

Ion channel mechanism of gastrointestinal motility and gastric hypersensitivity in functional dyspepsia: a review

Terigele Bao¹, Feng Lan¹, Guorui Li², Xiyele Mu¹, Ta Na¹, Minghai Fu¹, Yongsheng Chen^{1,2*}

¹School of Mongolian Medicine, Inner Mongolia University for Nationalities, Tongliao 028000, Inner Mongolia Autonomous Region, China. ²School of Life Sciences, Inner Mongolia University for Nationalities, Tongliao 028000, Inner Mongolia Autonomous Region, China.

*Corresponding to: YongSheng Chen, School of Mongolian Medicine, Inner Mongolia University for Nationalities, No.536, Huolinhe Street West, Korqin District, Tongliao City, Inner Mongolia Autonomous Region 028000, China. E-mail: Chenys_2000@163.com.

Author contributions

Minghai Fu and Yongsheng Chen were responsible for the conception and design of the manuscript. Terigele Bao, Feng Lan, Xiyele Mu and Ta Na wrote the draft. Yongsheng Chen and Guorui Li performed critical review and revision of the manuscript.

Competing interests

The authors declare no conflicts of interest.

Acknowledgments

"Double first-class" construction project of Mongolian medicine scientific research and innovation team fund (190301); International cooperative scientific and technological innovation project of Mongolian medicine standardization research (MDK2018009); 2018 National Civil Affairs Commission-Ministry of Education Mongolian Medicine R&D Engineering Key Laboratory Open Project (MDK2018056); Mongolian Medicine R&D National and Local Joint Engineering Research Center Open Fund Project (MDK2019044).

Abbreviations

FD, functional dyspepsia; SMC, smooth muscle cells; VIP, vasoactive intestinal polypeptide; NO, nitric oxide synthesis; ICC, interstitial cells of Cajal; ATP, adenosine triphosphate; TRP, transient receptor potential.

Peer review information

Gastroenterology & Hepatology Research thanks all anonymous reviewers for their contribution to the peer review of this paper.

Citation

Bao T, Lan F, Li GR, Mu X, Na T, Fu MH, Chen YS. Ion channel mechanism of gastrointestinal motility and gastric hypersensitivity in functional dyspepsia: a review. *Gastroenterol Hepatol Res.* 2022;4(3):15. doi: 10.53388/ghr2022-09-057.

Executive editor: Miao Peng.

Received: 10 August 2022; Accepted: 23 September 2022;

Available online: 28 September 2022.

© 2022 By Author(s). Published by TMR Publishing Group Limited.

This is an open access article under the CC-BY license.

(<https://creativecommons.org/licenses/by/4.0/>)

Abstract

Functional dyspepsia (FD) is a regularly diagnosed clinical gastrointestinal ailment with a high incidence rate that can considerably impact patients' health and quality of life and impose a substantial financial burden. Modern research on the pathophysiology of functional dyspepsia has not thoroughly explained the underlying reasons. The condition does not manifest any significant organ abnormalities, which raises the disease's difficulty coefficient. Major pathogenic exceptions in FD include gastrointestinal motor dysfunction, gastrointestinal hormone secretion problem, visceral hypersensitivity, and brain-gut axis. Several ion channels have reportedly been implicated in the pathophysiological process of FD. Therefore, it is crucial to comprehend the probable activities of various ion channels in FD. This study focuses on the current state of research on the possible role of several ion channels in the pathogenesis of FD.

Keywords: functional dyspepsia; ion channel; gastrointestinal motility; gastric hypersensitivity; smooth muscle

Introduction

Ion channels are macromolecular protein structures found in organelle plasma membranes or cell endoplasmic membranes capable of displaying distinct selective ion transmembrane transport capabilities and playing a crucial role in maintaining the specific permeability barrier [1]. It has been stated that about 400 linked protein families account for over 1 percent of the human gene pool. Malykhina originally introduced the idea of “channel illnesses” in 2004 and emphasized that several significant clinical symptoms of smooth muscle contraction may be controlled at the level of ion channels [2]. It has been found that ion channel anomalies are strongly associated with gastrointestinal illnesses. Thus, unraveling the molecular basis of gastrointestinal ion channel illnesses can offer a foundation for developing innovative diagnostic, differential diagnostic, and therapeutic techniques for Functional dyspepsia (FD) [3]. The classification of ion channels is based on their specific ion types, gating methods (activation or control), and molecular shapes. The essential ion channels are Na^+ , K^+ , Ca^{2+} , and Cl^- , as they predominate in smooth muscle cells of the gastrointestinal tract and govern their contraction [4].

FD is a functional gastrointestinal disorder that can present a variety of complicated gastrointestinal symptoms [5]. The most commonly diagnosed symptoms include epigastric distension, discomfort, postprandial fullness, early satiety, epigastric pain, etc. However, its clinical presentations may not be fully explained by organic, systemic, or metabolic illnesses [6]. Comorbidity, acute gastroenteritis, being female, smoking, using nonsteroidal anti-inflammatory medications, and helicobacter pylori infection are risk factors [7]. Rome III diagnostic criteria [8] provide that one or more of the following must be met: 1. early satiety, postprandial fullness, epigastric discomfort, and epigastric burning sensation; 2. no organic condition can explain the aforementioned symptoms; 3. at least six months before to diagnosis and approximately three months following diagnosis. According to a recent study, FD accounts for about one-third of outpatient disorders in the digestive medicine department, with an incidence rate of 18 to 23 percent [9]. In recent years, as a result of the tremendous growth of the economy, societal demand to preserve life has increased dramatically. It has been observed that the incidence rate of FD is growing, which might negatively impact the quality of life and social functions [10, 11]. Currently, the etiology and pathophysiology of FD are unknown; nevertheless, it has been proposed that the incidence and development of FD may be associated with many pathological processes, such as gastrointestinal motility disturbance, gastric hypersensitivity, and impaired stomach compliance [12]. Numerous investigations have demonstrated that ion channels are not only engaged in controlling gastrointestinal motility but also play a significant role in gastric hypersensitivity; these two activities have been identified as key contributors to the incidence and

progression of FD [5]. This article will examine the possible roles of several ion channels in the etiology and therapy of FD.

Ion channels can cause gastrointestinal motility disorders

Several related abnormalities are seen in patients with FD, including postprandial hypomotility of the stomach antrum, change in gastric rhythm, poor gastric-duodenal coordination delayed gastric emptying, and so on. The abnormality rate is between 20% and 54%, which has been linked to alterations in the patient's duodenal acid and lipid metabolism, excessive phasic contraction of the proximal stomach after a meal, and gastric dysfunction [12–14]. The cause of delayed gastric emptying is reduced gastric motility, which can severely reduce the motility of the stomach antrum. One of the critical reasons for FD [15, 16] is a failure of the stomach's motility, which can result in antrum overload. In FD, nausea, vomiting, and postprandial satiety are also believed to be associated with stomach emptying [17], and faster gastric emptying has been shown to alleviate these symptoms significantly [18]. In recent years, the successful use of calcium channel antagonists in the treatment of functional gastrointestinal motility disorders has suggested that the occurrence of these diseases may be linked to an imbalance of calcium homeostasis in smooth muscle cells; however, the mechanism of action requires further investigation.

The modulation of gastrointestinal motility has been linked to the incidence, development, and alleviation of functional dyspepsia. As a centrally regulated muscular organ, the gastrointestinal tract is made of multiple layers of smooth muscles. The fast contractions of smooth muscle cells (SMC) accompany intestinal motility. Therefore, it has been shown that the incidence of gastrointestinal motility problems is directly connected to changes in smooth muscle contraction. In the gastrointestinal SMC, there are three distinct ion channels (Figure 1): 1. voltage-gated sodium channels, potassium channels, calcium channels, etc. 2. ligand-gated channels. 3. mechanically-gated channels such as stretch-sensitive channels and volume-sensitive channels, etc. [19]. The fast contraction of smooth muscle cells is mediated by an increase in cytosolic Ca^{2+} concentration [20]. Calcium ions can bind to calmodulin, and the calcium-calmodulin complex can activate myosin light chain kinase and control myosin light chain by phosphorylating 20 kDa (MLC20). This process is the foundation for developing a single bridge between actin and myosin heavy chain and can govern smooth muscle contraction. L-type voltage-dependent calcium channels (L-Ca^{2+}) are the most prevalent and vital calcium channels in the smooth muscle of the digestive tract. It preferentially permits Ca^{2+} to cross the membrane, can participate in smooth muscle excitation and contraction coupling, and can be inhibited by nifedipine medicines. It has a high threshold, large conductance, slow inactivation, and inactivation dependent on intracellular Ca^{2+} concentration, etc. [21].

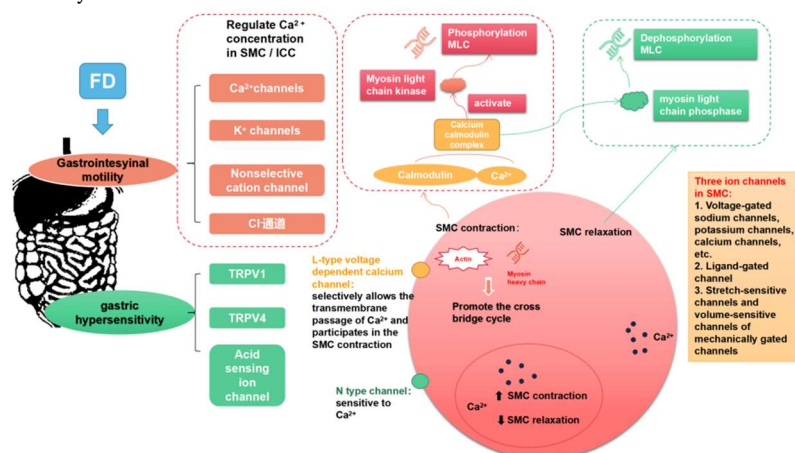


Figure 1: Ion channel and SMC constriction mechanism of gastrointestinal motility and visceral hypersensitivity observed in FD

Calcium ion channels regulate intracellular Ca^{2+} concentration changes

Extracellular calcium influx and intracellular calcium release have been shown as critical factors of SMC contraction. It has been shown that the contractile activity of the gastrointestinal smooth muscle is strongly connected to variations in intracellular Ca^{2+} concentration and that intracellular Ca^{2+} release can also serve as an efficient means of regulating smooth muscle contraction and relaxation [22]. When cells are stimulated externally, the intracytoplasmic Ca^{2+} concentration rises, which can bind to calmodulin and activate myosin light chain Kinase (MLCK). Activated MLCK can then phosphorylate the myosin light chain (MLC), which promotes the formation of cross-bridge cycles between actin and myosin and causes smooth muscle contraction. Myosin light chain phosphatase (MLCP) can dephosphorylate the myosin light chain and promote smooth muscle relaxation [23]. Ca^{2+} is the central transmitter that may successfully stimulate smooth muscle contraction, and it functions as the cell's ubiquitous secondary messenger. Thus, precise Regulation of intracellular Ca^{2+} release is an efficient means of controlling smooth muscle contraction and relaxation [24].

At present, smooth muscle cell calcium channels are categorized into four main categories: according to their various pharmacological and biophysical features, voltage-dependent calcium channels are classified as L-type, T-type, P-type, and N-type calcium channels. The L-type calcium channel is distinguished by its rapid inactivation, delayed and persistent attenuation, high threshold, large conductance, and intracellular Ca^{2+} concentration-dependent inactivation. It can manifest as a continuous inward current, typically triggered at -40–30 mV and 0–10 mV generates the most inward current. It is the predominant calcium channel found in the smooth muscle of the gastrointestinal tract. It can form action potentials in smooth muscle and can regulate the contraction activity of smooth muscle directly. Kurjak revealed that intestinal synaptosomes could display voltage-dependent Ca^{2+} channel properties and stimulate vasoactive intestinal polypeptide (VIP) release and nitric oxide synthesis (NO).

VIP release is predominantly associated with P- and N-type Ca^{2+} channels, but NO synthase-dependent Ca^{2+} can also pass via L-type Ca^{2+} channels. On the contrary, N-type calcium channels have not been cloned on smooth muscle cells of the gastrointestinal tract and interstitial cells of Cajal (ICC). Calcium channels of the T type can exhibit slow inactivation, quick decay, a low threshold, and poor conductance. It can appear as a transient inward current, often triggered around -70–60 mV, predominantly in neurons and myocardium. Receptors can modulate calcium channels, which are subdivided into ligand-gated calcium channels and, second messenger-gated calcium channels, stretch-activated calcium channels; background calcium channels [25].

ICC is mainly situated between the longitudinal and circular muscles of the gastrointestinal tract, near the myenteric plexus, and has been discovered to be strongly connected to gastrointestinal motor neurons and smooth muscle cells. Mesenchymal cells play a crucial part in the conductance of electrical activity and dyskinesia processes in the gastrointestinal tract. A recent article revealed that FD might be coupled with a decrease in the number of ICC or network destruction [26]. Interstitial cells are the primary pacemaker cells among interstitial cells, which can generate rhythmic depolarization and produce slow waves. Pacemaker current refers to the spontaneous inward current produced by these interstitial cells serving as pacemaker cells. The “pacing unit” consists of four parts: inosine triphosphate (IP3), ryanodine-sensitive calcium storage, mitochondria, and calcium-sensitive channels. It is the fundamental component that promotes automatic depolarization. The activation of the receptor causes the IP3 and ryanodine receptor-sensitive drug depots to release calcium, which can then trigger the process of mitochondrial uptake and consumption, which results in a significant decrease in the calcium ion concentration in a specific area of the cell, and can finally trigger the opening of low calcium sensitive channels. Moreover, the ion flow might induce the potential change, resulting in a repetitive electric likely change [27]. It was shown that calcium-activated chloride channels, voltage-dependent calcium channels, and intracellular calcium ion concentration are intimately associated with the formation of ICC pacing potential [28].

Table 1: Ion channels affecting gastrointestinal motility disorders in FD

Name	Type	Expression	
Calcium activated potassium channel	High conductance (BKCa), medium conductance (IKCa), and small conductance (SKCa)	Gastrointestinal smooth muscle, ICC	[28, 29]
Inward rectifier potassium channel	Classical (Kir2.1–2.4) K^+ transport channels (4.2, 5.1, 7.1) g protein gated (kir3.1–3.4) ATP sensitive (Kir6.1–6.2) channels	Some were expressed in smooth muscle cells, kir1.1, 4.1 in gastrointestinal smooth muscle and ICC	[30]
ATP sensitive potassium channel	-	Increase pacing frequency and action potential frequency of muscle cells	[31]
Delay rectifier channel	KV1.1, KV1.2, KV1.5, KV1.6, KV2.2		
Kv4 channel	-		
Slow delay rectifier channel	KCNQ or KV7		
HERG channel (KV11.1)	-	Mostly expressed in SMC	[32, 33]
BK channel (KCa1.1)	-		
SK channel (KCa2)	-		
IK channel (KCa3.1)	-		
Trek channel (K2P2.1)	-		
KV7	-	Rat proximal gastric muscle tension	[34]
Potassium bisphosphate (K2P) channel family	K2P2.1 and K2P5.1	Regulation of gastrointestinal smooth muscle contraction	[35]

K^+ channels and gastrointestinal motility cells

K^+ channels are the most numerous and diverse family of ion channels. All K^+ channels have been discovered to have a role in maintaining the membrane potential. Voltage-gated potassium channels and ligand-gated potassium channels can be distinguished among K^+ channels. Ligand-gated potassium channels include sodium-activated potassium channels, ATP-sensitive potassium

channels, adenosine-sensitive potassium channels, muscarinic potassium channels, and phosphatidylcholine-activated potassium channels.

Calcium-activated potassium channels are time- and voltage-dependent and primarily depends on the intracellular Ca^{2+} concentration. There are three different types, which are high-conductance (BKCa), medium-conductance (IKCa), and small-conductance (SKCa) calcium-activated potassium channels. BKCa, the most important of these three, is widely distributed in the

gastrointestinal smooth muscle and ICC because of its maximum conductivity [29, 30]. Moreover, inward rectifier potassium channels (Kir channels) are divided into seven types and 15 subtypes, and some are explicitly expressed in smooth muscle cells. Kir channels can be divided into four different functional groups: classic (Kir2.1–2.4), K⁺ transport channels (Kir1.1, 4.1 gastrointestinal smooth muscle and ICC, 4.2, 5.1, and 7.1), G protein gating (Kir3.1–3.4) and ATP-sensitive (Kir6.1–6.2) channels [31].

There are ATP-sensitive potassium channels located in the ICC of the small intestine and colon, which can maintain the resting potential of the ICC at -70 mV by regulating the regular influx of potassium ions. It has been reported that when the ATP-sensitive potassium channel is inhibited, the cell membrane can undergo depolarization, thereby causing ICC calcium influx, increasing the pacing frequency and the action potential frequency of the muscle cells [32]. A number of potassium channels are expressed in the cells that constitute SMC-ICC-PDGFR α ⁺ cell (SIP) syncytium, the most important among which are: delayed rectification (KV1.1, KV1.2, KV1.5, KV1.6, KV2.2) channel, type A (KV4) channel, slow delayed rectification (KCNQ or KV7) channel, HERG (KV11.1) channel, BK (KCa1.1) channel, SK (KCa2) channel, IK (KCa3. 1) channel, TREK (K2P2.1) channel [33, 34]. Most of these channels are expressed in SMC, including the smooth muscle of the gastrointestinal tract. The most important Kir channels in SIP syncytia are Kir2.1, 3, and 6 channels play a pivotal role in the regulation of smooth muscle movement. The former is expressed in ICC, and the latter is expressed in SMC [35].

Other studies have also shown that the KV7 channel can partially mediate the proximal gastric relaxation induced by VIP, which is one of the most important inhibitory neurotransmitters in the gastrointestinal tract [36]. It has been suggested that the KV7 activator might be considered a new possible drug for treating gastrointestinal dyskinesia in functional gastrointestinal diseases. In addition, the K2P channel family includes the K2P2.1 and K2P5.1 channel among 15 channels, which can play important physiological roles in the regulation of gastrointestinal smooth muscle contraction. Their opening probability increases significantly with stretching and in response to NO. These characteristics make them a vital constituent regulating the stability and relaxation of the smooth muscle membrane of the gastrointestinal tract [37].

Other ion channels and gastrointestinal motor cells

Currently, the existence of nonselective cation channels (NSCC) in a variety of smooth muscle cells has been confirmed. NSCC is the most prevalent form of stretch-activated ion channels (SAC channels). These channels display the following attributes: 1. they are primarily permeable to cations and demonstrate low permeability towards anions; 2. the permeability of cation selection is not high, and it can exhibit permeability towards various cations, including K⁺, Na⁺, Ca²⁺, and Mg²⁺; 3. the average value of single ion conductance is between 20 and 40 pS; 4. Gd³⁺ can effectively block it. In addition, investigations have demonstrated that the nonselective cation channel implicated in the pacing activity of gastrointestinal ICC-MY (ICC inside the intermuscular gap between myenteric areas) is a specialized kind of transient receptor potential (TRP) channel [38]. The TRP channel is predominantly a nonselective six-transmembrane cation channel with seven variants, TRPC4 and TRPM7, associated with the gastrointestinal ICC-MY pacing current [39]. It has been discovered that heterologous production of these two channels on HEK293 cells generates an inward current comparable to the ICC pacing current, suggesting that the pacing channel may be a member of the TRP channel family. Another study showed that the TRPV2 ion channel contributes considerably to mouse gastric adaption relaxation and stomach emptying. Mechanism-sensitive TRPV2 is expressed in the inhibitory motor neurons of the mouse stomach and is capable of inducing GAR and GE. Thus, TRPV2 may be a good target for GAR-impaired individuals [40].

The chloride channel is the primary anion channel on the cell membrane and is present in nearly all investigated cells. The chloride

ion channels serve various purposes, including transmembrane transport of ions and liquids, modulation of cell volume, and stabilization of cell membrane potential, among others. Many studies have discovered that the gastrointestinal tract contains a specific ANO1 protein (anoctamin 1/TMEM16A), also known as calcium-activated chloride channel protein, which functions as a particular marker protein of gastrointestinal ICC but has also been implicated in the production of slow waves [41–43]. This data suggested that the chloride channel may participate in the pacing processes. In addition, ICC-MY possesses a pK^a and ATP-activated chloride channel (PacC) that ATP and PKA can activate, produce a tail current, and only actively engage in the post-depolarization process of ICC-MY pacing activity [44]. Consequently, it is possible that these ion channels play a crucial role in FD-associated gastrointestinal motility abnormalities and contribute to the creation and modulation of pacing activity with other channel types.

Abnormal ion channels cause FD gastric hypersensitivity

Gastric hypersensitivity is commonly regarded as an additional significant element in the pathophysiology and development of FD. It has been observed that FD is more responsive than healthy volunteers to mechanical stimulation such as gastric distension [45]. The molecular basis of FD is yet unknown. However, gastric hypersensitivity and aberrant perception of gastrointestinal mechanical strain, temperature, and environmental variables may play a significant role in its formation. It has been demonstrated that sensory neuron-specific targets may be categorized into three distinct groups: receptors and sensors of peripheral nerve terminals, ion channels, and transmitter receptors associated with nerve excitability and conduction. Moreover, targets expressed explicitly by the numerous afferent neurons, such as those linked with transient receptor potential channels TRPV1, acid-sensing ion channels, and anti-tetraketotoxin Na channels [46], have the most significant therapeutic potential.

TRPV4 is a nonselective cation channel family that can be stretched and activated and is one of the drug candidates that can mediate visceral hypersensitivity [47]. TRPV4 was initially identified as a low molar concentration sensitive ion channel, which can be activated by mechanical stimulation, hyperthermia, and epoxyeicosatrienoic acid. It has been found that ATP released by Enterobacteria, activated submucosal basophils, and mast cells can also stimulate inhibitory nerve endings in the intestine. It has been reported that TRPV4 itself is significantly up-regulated in the inflamed gastrointestinal mucosa; therefore, this stretch-activated family of nonselective cation ion channels may interact with the different inflammatory mediators (IL-1 β , TNF- α , IL-6) in the intestine and/or the mucosal immune system [48, 49].

The acid-sensing ion channel (ASIC) is a member of the degraded protein/epithelial sodium channel (DEG/ENaC) superfamily. It can actively participate in the process of nociception by sensing protons and is involved in regulating inflammatory chronic visceral hyperalgesia [50, 51]. There is ample evidence to suggest that ASICs, especially ASIC1 and ASIC3, can cause major chemical and mechanical allergies related to inflammatory and non-inflammatory conditions in the gastrointestinal tract [52–55]. Acid-sensitive ion channel 5 (ASIC5) mRNA was expressed in duodenal epithelial cells. It was found that ASIC5 may act as a pain-related chemical receptor or a sensor of bile acid [56]. These results indicated that ASIC5 might be related to the gallbladder and Oddi sphincter disorders in FD. There is also some evidence that TRPV1 might be related to hyperalgesia throughout the digestive tract. TRPV1 can also serve as an important marker in treating upper gastrointestinal pain in FD [57].

Discussion

Mounting evidence indicates that ion channels are attractive targets for FD that can be effective in various pathophysiological conditions. Numerous studies have initiated research tracks to identify the

treatment of FD through ion channel mechanisms. The study on the interventional mechanism of electroacupuncture (EA) of “Zusanli” (ST36) based on the involvement of mast cells/TRPV1 signaling pathway in relieving visceral hypersensitivity in FD rats reveals that gastric compliance was significantly decreased, and the levels of visceral sensitivity increased in the FD rats. TRPV1 immunofluorescence intensity, expression of PAR2 and TRPV1 proteins, and contents of SP and CGRP in the stomach were considerably up-regulated in FD rats, and these symptoms were improved after EA intervention, which may be related to its effects in inhibiting the activation of down-regulating the expression of gastric PAR2 and TRPV1 proteins [58]. Another study on quercetin in the treatment of FD showed that quercetin-induced relaxation of human gastric smooth muscle occurs directly through K^+ channels and suggested that quercetin is a potential nutraceutical in the treatment of functional dyspepsia [59].

Currently, treating FD through ion channels includes endogenous, natural compounds, prescription, and physical intervention. The ion channel may be a better candidate for the therapy of FD as it is specifically relevant to gastrointestinal motility and gastric hypersensitivity. However, based on the massive family of ion channels, it will not only be a valuable method but also be a challenge for elucidating the pathogenesis and treatment of FD through ion channels.

Conclusion

In conclusion, various distinct ion channels or subtypes can target diverse gastrointestinal cells to regulate some crucial gastrointestinal activities, including functional dyspepsia. It has been discovered that channel blockers such as pinaverium bromide and otilonium bromide can greatly alleviate gastrointestinal smooth muscle spasms and diminish visceral hypersensitivity, albeit with a limited impact. Expanding study is being conducted on the mechanics of ion channels in functional dyspepsia. Therefore, the multi-ion channel modifying components may be considered a viable novel FD medication therapy strategy. Moreover, innovative features or multi-targeted natural medicines should be created to selectively adjust the particular assembly of these channel subunits and their associated accessory proteins. If these strategies are effective, ion channel modulators may become essential medications for treating FD symptoms and may have a substantial role in treating other gastrointestinal tract-related illnesses.

References

1. B. Hille. Ionic channels: Molecular pores of excitable membranes. *Harvey Lect* 1986;82:47–69. Available at: <https://pubmed.ncbi.nlm.nih.gov/2452140>
2. Yang H, Hou C, Xiao W, Qiu Y. The role of mechanosensitive ion channels in the gastrointestinal tract. *Front Physiol* 2022;13:904203. Available at: <https://doi.org/10.3389/fphys.2022.904203>
3. Sugiyama T, Shiotani A. The Cutting Edge Research of Functional Gastrointestinal Disorders in Japan: Review on JGA Core Symposium 2018-2020. *Digestion* 2021;102:6–11. Available at: <https://doi.org/10.1159/000510680>
4. Alexander SP, Benson HE, Faccenda E, et al. The Concise Guide to PHARMACOLOGY 2013/14: Ion channels. *Br. J. Pharmacol* 2013;170:1607–1651. Available at: <https://doi.org/10.1111/bph.12447>
5. Ford AC, Mahadeva S, Carbone MF, Lacy BE, Talley NJ. Functional Dyspepsia. *Lancet* 2020;10:1–14. Available at: [https://doi.org/10.1016/S0140-6736\(20\)30469-4](https://doi.org/10.1016/S0140-6736(20)30469-4)
6. Drossman DA, Hasler WL. Rome IV-Functional GI Disorders: Disorders of Gut-Brain Interaction. *Gastroenterology* 2016;150(6):1257–61. Available at: <https://doi.org/10.1053/j.gastro.2016.03.035>
7. Zhang SS, Zhao LQ. Consensus opinion of experts in TCM diagnosis and treatment of functional dyspepsia. *Chinese Journal of traditional Chinese medicine* 2017;32:2595–2598. (Chinese) Available at: <http://125.221.83.226:18/rwt/CNKI/https://NNYHGLUDN3WX TLUPMW4A/kcms/detail/detail.aspx?FileName=BXY201706072&DbName=CJFQ2017>
8. Sperber AD, Bangdiwala SI, Drossman DA, et al. Worldwide prevalence and burden of functionalgastrointestinal disorders, results of Rome foundation global study. *Gastroenterology* 2021;160:99–114. Available at: <https://doi.org/10.1053/j.gastro.2020.04.014>
9. Ford AC, Marwaha A, Sood R, Moayyedi P. Global prevalence of, and risk factors for, uninvestigated dyspepsia: a meta-analysis. *Gut* 2015;64:1049–1057. Available at: <https://doi.org/10.1136/gutjnl-2014-307843>
10. Ji S, You Y, Peng B, et al. Multi-omics analysis reveals the metabolic regulators of duodenal low-grade inflammation in a functional dyspepsia model. *Front Immunol* 2022;24:944591. Available at: <https://doi.org/10.3389/fimmu.2022.944591>
11. Black CJ, Paine PA, Agrawal A, et al. British Society of Gastroenterology guidelines on the management of functional dyspepsia. *Gut* 2022;71:1697–1723. Available at: <https://doi.org/10.1136/gutjnl-2022-327737>
12. Singh R, Zogg H, Ghoshal UC, Ro S. Current Treatment Options and Therapeutic Insights for Gastrointestinal Dysmotility and Functional Gastrointestinal Disorders. *Front Pharmacol* 2022;13:808195. Available at: <https://doi.org/10.3389/fphar.2022.808195>
13. Bisschops R, Karamanolis G, Arts J, et al. Relationship between symptoms and ingestion of a meal in functional dyspepsia. *Gut* 2008;57:1495–1503. Available at: <https://doi.org/10.1136/gut.2007.137125>
14. van Lelyveld N, Schipper M, Samsom M. Samsom. Lack of relationship between chronic upper abdominal symptoms and gastric function in functional dyspepsia. *Dig. Dis. Sci* 2008;53(5):1223–1230. Available at: <https://doi.org/10.1007/s10620-007-0012-1>
15. Min YW, Lee H, Ahn S, et al. Eosinophil and Mast Cell Counts in the Stomach and Duodenum of Patients with Functional Dyspepsia without a Helicobacter pylori infection. *Korean J Gastroenterol* 2022;80:28–33. Available at: <https://doi.org/10.4166/kjg.2022.036>
16. Tang L, Zeng Y, Li L, et al. Electroacupuncture Upregulated Ghrelin in Rats with Functional Dyspepsia via AMPK/TSC2/Rheb-Mediated mTOR Inhibition. *Dig Dis Sci* 2022;65:1689–1699. Available at: <https://doi.org/10.1007/s10620-019-05960-5>
17. Sarnelli G, Caenepeel P, Geypens B, Janssens J, Tack J. Symptoms associated with impaired gastric emptying of solids and liquids in functional dyspepsia. *Am. J. Gastroenterol* 2003;98:783–788. Available at: <https://doi.org/10.1053/j.gastro.2019.01.249>
18. Vijayvargiya P, Camilleri M, Chedid V, Mandawat A, Erwin PJ, Murad MH. Effects of promotility agents on gastric emptying and symptoms:a systematic review and meta-analysis. *Gastroenterology* 2019;156:1650–1660. Available at: <https://doi.org/10.1111/j.1572-0241.2003.07389.x>
19. Li Z, Xu W. Ion channels in gastrointestinal smooth muscle cells and their modulation. *Basic Medicine and Clinic* 2003;23:19. (Chinese) Available at: http://125.221.83.226:18/rwt/CNKI/https://PRYGG5ULNEYG63 LV/kcms/detail?v=1BH6_3vDKzMGQa1Dc8ckQb1r9C13JY4M 2xMLi3JEpdCPQTVLk1f8zVsARLtxEYq3zwhskZms-AR3CwOo3 woE3StWzzXqheRSe2LIspkHDY_NF8A8VebA= =&uniplatform =NZKPT&language=CHS
20. Sanders KM, Koh SD, Ro S, Ward SM. Regulation of gastrointestinal motility-insights from smooth muscle biology.

- Nat Rev Gastroenterol Hepatol* 2012;9:633–645. Available at: <https://doi.org/10.1038/nrgastro.2012.168>
21. Yang B. Ion channel pharmacology. 1st edition. *Beijing: People's Health Publishing House* 2015;15. (Chinese) Available at: ISBN: 9787117066341
 22. Kitazawa T, Matsui T, Katsuki S, et al. A temporal Ca^{2+} desensitization of myosin light chain kinase in phasic smooth muscles induced by CaMKK β /PP2A pathways. *Am J Physiol Cell Physiol* 2021; 321:549–558. Available at: <https://doi.org/10.1152/ajpcell.00136.2021>
 23. Filter JJ, Williams BC, Eto M, Shalloway D, Goldberg ML. Unfair competition governs the interaction of pCPI-17 with myosin phosphatase (PP1-MYPT1). *Elife* 2017;6:e24665. Available at: <https://doi.org/10.7554/eLife.24665>
 24. Sanders KM. Spontaneous Electrical Activity and Rhythmicity in Gastrointestinal Smooth Muscles. *Adv Exp Med Biol* 2019;1124: 3–46. Available at: https://doi.org/10.1007/978-981-13-5895-1_1
 25. Liu J. Cell information and regulation. *China Union Medical College Press* 2004;220. (Chinese) Available at: ISBN: 7810724606
 26. Mikkelsen HB. Interstitial cells of Cajal, macrophages and mast cells in the gut musculature: morphology, distribution, spatial and possible functional interactions. *J Cell Mol Med* 2010;14:818–832. Available at: <https://doi.org/10.1111/j.1582-4934.2010.01025.x>
 27. Wang Z, Xu W. Latest trends in the study of gastrointestinal smooth muscle pacing function. *World Chinese Journal of digestion* 2010;18:319–323. (Chinese) Available at: <http://125.221.83.226:18/rwt/CNKI/https://NNYHGLUDN3WX.TLUPMW4A/kcms/detail/detail.aspx?FileName=XXHB201004003&DbName=CJFQ2010>
 28. Di G, Lei S. Research progress on Cajal cells and their ion channel regulation mechanism. *Medical Review* 2017;23:1457–1460 + 1465. (Chinese) Available at: <http://125.221.83.226:18/rwt/CNKI/https://NNYHGLUDN3WX.TLUPMW4A/kcms/detail/detail.aspx?FileName=YXZS201708001&DbName=CJFQ2017>
 29. Wang W, Huang H, Hou D, et al. Mechanosensitivity of STREX-lacking BKCa channels in the colonic smooth muscle of the mouse. *Am J Physiol* 2010;299:G1231–1240. Available at: <https://doi.org/10.1152/ajpgi.00268.2010>
 30. Shen XX, Zhang L, Jiang L, et al. Alteration of sphingosine-1-phosphate with aging induces contractile dysfunction of colonic smooth muscle cells via Ca^{2+} -activated K^+ channel (BKCa) upregulation. *Neurogastroenterol Motil* 2021;33:14052. Available at: <https://doi.org/10.1111/nmo.14052>
 31. Hibino H, Inanobe A, Furutani K, Murakami S, Findlay I, Kurachi Y. Inwardly rectifying potassium channels: their structure, function, and physiological roles. *Physiol Rev* 2010;90:291–366. Available at: <https://doi.org/10.1152/physrev.00021.2009>
 32. Na JS, Hong C, Kim MW, et al. ATP-sensitive K^+ channels maintain resting membrane potential in interstitial cells of Cajal from the mouse colon. *Eur J Pharmacol* 2017;809:98–104. Available at: <https://doi.org/10.1016/j.ejphar.2017.05.029>
 33. Koh SD, Ward SM, Sanders KM. Ionic conductances regulating the excitability of colonic smooth muscles. *Neurogastroenterol Motil* 2012;24:705–718. Available at: <https://doi.org/10.1111/j.1365-2982.2012.01956.x>
 34. Sanders KM. Regulation of smooth muscle excitation and contraction. *Neurogastroenterol Motil* 2008;20:39–53. Available at: <https://doi.org/10.1111/j.1365-2982.2008.01108.x>
 35. Ipavec V, Martire M, Barrese V, Tagliatela M, Currò D. KV7 channels regulate muscle tone and nonadrenergic noncholinergic relaxation of the rat gastric fundus. *Pharmacol Res* 2011;64:397–409. Available at: <https://doi.org/10.1016/j.phrs.2011.06.016>
 36. Huang X, Lee SH, Lu H, Sanders KM, Koh SD. Molecular and functional characterization of inwardly rectifying K^+ currents in murine proximal colon. *J Physiol* 2018;596:379–391. Available at: <https://doi.org/10.1113/JP275234>
 37. Alcaino C, Farrugia G, Beyder A. Chapter Eight- Mechanosensitive Piezo Channels in the Gastrointestinal Tract. *Curr Top Membr* 2017;79:219–244. Available at: <https://doi.org/10.1016/bs.ctm.2016.11.003>
 38. Lee JH, Wu WH, Huang XY, Jun JY, Choi S. Transient Receptor Potential Canonical 4 and 5 Channel Antagonist ML204 Depolarized Pacemaker Potentials of Interstitial Cells of Cajal. *Neurogastroenterol Motil* 2020;26:521–528. Available at: <https://doi.org/10.5056/jnm20064>
 39. Kim BJ, Lim HH, Yang DK, et al. Melastatin-type transient receptor potential channel 7 is required for intestinal pacemaking activity. *Gastroenterology* 2005;129:1504–1517. Available at: <https://doi.org/10.1053/j.gastro.2005.08.016>
 40. Mihara H, Suzuki N, Yamawaki H, Tominaga M, Sugiyama T. TRPV2 ion channels expressed in inhibitory motor neurons of gastric myenteric plexus contribute to gastric adaptive relaxation and gastric emptying in mice. *Am J Physiol Gastrointest Liver Physiol* 2013;304:G235–40. Available at: <https://doi.org/10.1152/ajpgi.00256.2012>
 41. Sforza L, Michelucci A, Morena F, et al. Piezo1 controls cell volume and migration by modulating swelling-activated chloride current through Ca^{2+} influx. *J Cell Physiol* 2022;237:1857–1870. Available at: <https://doi.org/10.1002/jcp.30656>
 42. Liu Y, Liu Z, Wang K. The Ca^{2+} -activated chloride channel ANO1/TMEM16A: an emerging therapeutic target for epithelium-originated diseases? *Acta Pharm Sin B* 2021;11:1412–1433. Available at: <https://doi.org/10.1016/j.apsb.2020.12.003>
 43. Baker SA, Hwang SJ, Blair PJ, et al. Ca^{2+} transients in ICC-MY define the basis for the dominance of the corpus in gastric pacemaking. *Cell Calcium* 2021;99:102472. Available at: <https://doi.org/10.1016/j.ceca.2021.102472>
 44. Parsons SP, Sanders KM. An outwardly rectifying and deactivating chloride channel expressed by interstitial cells of cajal from the murine small intestine. *J Membr Biol* 2008;221:123–132. Available at: <https://doi.org/10.1007/s00232-007-9084-2>
 45. Duan S, Kondo T, Miwa H, et al. Eosinophil-associated microinflammation in the gastroduodenal tract contributes to gastric hypersensitivity in a rat model of early-life adversity. *Am J Physiol Gastrointest Liver Physiol* 2021;320:G206–G216. Available at: <https://doi.org/10.1152/ajpgi.00313.2020>
 46. Holzer P. Gastrointestinal pain in functional bowel disorders: sensory neurons as novel drug targets. *Expert Opin Ther Targets* 2004;8:107–23. Available at: <https://doi.org/10.1517/14728222.8.2.107>
 47. Balemans D, Boeckstaens GE, Talavera K, Wouters MM. Transient receptor potential ion channel function in sensory transduction and cellular signaling cascades underlying visceral hypersensitivity. *Am J Physiol* 2017;312:G635–648. Available at: <https://doi.org/10.1152/ajpgi.00401.2016>
 48. Rayees S, Joshi JC, Tauseef M, et al. PAR2-mediated camp generation suppresses TRPV4-dependent Ca^{2+} signaling in alveolar macrophages to resolve TLR4-induced inflammation. *Cell Rep* 2019;27:793–805. Available at: <https://doi.org/10.1016/j.celrep.2019.03.053>
 49. Li M, Fang XZ, Zheng YF, et al. Transient receptor potential vanilloid 4 is a critical mediator in LPS mediated inflammation

- by media ting calcineurin/NFATc3 signaling. *Biochem Biophys Res Commun* 2019;513:1005–1012. Available at: <https://doi.org/10.1016/j.bbrc.2019.04.020>
50. Wemmie JA, Taugher RJ, Kreple CJ. Kreple. Acid-sensing ion channels in pain and disease. *Nat Rev Neurosci* 2013;14:461–471. Available at: <https://doi.org/10.1038/nrn3529>
 51. Hummel M, Knappenberger T, Reilly M, Whiteside GT. Pharmacological evaluation of NSAID-induced gastropathy as a “Translatable” model of referred visceral hypersensitivity. *World J Gastroenterol* 2017;23:6065–6076. Available at: <https://doi.org/10.3748/wjg.v23.i33.6065>
 52. Holzer P. Acid-sensing ion channels in gastrointestinal function. *Neuropharmacology* 2015;94:72–79. Available at: <https://doi.org/10.1016/j.neuropharm.2014.12.009>
 53. Wang HJ, Xu X, Zhang PA, et al. Epigenetic upregulation of acid-sensing ion channel 1 contributes to gastric hypersensitivity in adult offspring rats with prenatal maternal stress. *Pain* 2020;161:989–1004. Available at: <https://doi.org/10.1097/j.pain.0000000000001785>
 54. Matricon J, Muller E, Accarie A, et al. Peripheral contribution of NGF and ASIC1a to colonic hypersensitivity in a rat model of irritable bowel syndrome. *Neurogastroenterol Motil* 2013;25:e740–754. Available at: <https://doi.org/10.1111/nmo.12199>
 55. Zhang L, Zheng L, Yang X, et al. Pathology and physiology of acid-sensitive ion channels in the digestive system (Review). *Int J Mol Med* 2022;50:94. Available at: <https://doi.org/10.3892/ijmm.2022.5150>
 56. Vyvers A, Schmidt A, Wiemuth D, Gründer S. Screening of 109 neuropeptides on ASICs reveals no direct agonists and dynorphin A, YFMRFamide and endomorphin-1 as modulators. *Sci Rep* 2018;8:18000. Available at: <https://doi.org/10.1038/s41598-018-36125-5>
 57. Sarnelli G, Pesce M, Seguela L, et al. Impaired Duodenal Palmitoylethanolamide Release Underlies Acid-Induced Mast Cell Activation in Functional Dyspepsia. *Cell Mol Gastroenterol Hepatol* 2021;11:841–855. Available at: <https://doi.org/10.1016/j.jcmgh.2020.10.001>
 58. Dong JZ, Rong PJ, Ma TM, Wang D, Wang XT, Qiao Y. Influence of electroacupuncture of zusanli (ST36) on mast cells/TRPV1 signaling pathway in visceral hypersensitivity rats with functional dyspepsia. *Zhen Ci Yan Jiu* 2022;47:592–7. (Chinese) Available at: <https://doi.org/10.13702/j.1000-0607.20210937>
 59. Modzelewska B, Drygalski K, Kleszczewski T, et al. Quercetin relaxes human gastric smooth muscles directly through ATP-sensitive potassium channels and not depending on the nitric oxide pathway. *Neurogastroenterol Motil* 2021; 33:e14093. Available at: <https://doi.org/10.1111/nmo.14093>