaule inhibits growth and induces apoptosis of the human stomach cancer cell line SGC-7901. Pharm Biol 2016, 54: 1133-1139.
- 107. Tian C, Zhang P, Yang C, et al. Extraction Process, Component Analysis, and In Vitro Antioxidant, Antibacterial, and Anti-Inflammatory Activities of Total Flavonoid Extracts from Abutilon theophrasti Medic. Leaves. Mediators Inflamm 2018.
- 108. Li Y, Xia H, Wu M, *et al.* Evaluation of the Antibacterial Effects of Flavonoid Combination from the Leaves of Dracontomelon dao by Microcalorimetry and the Quadratic Rotary Combination Design. Front Pharmacol 2017, 8: 70
- 109. Chen Y, Zhao J, Qiu Y, et al. Prenylated flavonoids from the stems and roots of Tripterygium wilfordii. Fitoterapia 2017, 119: 64-68.
- 110. Fan GW, Zhang Y, Jiang X, et al. Anti-inflammatory activity of baicalein in LPS-stimulated RAW264.7 macrophages via estrogen receptor and NF-kappaB-dependent pathways. Inflammation 2013, 36: 1584-1591.
- 111. Zhao QY, Yuan FW, Liang T, *et al.* Baicalin inhibits Escherichia coli isolates in bovine mastitic milk and reduces antimicrobial resistance. J Dairy Sci 2018, 101: 2415-2422.
- 112. Zeng Z, Qian L, Cao L, *et al.* Virtual screening for novel quorum sensing inhibitors to eradicate biofilm formation of Pseudomonas aeruginosa. Appl Microbiol Biotechnol 2008, 79: 119-126.
- 113. Yao J, Zhang Q, Min J, et al. Novel enoyl-ACP reductase (FabI) potential inhibitors of Escherichia coli from Chinese medicine monomers. Bioorg Med Chem Lett 2010, 20: 56-59.
- 114. Guan X, Zhou Y, Liang X, *et al.* Effects of compounds found in Nidus Vespae on the growth and cariogenic virulence factors of Streptococcus mutans. Microbiol Res 2012, 167: 61-68.
- 115. Xia F, Li X, Wang B, *et al.* Combination Therapy of LysGH15 and Apigenin as a New Strategy for Treating Pneumonia Caused by Staphylococcus aureus. Appl Environ Microbiol 2016, 82: 87-94.
- 116. Yu CH, Yu WY, Fang J, et al. Mosla scabra flavonoids ameliorate the influenza A virus-induced lung injury and water transport abnormality via the inhibition of PRR and AQP signaling pathways in mice. J Ethnopharmacol 2016, 179: 146-155.
- 117. Yan S, Wu B, Lin Z, et al. Metabonomic characterization of aging and investigation on the anti-aging effects of total flavones of Epimedium. Mol Biosyst 2009, 5: 1204-1213.
- 118. Zhang HX, Lunga PK, Li ZJ, *et al.* Flavonoids Submit a manuscript: https://www.tmrjournals.com/tmr

- and stilbenoids from Derris eriocarpa. Fitoterapia 2014, 95: 147-153.
- 119. Wang T, Liu Y, Li X, *et al.* Isoflavones from green vegetable soya beans and their antimicrobial and antioxidant activities. J Sci Food Agric 2018, 98: 2043-2047.
- 120. Jiang J, Wang RP, Hou MH, *et al.* Hydromethanolic extract of Rehum emodi exhibits significant antimicrobial activity against acute gastroenteriti bacterial strains. Microb Pathog 2018, 115: 179-182.
- 121. Zhou Z, Han N, Liu Z, *et al.* The antibacterial activity of phytochemically characterised fractions from Folium Syringae. Natur Prod Res 2014, 28: 1495-1498.
- 122. Njume C, Gqaza BM, Rozani C, et al. Studies on bioactivity and secondary metabolites of crude extracts of Bidens pilosa L. (Asteraceae): A medicinal plant used in the Transkei region of South Africa. Pak J Pharm Sci 2016, 29: 877-885.
- 123. Mu J, Liu T, Jiang L, *et al.* The Traditional Chinese Medicine Baicalein Potently Inhibits Gastric Cancer Cells. J Cancer 2016, 7: 453-461.
- 124. Xia J, Rong L, Sawakami T, et al. Shufeng Jiedu Capsule and its active ingredients induce apoptosis, inhibit migration and invasion, and enhances doxorubicin therapeutic efficacy in hepatocellular carcinoma. Biomed Pharmacother 2018, 99: 921-930.
- 125. Ren X, Zhang Z, Tian J, *et al.* The downregulation of c-Myc and its target gene hTERT is associated with the antiproliferative effects of baicalin on HL-60 cells. Oncol Lett 2017, 14: 6833-6840.
- 126. Tao Y, Zhan S, Wang Y, *et al.* Baicalin, the major component of traditional Chinese medicine Scutellaria baicalensis induces colon cancer cell apoptosis through inhibition of oncomiRNAs. Sci Rep 2018, 8: 14477.
- 127. Qiu P, Dong Y, Li B, *et al.* Dihydromyricetin modulates p62 and autophagy crosstalk with the Keap-1/Nrf2 pathway to alleviate ethanol-induced hepatic injury. Toxicol Lett 2017, 274: 31-41.
- 128. Deng J, Wang DX, Liang AL, et al. Effects of baicalin on alveolar fluid clearance and alpha-ENaC expression in rats with LPS-induced acute lung injury. Can J Physiol Pharmacol 2017, 95: 122-128.