

Chemical constituents, pharmacology and safety of isoflavones in *Puerariae Lobatae Radix*

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Author contributions

The study was conceived by Han-Qing Pang and Yong-Juan Zhao, the review was designed by Xiao-Yu Shang, Jia-Lu Li, Tong-Yu Zhou, Hao-Feng Wang and Han-Qing Pang. Literature collection by Xiao-Yu Shang, Jia-Lu Li, Yan Zhu and Tong-Yu Zhou. The manuscript was drafted by Xiao-Yu Shang, Jia-Lu Li, Yan Zhu, Tong-Yu Zhou, Zhe Tao, and Hao-Feng Wang, with figures drawn by Tong-Yu Zhou, Hao-Feng Wang, Yang Zhou, Tong Su and Bin-Bin Zeng. Language polishing was performed by Han-Qing Pang, Yong-Juan Zhao and Yu Chen.

Competing interests

The authors declare no conflicts of interest.

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Abbreviations

PLR, Pueraria Lobatae Radix.

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Abstract

Pueraria Lobatae Radix (PLR), also known as kudzu root, is abundant in various of active compounds. Among these compounds, isoflavones (puerarin, daidzein, genistein, etc.) is extensive studied due to their extensive pharmacological properties. This review focuses on the chemical compounds, pharmacological effects, action mechanisms and clinical studies of the isoflavone in PLR to offer new insights for prospective research of PLR. Isoflavones in PLR possessed multiple pharmacological effects, such as anti-inflammatory, anti-oxidation and neuroprotection. Studies have shown that isoflavones are expected to be applied in cardiovascular diseases, intestinal diseases, diabetes, liver disorders, and neurological conditions. Although isoflavones derived from PLR exhibit therapeutic potential for treating a variety of diseases, they may also lead to adverse reactions, such as gastrointestinal discomfort, estrogen-like effects, hepatotoxicity and nephrotoxicity. Therefore, clinical investigations should be carried out to ascertain the pharmacological actions of isoflavones obtained from the laboratory and animal studies, ensuring their safety and effectiveness in humans. More studies on developing advanced drug delivery approaches to improve the bioavailability of isoflavones and their effectiveness, as well as exploring their precise molecular and cellular mechanisms, will be useful in developing new drugs and novel therapies.

Keywords: Pueraria Lobatae Radix; isoflavones; pharmacological effects; side effects; safety and risks

Introduction

Pueraria Lobatae Radix (PLR), commonly known as kudzu, is a climbing plant native to East Asia and widely used in traditional Chinese medicine [1]. PLR contained isoflavones, flavonoids, triterpenoids and saponins, organic acids, polysaccharides and alkaloids, among which isoflavones are the highest and are also the active components of PLR. The isoflavones (puerarin, daidzein, genistein, and tectoridin, etc.) have been extensively studied for their potential benefits in cardiovascular diseases, diabetes, liver diseases, neurological disorders. Recent research suggested that these isoflavones executed their biological effects by influencing key cellular signaling pathways, such as reducing the generation of inflammatory mediators, protecting cells from oxidative damage, and regulating blood sugar levels [2], and endocrine regulation. The molecular structure of these isoflavones were similar with that of estrogen, which could affect estrogen-related receptors, thereby providing therapeutic potential for various diseases [3].

Despite isoflavones in PLR exerted various pharmacological activities, the development of these isoflavones in clinical applications also faced some challenges. The most significant ones included their low bioavailability and potential safety issues. The low absorption and metabolism efficiency of these isoflavones limited their effectiveness and application scope. Advanced drug delivery systems such as nanotechnology have been utilized to enhance the bioavailability of these compounds [4]. Moreover, due to their estrogen-like effects, there may be safety risks for specific populations, such as patients with hormone-sensitive diseases [5]. To reduce the potential risks caused by isoflavones from PLR, some clinical studies have been used to validate their safety and efficacy, especially their potential applications in human health and disease treatment [6].

The present review summarized the modern research progress on the chemical compounds, pharmacological actions, side effects, and clinical application of isoflavones from PLR. It offers overall knowledge of isoflavones from PLR and some deficiencies were also proposed, which could accelerate the further development of PLR isoflavones related products. The whole plant and Chinese herbal medicines of PLR were shown in Figure 1.

Materials and methods

The accessible literatures on isoflavones from PLR were obtained from published materials of electronic databases, such as SCI finder, PubMed, Wed of Science, Springer and Google Scholar. The relevant information was also acquired from *Chinese Pharmacopoeia* and Chinese herbal classic books. ChemDraw Ultra 20.0 software was used to draw the chemical structures. Figdraw was utilized to draw the action mechanisms of isoflavones in nervous system, cardiovascular system and intestinal tract diseases.

Ingredients of Pueraria mirifica isoflavones

Composition and contents

Currently, more than 100 types of isoflavones have been isolated in PLR, mainly including puerarin, daidzein, tectoridin, and irisolidone [7] (see Table 1 and Figure 2). The types and contents of isoflavones varied significantly among different plants parts of PLR. The roots and leaves are main sources of isoflavones, and the total contents of isoflavones decreased in the following order: roots > leaves > stems > flowers. The content of puerarin in roots was the highest, while the contents of irisolidone and daidzein were relatively high in leaves [8]. Moreover, the growth environment also significantly affected the contents and types of PLR isoflavones [9]. Some studies have indicated that puerarin, irisolidone, and daidzein and their derivatives were the main PLR isoflavones [9].

Structures

Isoflavones in PLR are a class of plant estrogens, most of which are glycosidic compounds with a 3-phenylchroman structure [10]. These isoflavones had a basic skeleton of C_6 - C_3 - C_6 combined with glycone or glycoside to form phenolic compounds. Isoflavone glycosides have a large π - π conjugated system [11, 12], forming an approximately planar structure in space. The types of isoflavones in PLR was extremely complex, containing many different isomers.



Figure 1 Whole plant and Chinese herbal medicines of *Pueraria lobata*. (A) Whole plant of *Pueraria lobata*. (B) Chinese herbal medicines of *Pueraria lobata*.

Table 1 Compound	l. molecular formula.	CAS and logn of	major isoflavones in PLR
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NO	Compound	Molecular formula	CAS	Logp	Reference
1	Puerarin	$C_{21}H_{20}O_{10}$	3681-99-0	-0.67	[12]
2	Daidzein	$C_{15}H_{10}O_4$	486-66-8	2.78	[13]
3	Genistein	$C_{15}H_{10}O_5$	446-72-0	2.96	[14]
4	Prunetin	C ₁₆ H ₁₂ O5	552-59-0	3.53	[15]
5	Genistein-7-O-β-D-glucoside	$C_{21}H_{20}O_{10}$	529-59-9	0.79	[16]
6	Daidzin	$C_{21}H_{20}O_9$	552-66-9	0.45	[17]
7	Kakkalide	$C_{28}H_{32}O_{15}$	58274-56-9	1.62	[18]
8	Glycitein	$C_{16}H_{12}O_5$	40957-83-3	2.57	[19]
9	Glycitin	$C_{22}H_{22}O_{10}$	40246-10-4	0.16	[20]
10	Tectoridin	$C_{22}H_{22}O_{11}$	611-40-5	0.29	[21]

PLR, Pueraria Lobatae Radix.

NO	Compound	Molecular formula	CAS	Logp	Reference
11	Tectorigenin	$C_{16}H_{12}O_{6}$	548-77-6	2.54	[22]
12	Formononetin	$C_{16}H_{12}O_4$	485-72-3	2.96	[23]
13	3'-Hydroxyl daidzein	$C_{15}H_{10}O_5$	485-63-2	2.58	[24]
14	3'-Methoxy daidzein	$C_{16}H_{12}O_5$	21913-98-4	2.54	[15]
15	3'-Methoxypuerarin	$C_{22}H_{22}O_{10}$	117047-07-01	1.72	[25]
16	3'-Hydroxypuerarin	$C_{21}H_{20}O_{10}$	117060-54-5	1.75	[15]
17	Isoformononetin	$C_{16}H_{12}O_4$	486-63-5	3.17	[25]
18	Irisolidone	$C_{17}H_{14}O_6$	2345-17-7	2.88	[26]
19	4;-methoxypuerarin	$C_{22}H_{22}O_9$	92117-94-7	2.14	[27]
20	BiochaninA	$C_{16}H_{12}O_5$	491-80-5	3.14	[28]
21	Ononin	$C_{22}H_{22}O_9$	486-62-4	0.63	[29]
22	NeopuerarinA	$C_{21}H_{20}O_9$	1150314-34-3	0.90	[30]
23	NeopuerarinB	$C_{21}H_{20}O_9$	1150314-39-8	0.90	[30]
24	8-prenyldaidzein	$C_{20}H_{18}O_4$	135294-00-8	4.87	[28]
25	Lupiwighteone	$C_{20}H_{18}O_5$	104691-86-3	5.05	[31]
26	Wighteone	$C_{20}H_{18}O_5$	51225-30-0	5.05	[32]
27	6,7,4'-Trihydroxyisoflavone	$C_{15}H_{10}O_5$	17817-31-1	2.17	[33]
28	IristectorigeninA	$C_{17}H_{14}O_7$	39012-01-6	2.59	[34]
29	Puerarin-6"-O-xyloside	$C_{26}H_{28}O_{13}$	114240-18-5	1.48	[35]
30	Mirificin	$C_{26}H_{28}O_{13}$ $C_{26}H_{28}O_{13}$	103654-50-8	1.21	[36]
31	Mirificin-4'-O-glucoside	$C_{32}H_{38}O_{18}$	168035-01-6	-1.44	[37]
32	Puerarin-4'-O-glucoside	C ₂₇ H ₃₀ O ₁₄	117047-08-2	-0.70	[37]
33	Genistein-8-C-glucoside	$C_{21}H_{20}O_{10}$	66026-80-0	0.09	[39]
33 34	6"-O-malonyl genistin	$C_{24}H_{22}O_{13}$	51011-05-3	2.22	[39]
35	Kakkalidone	$C_{23}H_{24}O_{11}$	6009-88-7	0.36	[40]
35 36	6"-O-xylosyl-glycitin	$C_{23}H_{24}O_{11}$ $C_{27}H_{30}O_{14}$	231288-18-9	1.37	[41]
30 37	Tectorigenin-7-O-xylosylglucoside	$C_{27}H_{30}O_{14}$ $C_{27}H_{30}O_{15}$	231288-19-0	1.37	[42]
38	Daidzein-4', 7-diglucoside	$C_{27}H_{30}O_{15}$ $C_{27}H_{30}O_{14}$	53681-67-7	-2.20	[42]
30 39	Ambocin	$C_{27}H_{30}O_{14}$ $C_{26}H_{28}O_{14}$	108044-05-9	-2.20 1.47	[43]
39 40	8-O-methyl retusin	$C_{26}H_{28}O_{14}$ $C_{17}H_{14}O_5$	37816-20-9	2.32	[33]
			2746-85-2		
41 42	Fujikinetin methyl ether	$C_{18}H_{14}O_6$		3.44	[45]
42 42	Calycosin 7-O-glucoside	$C_{22}H_{22}O_9$	20633-67-4	0.09	[46]
43	Psi-Tectorigenin	$C_{16}H_{12}O_{6}$	13111-57-4	2.22	[47]
44	Pseudobaptigenin	$C_{16}H_{10}O_5$	90-29-9	3.06	[48]
45	5-Hydroxypseudobaptigenin	$C_{16}H_{10}O_{6}$	40624-03-1	3.24	[15]
46	4',7-Dimethoxyisoflavone	$C_{17}H_{14}O_4$	1157-39-7	3.43	[25]
47	Isoflavone	$C_{15}H_{10}O_2$	574-12-9	3.58	[49]
48	3'-methoxy-daidzin	$C_{22}H_{20}O_{10}$	200127-80-6	0.22	[25]
49	Neobavaisoflavone	$C_{20}H_{18}O_4$	41060-15-5	4.87	[50]
50	Corylin	$C_{20}H_{16}O_4$	53947-92-5	4.45	[51]
51	IristectorigeninB	$C_{17}H_{14}O_7$	86849-77-6	2.17	[52]
52	Puerarone	$C_{20}H_{16}O_5$	116107-15-4	5.09	[53]
53	Parvisoflavanone	$C_{17}H_{16}O_7$	49776-79-6	2.87	[54]
54	Calycosin	$C_{16}H_{12}O_5$	20575-57-9	2.41	[55]
55	Maximaisoflavone J	$C_{21}H_{20}O_4$	16277-87-5	5.32	[56]
56	Artocarpanone	$C_{16}H_{14}O_{6}$	520-25-2	2.65	[57]
57	Piscerythrone	$C_{21}H_{20}O_7$	6506-96-3	4.76	[58]
58	Dihydrodaidzein	$C_{15}H_{12}O_4$	17238-05-0	2.79	[59]

PLR, Pueraria Lobatae Radix.

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Figure 2 Chemical structures of major isoflavones in Pueraria Lobatae Radix

Quantitative analysis

Currently, high performance liquid chromatography and liquid chromatography coupled with mass spectrometry techniques have been utilized to determine the contents of isoflavones in PLR. Ultra-high performance liquid chromatography has been used for the simultaneous quantification of major isoflavones in PLR within 10 minutes [19]. To increase the extraction efficiency of isoflavones, subcritical water extraction has been used to extract the isoflavones from PLR, which was a greener and more sustainable extraction approach [60]. Moreover, proton nuclear magnetic resonance was used to effectively quantify seven isoflavones, providing a reliable method for the standardization of these isoflavones in PLR [61].

Biosynthesis

Roots and stems are the major sites of isoflavone biosynthesis in PLR [62]. Isoflavone biosynthesis is a branch of phenylpropanoid metabolism that occurs through the hybridization pathway of phenylpropanoid and isoflavones [63]. The 2-hydroxylation of the C ring of flavanones could be catalyzed by 2-HIS, and the products were then catalyzed by 2-HID to yield isoflavone products, such as daidzein or genistein [23]. 2-HIS is a membrane-associated cytochrome P450 enzyme belonging to the CYP93C subfamily, and it is the first key enzyme catalyzing the synthesis of isoflavones [24]. Puerarin is biosynthesized by hydroxylation at the C-2 position via the phenylpropanoid pathway, generating its isoflavone backbone [64]. Transcription factors play an important role in the biosynthesis of secondary metabolites [63]. The regulation of transcription factors of flavonoids has been extensively studied in many plant species, while the role of isoflavone biosynthesis in PLR is seldom reported [64]. Shen et al. found that the transcript levels of the PlMYB1. PlHLH3-4 and PlWD40-1 genes were closely related to the isoflavone biosynthesis in different tissues of PLR [24, 64]. The biosynthesis pathway of isoflavone in PLR was shown in Figure 3.

Pharmacological studies

Nervous system disease

Isoflavones in PLR, especially puerarin, play a neuroprotective role by inhibiting oxidative stress, cytotoxicity, and apoptosis [65]. The neuroprotective effects of PLR isoflavones were shown in Table 2. Transient receptor potential melastatin-related 2 is an ion channel that regulates pyramidal neuron death in the CA1 region of the hippocampus. The TRPM2/NMDAR pathway was blocked in neurons of bilateral common carotid artery occlusion rats treated with puerarin, preventing the overproduction of reactive oxygen species [66]. Puerarin reduced the blood-brain barrier damage, and the possible mechanism was the inhibition of the NLRP3/Caspase-1/GSDMD-mediated classical pyroptosis pathway [67]. Moreover, rhodopsin in PLR could activate the BDNF-TRK pathway and increase neuronal cell viability and proliferation [30]. Isoflavones in PLR could exert antidepressant effects through increasing protein expression of AKT1 and FOS, and decreasing protein expression of CASP3, STAT3 and TNF-a [68]. The antidepressant effects observed in the treatment of ovariectomy mice with puerarin may be related to the inhibition of hypothalamic-pituitary-adrenal axis or the up-regulation of BDNF mRNA expression in the hippocampus [69]. Mechanisms of isoflavones in nervous system have been summarized in Figure 4.

Cardiovascular system

Numerous evidences indicated that isoflavones in PLR had excellent effects in cardiovascular system. Puerarin promoted cardiac function and prevented myocardial infarction by regulating the PPAR- γ /NF- κ B and Akt/HO-1 pathways, thereby decreasing the cardiovascular risk Furthermore, [1. 31. puerarin protected from ischemia/reperfusion-induced myocardial injury by enhancing VEGFA and Ang-1, so as to reverse cardiovascular fibrosis through the Nrf2/ROS pathway [70]. Iron death has been discovered as a new cell death mechanism of the failing heart, and some studies have demonstrated that puerarin could counter iron death caused by cardiovascular diseases [5]. Aside from puerarin, other isoflavones derived from PLR also played a crucial role in the cardiovascular system. Daidzein had anti-inflammatory and antioxidant properties, which could maintain the vascular homeostatic state [7]. Mirificin could diminish oxidative injury to cardiovascular tissues, it has potential to reduce blood pressure and enhance the heart's function

[71]. The cardiovascular effects of isoflavones in PLR were summarized in Table 3.

Isoflavone in PLR could exert anti-inflammation, anti-oxidation, anti-apoptotic, and inhibiting iron death effects through regulating a variety of important cardiac vasculature pathways, thus enhancing heart function. Mechanisms of PLR isoflavones in cardiovascular system were shown in Figure 5.

Intestinal tract diseases

Recent studies have shown that the isoflavones in PLR showed benefits for intestinal function and microecology. Isoflavones exerted their effects on the gut diseases through diverse mechanisms. The effects of isoflavones in PLR against intestinal tract diseases were listed in Table 4. A few PLR isoflavones exerted significant effects on gut metabolism, such as puerarin, genistein, and daidzin.



Figure 3 The biosynthesis pathway of isoflavone in PLR. PLR, *Pueraria Lobatae Radix*; CHS, Chalcone Synthase; CHR, Chalcone Reductase; CHI, Chalcone Isomerase; OGT, O-GlcNAc transferase.

Component	Dose	Animal models	Effects	Mechanism	Ref
Puerarin	20, 40 and 80 mg/kg	Early brain injury rat model	Reducing neurological dysfunction, and oxidative stress injury	Regulating AMPK/PGC1α/Nrf2- signaling pathway	[72]
Daidzein	50 mg/kg	High-fat diet rat model	Increasing SGZ cell proliferation and educing hippocampal apoptosis and gliosis	Regulating caspase 3, FosB, GFAP and Iba1	[73]
Daidzein	10 and 20 μΜ	LPS induced BV2 cells model	Exerting neuroprotective effects	Preventing against mitochondrial oxidative stress	[74]
Daidzein	200 mg/kg	High-fat diet rat model	Neuroprotective effects of DZ in the cerebellar layers	Activating TrkB signaling event	[75]
Formononetin	30 mg/kg	Male Sprague Dawley rats	Alleviating the neurological deficit and the pathological state of brain tissues and reducing the volume of cerebral infarction	Regulating JAK2/STAT3 signaling pathway	[76]
Formononetin	10, 30 mg/kg	Traumatic brain injury rat model	Exerting the neuroprotective and antioxidant effects against TBI	Regulating Nrf2-dependent antioxidant pathway	[77]
Formononetin	25 mg/kg	Chronic inflammatory pain mouse model	Anxiolytic effect	Inhibiting microglia activation by NF-кВ p65 signaling pathway	[78]
Genistein	15 and 30g/kg	Ovariectomized rat model	Reducing the neural apoptosis	Attenuating oxidative stress, lipid peroxidation and the mitochondria-mediated apoptotic pathway	[79]
Daidzin	1, 5, 10 mg/kg	Pentylenetetrazole-ind uced mice model	Antioxidant and anti-epileptic properties	Inhibiting VEGF signaling pathway	[80]
Tectorigenin	5, 10 mg/kg	LPS induced BV2 cells model	Exerting anti-neuroinflammatory activity	Suppressing NF-ĸB/ERK/JNK- related signaling pathways	[81]
Tectorigenin	25, 50 and 100 μM	Rat C6 astroglioma cells	Inhibiting oxidative stress	Regulating HO-1/NQO1 signaling pathways	[82]

Table 2 The neuro	protective effect	s of maior iso	flavones in P	ueraria Lol	batae Radix
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Figure 4 Mechanisms of major isoflavones actions in nervous system. JAK, janus kinase; STAT, recombinant signal transducer and activator of transcription 1; IRF, interferon regulatory factor; ERK, extracellular regulated protein kinases; JNK, c-Jun N-terminal kinase; p38, mitogen-activated protein kinase; NF-κB, nuclear factor kappa-B; NLRP3, NOD-like receptor thermal protein domain associated protein 3; CASP1, caspase1; GSDMD, gasdermin D; HO-1, heme oxygenase-1; NQO1, NAD(P)H quinone oxidoreductase 1; Nrf2, nuclear factor erythroid-2-related factor 2; Keap1, kelch-like ECH-associated protein 1; ER, estrogen receptor; ERE, estrogen response element; VEGF, vascular endothelial growth factor; HIF-1α, hypoxia inducible factor-1; TRPM2, transient receptor potential melastatin 2; BCL-2, B-cell lymphoma-2; SMAC, second mitochondria-derived activator of caspases; BAK, recombinant Bcl2 antagonist/killer; ADPR, ADP-ribosylation; PRAP, proline-rich acidic protein; PARG, eukaryotic poly ADP ribose glycohydrolase. The figure was created by Figdraw (www.figdraw.com).

Component	Dose	Effects	Mechanism	Ref
Puerarin	100 mg/kg	Protecting against sepsis-induced myocardial injury	Exerting cardioprotective effects by attenuating inflammation and oxidative damage	[83]
Puerarin	60 mg/kg	Having a promising potential for treating chronic heart failure	Increasing the expression of PPAR α and its downstream target genes GLUT4 and CD36	[84]
Puerarin	50, 100 and 150 mg/kg	Reducing myocardial fibrosis, inhibiting mitochondrial damage and improving myocardial contractile function	Inhibiting the activation of p38MAPK and its downstream activation by Na $^+/{\rm H}^+$ exchange isoform 1	[85]
Daidzein	5, 10 mg/kg	Attenuating endothelium-intact aortas	Regarding oxidative stress markers, daidzein treatment attenuated the increased malondialdehyde content and reduced activity of superoxide dismutase	[86]
Daidzein	10 mg/kg	Treating doxorubicin-induced heart failure	Ameliorating cardiac inflammation and fibrosis, cardiac apoptosis, oxidative stress and cardiac energy imbalance	[87]
Daidzein	10 µM to 100 µM	Improving myocardial infarction-induced cardiac dysfunction and cardiac fibrosis	Reducing TGF- β 1-induced cardiac fibroblast activation by regulating the TGF- β 1/SMAD2/3 signaling pathway	[88]
Formononetin	10 mg/kg	Attenuating the development of atherosclerosis	Regulating the interplay between KLF4 and SRA	[89]
Formononetin	10, 30 and 100 µmol	Decreasing the arterial pressure	Regulating NO release and Ca ²⁺ channels	[90]
Formononetin	0.5, 1, 2, 5, 8 and 10 μM	Exhibiting a protective effect on HUVECs	Increasing vascular endothelial growth factor and p-ERK1/2 expression levels	[91]
Genistein	1 mg/kg	Rescuing pulmonary vascular remodeling	Downregulating the estrogen receptor- β expression	[92]
Genistein	5 and 25 mg/kg	Attenuating myocardial fibrosis in type 1 diabetic rats	Inhibiting the TGF- β 1/Smad3 signaling pathway and regulating collagen expression in dilated cardiomyopathy	[93]
Daidzin	60 µM	High glucose-induced cardiomyocyte injury	Inhibiting the activities of ALDH2	[94]
Tectorigenin	0.1, 0.2, 0.5, 1 and 10 μmol/L,	$\begin{array}{llllllllllllllllllllllllllllllllllll$	Activating the PI3K/Akt pathway	[95]
Tectorigenin	50 mg/kg	Improving cardiac fibrosis and cardiac function in diabetic cardiomyopathy mice	Upregulating the phosphorylation of adenosine 5'-monophosphate-AMPK by preventing the ubiquitination of AdipoR1	[96]

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Figture 5 Mechanisms of PLR isoflavones in cardiovascular system. PPAR, peroxisome proliferator-activated receptors; IGF, insulin-like growth factor; Shc, Src homology 2 domain containing; Grb2, growth factor receptor-bound protein 2; SOS, son of sevenless; RAS, rat sarcoma; MEK, mitogen-activated protein kinase; RAF, raf protein kinase; ERK-1/2, mitogen-activated protein kinase; NF-B, nuclear factor kappa-B; VEGF, vascular endothelial growth factor; GPCRs, G protein-coupled receptors; L-Arg, arginine; SMAD, mothers against DPP homolog; JNK, c-Jun N-terminal kinase; ALDH2, aldehyde dehydrogenase; TGF- β , transforming growth factor- β ; AKT, proteinkinase B; mTOR, mammalian target of rapamycin; S6K1, ribosomal protein S6 kinase beta-1; 4EPB, 4E-bind-ing protein 1; IL-1, interleukin-1 β ; IL-6, interleukin-6; TNF- α , transforming growth factor- α ; PI3K, phosphatidylinositol 3-kinase. The figure was created by Figdraw (www.figdraw.com).

Table 4 The intestinal effects of major isoflavones in Pueraria Lobatae Radix

Component	Dose	Effects	Mechanism	Reference
Puerarin	10, 50 mg/kg	Preventing the breakdown of barrier integrity	Increasing levels of tight junction proteins	[97]
Puerarin	200 mg/kg	Relieving ulcerative colitis	Increasing the levels of SCFAs	[98]
Puerarin	100 mg/kg	Reversing the impairment of the intestinal caused by influenza virus infection	Reducing the levels of TLRs and inflammatory factors in the intestines and attenuating inflammatory damage	[99]
Daidzein	12.5–200 μM	Resisting intestinal epithelial barrier injury	Suppressing the PI3K/AKT and P38 pathways	[100]
Daidzein	50 mg/kg	Protecting from intestinal ischemia–reperfusion injury	Reducing caspase-6 expression	[101]
Daidzein	5 and 10 mg/kg	Inhibiting colon cancer	Inhibiting the p-ERK/ERK and p-AKT/AKT pathways	[102]
Formononetin	25, 50 and 100 mg/kg	Suppressing gastric ulcer	Inhibiting NF-кВ signaling pathway	[103]
Formononetin	50, 250, and 500 mg/kg	Exerting gastroprotective effect	Decreasing gastric secretion volumes and increasing mucus production	[104]
Formononetin	10, 30, 50, 80, and 100 μM	Inhibiting the growth and aggressiveness of GC cells	Reducing the levels of miR-542-5p	[105]
Genistein	140 mg /kg	Preventing colon cancer	Reducing the mRNA expression of COX2, TNF, and FRAT-1	[106]
Genistein	0.01–50.00 μmol/L	Inhibiting the contractile activity of gastrointestinal smooth muscle	Activating α -adrenergic receptors, NO and cAMP pathways, and KATP channels	[107]
Genistein	100 mg/kg	Protecting against acetic acid-induced ulcerative colitis	Upregulating the INF- $\gamma/JAK1/STAT1$ and INF- γ /TLR-4/NF- κB signaling pathways	[108]
Irisolidone	20 or 50 mg/kg	Attenuating ethanol-induced gastritis	Inhibiting IL-8 secretion and NF-KB activation	[109]

Puerarin played significant roles in the gut diseases mainly through three probable mechanisms. Firstly, puerarin significantly altered the gut flora by increasing the Akkermansia muciniphila, so as to achieve anti-inflammatory and anti-obese benefits [110]. Puerarin also strengthened the intestinal barrier functions by increasing the overexpression of Muc2 and ZO-1 [111]. Furthermore, puerarin could inhibit the expression of various inflammatory cytokines in the gut such as TNF- α and IL-6, thus further strengthening the gut [112]. These findings make puerarin a potential natural adjuvant for gut pathologies prevention.

Genistein could undergo methylation and hydroxylation reactions to produce multiple active metabolites [64]. These metabolites have obvious physiological activities, such as antithrombotic and antiallergic effects. For example, genistein metabolites played essential roles in platelet aggregation and immune response [113]. Genistein-related metabolites could also influence intestine cell proliferation to improve the intestinal health via activating the estrogen-related pathway [114]. These findings have shown that the metabolic transformation of Genistein could maintaining gut health.

Daidzein enhanced the integrity of the intestinal epithelial barrier by upregulating the expression of tight junction proteins like ZO-1, occluding, and claudin-1, while also dampened the inflammatory response through inhibiting PI3K/AKT and P38 MAPK pathways [100]. Moreover, daidzein have been discovered to enhance intestinal mucosal barrier function by boosting the expression of antioxidant and anti-inflammatory factors [115]. Mechanisms of PLR isoflavones in intestinal diseases were summarized in Figure 6.

Antitumor

Various biological pathways are involved in the mechanisms of PLR isoflavones for the treatment of tumors. Isoflavones in PLR significantly suppressed the proliferation of human breast cancer cells by enhancing mitochondria-dependent and non-dependent apoptotic pathways [6]. The isoflavones in PLR also possessed potential anti-tumor activity by downregulating the levels of TNF- α and IL-6 [9]. Puerarin obviously reduced the oxidative and inflammatory

processes, thereby protecting from liver tumors [116]. 3'-methoxyneopuerarin A and 3'-methoxyneopuerarin B, two new isoflavones in PLR, exerted their antitumor effects through activating the apoptotic pathway, suppressing the inflammatory responses and oxidative stress [31]. Thus, these studies indicated that isoflavones in PLR had the potential application in cancer treatment.

Diabetes

The therapeutic effects of PLR isoflavones on diabetes were involved with several molecular mechanisms, including the promotion of pancreatic β -cell division and the reduction of cell apoptosis, leading to the normal insulin secretion. The treatment effects were closely related to the activation of cAMP/PKA signaling pathway, which could increase cellular response to insulin [117]. Isoflavones in PLR also prevented diabetes-induced inflammation and oxidative stress through anti-inflammatory and antioxidant activities, thereby protecting the pancreas islet cells [118]. Moreover, these isoflavones could enhance gut health and relieve from diabetes through the modulation of gut microbial composition [119]. These multi-mechanisms were geared towards the potential value of PLR isoflavones in diabetes.

Liver diseases

In recent times, isoflavones in PLR have exhibited favorable pharmacological effects in liver ailments by influencing metabolic and inflammatory pathways. Isoflavones in PLR, such as puerarin and daidzin, could effectively regulate lipid and alcohol metabolism, diminish lipid buildup and inflammation by activating AMP/AMPK pathways, thereby mitigating liver damage [120]. The anti-diabetic properties of these isoflavones also contributed to their efficacy in treating non-alcoholic fatty liver disease. Lots of isoflavones in PLR (puerarin, daidzein, genistein, and daidzein, etc.) exerted the obvious liver-protection effects by inhibiting oxidative stress and inflammatory response, as well as regulating liver function and lipid metabolism [121]. These findings heightened the potential of PLR isoflavones for the developments of novel hepatoprotective drug.



Figure 6 Mechanisms of major isoflavones actions in intestinal tract. Muc2, Mucin 2; ZO-1, zonula occludens-1; TNF-α, transforming growth factor-α; IL-6, interleukin-6; PI3K, phosphatidylinositol 3-kinase; AKT, proteinkinase B; mTOR, mammalian target of rapamycin; S6K1, ribosomal protein S6 kinase beta-1; 4EPB, 4E-bind-ing protein 1; NF-κB, nuclear factor kappa-B; IL-1β, interleukin-1β; ERK, extracellular regulated protein kinase; JNK, c-Jun N-terminal kinase; p38, mitogen-activated protein kinase; IKB, recombinant inhibitory subunit of NF Kappa B Delta; JAK, januskinase; STAT, signal transducer and activator of transcription; IRF-9, recombinant interferon regulatory factor 9; ROS, reactive oxygen species; TLR4, toll-like receptor 4. The figure was created by Figdraw (www.figdraw.com).

Other diseases

Isoflavones in PLR, such as calycosin and daidzein, could suppress the activity of tyrosinase (a key enzyme in the synthesis of melanin), thus reducing skin aging and pigmentation [56]. These isoflavones prevented ocular diseases (particularly macular degeneration) through protecting retinal cells [122]. Puerarin has been demonstrated to prevent osteoarthritis development through anti-inflammatory effects [123].

Negative reaction/evaluation of safety

Although isoflavones in PLR showed potential in treating various diseases, they also caused some adverse reactions, especially for specific populations. These isoflavones in PLR may cause gastrointestinal discomfort [61], liver toxicity [116], and drugs interactions [124], affecting the efficacy and safety of medications. Moreover, due to the phytoestrogen properties, PLR isoflavones at high doses might influence hormonal balance and reproductive health [125]. For individuals with kidney issues, high doses of these isoflavones may exacerbate renal stress. Therefore, it was important to consider these potential risks when using PLR isoflavones, and they should be used under professional medical guidance.

Drug delivery systems

The drug delivery system for PLR isoflavones has significantly promoted their bioavailability and therapeutic efficacy. The glycosylation modification was critical for improving the solubility and absorption of the isoflavones [126]. It also ensured the easier solubility of the isoflavones and more efficient intestinal uptake [127]. Furthermore, the role of the gut microbiota in the system is significant. The isoflavones could be metabolized into more active or easier to absorb forms, thus enhancing pharmacological effects [60]. The above modifications could enhance bioavailability and reduce toxicity of PLR isoflavones.

Pharmacokinetics in isoflavones

The PLR isoflavones, especially puerarin, exhibit widespread tissue distribution. After intravenous injection of puerarin, it has been detected in multiple organs, including the brain, heart, stomach, lungs, intestines, and spleen [128]. Puerarin can cross the blood-brain barrier and distribute in various regions of the brain, such as the hippocampus, cerebral cortex, and striatum, where it exerts neuroprotective effects [129]. The brain penetration index (AUC_{brain}/AUC_{plasma}) for puerarin, 3'-Methoxypuerarin, daidzein, and daidzein-8-C-apiosyl-glycoside were 9.29%, 7.25%, 11.96%, and 4.21%, respectively [130]. Compared to other isoflavones, puerarin can relatively quickly cross the brain and exert its effects. Moreover, puerarin can also cross the placental barrier and maintain high concentrations in fetal plasma [131]. After oral administration, puerarin reached a peak concentration (Cmax) in the blood of approximately 140-230 μ g/L within 1 hour, with an absolute oral bioavailability of 7% [132]. Glucuronidation related metabolites (puerarin-7-O-glucuronide, puerarin-4'-O-glucuronide, etc.) are the major metabolites of puerarin,), and these metabolites could be easily excreted through urine and feces [133]. Although these isoflavones can cross the blood-brain barrier, their distribution in the brain is quite limited.

Discussion and future perspectives

The isoflavones in PLR have been widely studied for their potential benefits in treating cardiovascular diseases, diabetes, liver diseases, neurological disorders. Up to date, more than 100 isoflavones have been identified from PLR. Among the various parts of PLR, the roots contained the highest concentration of puerarin, while the leaves had relatively high levels of irisolidone and daidzein. Moreover, the isoflavones contents in PLR was significantly influenced by its growing environment. Isoflavones are a type of plant estrogen, primarily composed of glycosidic compounds with a 3-phenylchroman structure. This structure possessed a fundamental $C_6-C_3-C_6$ skeleton that combined with glycone or glycoside to form phenolic compounds. PLR contained a variety of active ingredients, brought great challenges for the qualitative and quantitative analysis of isoflavones in PLR. Modern chromatographic techniques (ultra-high performance liquid chromatography, ultra-high performance liquid chromatography-mass spectrometry, etc.) have significantly improved the quantitative analysis of isoflavones, providing greater accuracy and efficiency in measuring the concentration of these bioactive compounds.

Isoflavones in PLR had multiple effects (neuroprotection, anti-tumor, heart-protection, etc.) by inhibiting oxidative stress, inflammation and apoptosis. Puerarin prevented myocardial infarction by regulating the PPAR- γ /NF- κ B and Akt/HO-1 pathways. Daidzein played an essential role in reducing blood pressure and protecting heart. Daidzein was considered a powerful isoflavone due to its excellent anti-inflammatory and antioxidant properties, which played a crucial role in cardiovascular protection by reducing oxidative stress and inflammation. Isoflavones also influenced the gut through various mechanisms, such as promoting β -cell neogenesis and regulating the short-chain fatty acid metabolism. Taken daidzein as an example, it could upregulate the expression of tight junction proteins to mitigate intestinal dysfunction and improve intestinal epithelial cells.

Remarkably, the isoflavones in PLR have notable pharmacological effects on liver diseases, offering good therapeutic prospects. Puerarin and other isoflavones could upregulate AMP-activated protein to reduce liver damage and protect liver from alcohol-induced damage. Isoflavones like puerarin, daidzein, genistein, and daidzein played a crucial role in protecting the liver by regulating liver function and lipid metabolism. As isoflavones in PLR had the similar structure of estrogen, they may lead to some adverse reactions (especially at the high dose of isoflavones in PLR), such as gastrointestinal discomfort, liver toxicity and kidney toxicity. Thus, further in vivo and in vitro experiments should be conducted to reduce the side effects from PLR isoflavones. Research is needed to explore the most effective drug formulations through developing new drug delivery systems.

Most PLR isoflavones studies focused on puerarin, daidzein, and genistein, the other isoflavones' pharmacological potential should also be explored. Through these studies, the researchers could discover new isoflavones for the treatment of diseases. The pharmacological activity of isoflavones in PLR varied in different labs. PLR isoflavones possessed considerable pharmacological actions in fundamental researches, randomized, double-blind, placebo-controlled studies of these active isoflavones should be further explored.

Conclusion

Recently, more than 200 compounds have been identified from PLR, including isoflavones, terpenes, steroids, and coumarins, among which isoflavones are the predominant type. The metabolism of phenylalanine affected the formation of isoflavones. These isoflavones possess a wide range of pharmacological activities and had potential therapeutic effects on diseases in the nervous system, circulatory system, digestive system, endocrine system, and others. Further studies should focus on the medicine modifications and safety evaluation of PLR isoflavones to promote the drug development of PLR isoflavones related products.

References

- Li X, Yuan T, Chen D, et al. Cardioprotective Effects of Puerarin-V on Isoproterenol-Induced Myocardial Infarction Mice Is Associated with Regulation of PPAR-^T/NF-κB Pathway. *Molecules*. 2018;23(12):3322. Available at: http://doi.org/10.3390/molecules23123322
- 2. Cai S-A, Hou N, Zhao G-J, et al. Nrf2 Is a Key Regulator on Puerarin Preventing Cardiac Fibrosis and Upregulating

Metabolic Enzymes UGT1A1 in Rats. *Front Pharmacol.* 2018;9:540. Available at:

http://doi.org/10.3389/fphar.2018.00540

- Yen P-T, Huang S-E, Hsu J-H, et al. Anti-Inflammatory and Anti-oxidative Effects of Puerarin in Postmenopausal Cardioprotection: Roles of Akt and Heme Oxygenase-1. *Am J Chin Med.* 2022;51(01):149–168. Available at: http://doi.org/10.1142/s0192415x2350009x
- 4. Tang T, Hu X, Liao D, Liu X, Xiang D. Mechanisms of microemulsion enhancing the oral bioavailability of puerarin: comparison between oil-in-water and water-in-oil microemulsions using the single-pass intestinal perfusion method and a chylomicron flow blocking approach. *Int J Nanomedicine*. 2013;8:4415–4426. Available at: http://doi.org/10.2147/ijn.S51469
- Liu B, Zhao C, Li H, Chen X, Ding Y, Xu S. Puerarin protects against heart failure induced by pressure overload through mitigation of ferroptosis. *Biochem Biophys Res Commun.* 2018;497(1):233–240. Available at: http://doi.org/10.1016/j.bbrc.2018.02.061
- Ahn S-Y, Jo MS, Lee D, et al. Dual effects of isoflavonoids from Pueraria lobata roots on estrogenic activity and anti-proliferation of MCF-7 human breast carcinoma cells. *Bioorg Chem.* 2019;83:135–144. Available at: http://doi.org/10.1016/j.bioorg.2018.10.017
- Xuan T, Liu Y, Liu R, et al. Advances in Extraction, Purification, and Analysis Techniques of the Main Components of Kudzu Root: A Comprehensive Review. *Molecules*. 2023;28(18):6577. Available at:

http://doi.org/10.3390/molecules28186577

- Ma Y, Shang Y, Zhong Z, et al. A new isoflavone glycoside from flowers of *Pueraria Montana var. lobata* (Willd.) Sanjappa & Pradeep. *Nat Prod Res.* 2021;35(9):1459–1464. Available at: https://doi.org/10.1080/14786419.2019.1655021
- Hu Q, Xiang H, Shan J, et al. Two pairs of diastereoisomeric isoflavone glucosides from the roots of Pueraria lobata. *Fitoterapia*. 2020;144:104594. Available at: http://doi.org/10.1016/j.fitote.2020.104594
- Lv Y-Q, Tan T-W. Modeling and prediction of the mixed-mode retention mechanisms for puerarin and its analogues on n-octylamine modified poly(glycidyl methacrylate-co-ethylene glycol dimethacrylate) monoliths. *Process Biochem*. 2009;44(11):1225–1230. Available at: http://doi.org/10.1016/j.procbio.2009.06.021
- Adolfo LM, Rao X, Dixon RA. Identification of Pueraria spp. through DNA barcoding and comparative transcriptomics. *BMC Plant Biol.* 2022;22(1):10. Available at: https://doi.org/10.1186/s12870-021-03383-x
- Yang Y, Zhao H, Zhu F, et al. Analysis of Isoflavones in Pueraria by UHPLC-Q-Orbitrap HRMS and Study on α-Glucosidase Inhibitory Activity. *Foods.* 2022;11(21):3523. Available at: http://doi.org/10.3390/foods11213523
- Cheng Y, Yu S, Xue F. Comparison of the Extraction Efficiency of Isoflavone Compounds from Puerariae lobatae by Ionic Liquids with 11 Anions and 8 Imidazolium-Based Cations. ACS Omega. 2020;5(15):8962–8971. Available at: http://doi.org/10.1021/acsomega.0c00724
- Fang X, Zhang Y, Cao Y, et al. Studies on Chemical Composition of Pueraria lobata and Its Anti-Tumor Mechanism. *Molecules*. 2022;27(21):7253. Available at: http://doi.org/10.3390/molecules27217253
- Hamed AB, El-Abhar HS, Abdallah DM, Ahmed KA, Abulfadl YS. Prunetin in a GPR30-dependent manner mitigates renal ischemia/reperfusion injury in rats via interrupting indoxyl sulfate/TLR4/TRIF, RIPK1/RIPK3/MLKL, and RIPK3/PGAM5/DRP-1 crosstalk. Saudi Pharm J. 2023;31(11):101818. Available at: http://doi.org/10.1016/j.jsps.2023.101818
- 16. Zhang Y, Xu D, Xing X, Yang H, Gao W, Li P. The chemistry and

activity-oriented characterization of isoflavones difference between roots of Pueraria lobata and P. thomsonii guided by feature-based molecular networking. *Food Chem.* 2023;422:136198. Available at:

https://doi.org/10.1016/j.foodchem.2023.136198

- Shao X-Y, He T, Lai Y-L, Chen M, Tong Z-H. Water-Soluble Polysaccharides Extracted from Pueraria lobata Delay Aging of Caenorhabditis elegans under Heat Stress. *Plant Foods Hum Nutr.* 2022;77(2):220–225. Available at: http://doi.org/10.1007/s11130-022-00964-5
- Shin J, Bae E-A, Lee YC, Ma J-Y, Kim D-H. Estrogenic Effect of Main Components Kakkalide and Tectoridin of Puerariae Flos and Their Metabolites. *Biol Pharm Bull.* 2006;29(6):1202–1206. Available at:
- http://doi.org/10.1248/bpb.29.1202 19. Fu M, Jahan MS, Tang K, et al. Comparative analysis of the
- medicinal and nutritional components of different varieties of Pueraria thomsonii and Pueraria lobata. *Front Plant Sci.* 2023;14:1115782. Available at: http://doi.org/10.3389/fpls.2023.1115782
- Zhu J, Cheng H, Zhou M, Li S, Tang T, Feng J. Determining three isoflavones from Pueraria lobata using magnetic ZIF-8 nanoparticle-based solid-phase extraction and pressurized capillary electrochromatography. *J Pharm Biomed Anal.* 2022;212:114592. Available at: http://doi.org/10.1016/j.jpba.2022.114592
- Wang Q, Cheng X-L, Li H, et al. Application of an efficient strategy for discovery and purification of bioactive compounds from Chinese herbal medicines, a case study on the Puerariae thomsonii Flos. *J Pharm Biomed Anal.* 2013;75:25–32. Available at:

http://doi.org/10.1016/j.jpba.2012.11.009

- Li C, Zhang Y. Glycosylation and methylation in the biosynthesis of isoflavonoids in Pueraria lobata. *Front Plant Sci.* 2023,14:1330586. Available at: http://doi.org/10.3389/fpls.2023.1330586
- Gao HY, Liu Y, Tan FF, Zhu LW, Jia KZ, Tang YJ. The Advances and Challenges in Enzymatic C-glycosylation of Flavonoids in Plants. *Curr Pharm Des.* 2022;28(18):1466–1479. Available at: http://doi.org/10.2174/1381612828666220422085128
- 24. Wang X, Pan H, Sagurthi S, Paris V, Zhuo C, Dixon RA. The protein conformational basis of isoflavone biosynthesis. *Commun Biol.* 2022;5(1):1249. Available at: http://doi.org/10.1038/s42003-022-04222-x
- Liu L, Ma Y, Chen X, Xiong X, Shi S. Screening and identification of BSA bound ligands from Puerariae lobata flower by BSA functionalized Fe₃O₄ magnetic nanoparticles coupled with HPLC–MS/MS. *J Chromatogr B*. 2012;887–888:55–60. Available at:

http://doi.org/10.1016/j.jchromb.2012.01.008

- Li J, Li C, Gou J, Wang X, Fan R, Zhang Y. An Alternative Pathway for Formononetin Biosynthesis in Pueraria lobata. *Front Plant Sci.* 2016;7:861. Available at: http://doi.org/10.3389/fpls.2016.00861
- Shukla R, Banerjee S, Tripathi YB. Antioxidant and Antiapoptotic effect of aqueous extract of Pueraria tuberosa (Roxb. Ex Willd.) DC. On streptozotocin-induced diabetic nephropathy in rats. *BMC Complement Altern Med.* 2018;18(1):156. Available at: https://doi.org/10.1186/s12906-018-2221-x
- 28. Xu Z, Talpur ZH, Yang W, et al. Dual-spectrum online monitoring of puerarin and total flavonoids contents during the extraction process of Pueraria lobata. *Talanta*. 2022;248:123608. Available at:

http://doi.org/10.1016/j.talanta.2022.123608

29. Wang X, Zhang Q, Jiang H, et al. Pueraria lobata leaf extract as green corrosion inhibitor for low carbon steel in 1.0 M HCl solution. *Res Chem Intermed* 2021;47(3):1051–1069. Available at: https://doi.org/10.1007/s11164-020-04316-3

30. Kavtaradze NSh, Alaniya MD, Getia MZ, et al. Isoflavonoids from Roots of Pueraria hirsuta Distributed in Georgia and their Hepatoprotective Activity. *Chem Nat Compd.* 2022;58(1):27–31. Available at:

http://doi.org/10.1007/s10600-022-03634-9

- Sun Y, Zhang H, Cheng M, et al. New hepatoprotective isoflavone glucosides from *Pueraria lobata* (Willd.) Ohwi. *Nat Prod Res.* 2019;33(24):3485–3492. Available at: https://doi.org/10.1080/14786419.2018.1484461
- Liga S, Paul C. Puerarin—A Promising Flavonoid: Biosynthesis, Extraction Methods, Analytical Techniques, and Biological Effects. Int J Mol Sci. 2024;25(10):5222. Available at: http://doi.org/10.3390/ijms25105222
- 33. Sharma A, Wairkar S. Flavonoids for treating pulmonary fibrosis: Present status and future prospects. *Phytother Res.* 2024;38(9):4406–4423. Available at: http://doi.org/10.1002/ptr.8285
- 34. Adolfo LM, Burks D, Rao X, Alvarez-Hernandez A, Dixon RA. Evaluation of pathways to the C-glycosyl isoflavone puerarin in roots of kudzu (*Pueraria montana lobata*). *Plant Direct.* 2022;6(9):e442. Available at: http://doi.org/10.1002/pld3.442
- 35. He Y-X, Liu M-N, Wu H, et al. Puerarin: a hepatoprotective drug from bench to bedside. *Chin Med.* 2024;19(1):139. Available at: http://doi.org/10.1186/s13020-024-01011-y
- 36. Qin W, Guo J, Gou W, et al. Molecular mechanisms of isoflavone puerarin against cardiovascular diseases: What we know and where we go. *Chin Herb Med.* 2022;14(2):234–243. Available at: http://doi.org/10.1016/j.chmed.2021.12.003
- 37. Guerra-Ordaz AA, González-Ortiz G, La Ragione RM, et al. Lactulose and Lactobacillus plantarum, a potential complementary synbiotic to control postweaning colibacillosis in piglets. *Appl Environ Microbiol*. 2014;80(16):4879–4886. Available at:

https://doi.org/10.1128/AEM.00770-14

- Zhang C-L, Ding X-P, Hu Z-F, et al. Comparative Study of Puerariae lobatae and Puerariae thomsonii by HPLC-Diode Array Detection-Flow Injection-Chemiluminescence Coupled with HPLC-Electrospray Ionization-MS. *Chem Pharm Bull.* 2011;59(5):541–545. Available at: http://doi.org/10.1248/cpb.59.541
- 39. Chen Y-G, Song Y-L, Wang Y, et al. Metabolic differentiations of Pueraria lobata and Pueraria thomsonii using 1H NMR spectroscopy and multivariate statistical analysis. J Pharm Biomed Anal. 2014;93:51–58. Available at: http://doi.org/10.1016/j.jpba.2013.05.017
- Nguyen VD, Min BC, Kyung MO, et al. Identification of a naturally-occurring 8-[alpha-D-glucopyranosyl-(1-->6)-beta-Dglucopyranosyl]daidzein from cultivated kudzu root. *Phytochem Anal.* 2009;20(6):450–455. Available at: http://doi.org/10.1002/pca.1146
- Chen I-C, Chang K-H, Chen Y-J, Chen Y-C, Lee-Chen G-J, Chen C-M. Pueraria lobata and Daidzein Reduce Cytotoxicity by Enhancing Ubiquitin-Proteasome System Function in SCA3-iPSC-Derived Neurons. Oxid Med Cell Longev. 2019;2019:1–18. Available at: http://doi.org/10.1155/2019/8130481
- 42. Liang J, Maeda T, Tao X, Wu Y, Tang H. Physicochemical Properties of Pueraria Root Starches and Their Effect on the Improvement of Buckwheat Noodle Quality. *Cereal Chem.* 2017;94(3):554–559. Available at: http://doi.org/10.1094/CCHEM-08-16-0219-R
- Yao M, Liao Y, Li GQ, Law FC, Tang Y. Quantitative analysis of two isoflavones in Pueraria lobata flowers from eleven Chinese provinces using high performance liquid chromatography. *Chin Med.* 2010;5(1):14. Available at: http://doi.org/10.1186/1749-8546-5-14
- 44. Hirayama K, Matsuzuka Y, Kamiya T, Ikeguchi M, Takagaki K,

Itoh K. Metabolism of Isoflavones Found in the Pueraria thomsonii Flower by Human Intestinal Microbiota. *Biosci Microflora*. 2011;30(4):135–140. Available at: http://doi.org/10.12938/bifidus.30.135

- Fatima A, Khan MS, Ahmad MdW. Therapeutic Potential of Equol: A Comprehensive Review. *Curr Pharm Des.* 2020;26(45):5837–5843. Available at: http://doi.org/10.2174/1381612826999201117122915
- 46. Li Z, Wang Y, Zeng G, et al. Increased miR-155 and heme oxygenase-1 expression is involved in the protective effects of formononetin in traumatic brain injury in rats. *Am J Transl Res.* 2017;9(12):5653–5661. Available at: https://pubmed.ncbi.nlm.nih.gov/29312517/
- Kapingu M, Mbwambo Z, Moshi M, et al. A Novel Isoflavonoid from Millettia puguensis. *Planta Med.* 2006;72(14):1341–1343. Available at: http://doi.org/10.1055/s-2006-951689
- Bai X, Xie Y, Liu J, Qu J, Kano Y, Yuan D. Isolation and Identification of Urinary Metabolites of Kakkalide in Rats. *Drug Metab Dispos*. 2009;38(2):281–286. Available at: http://doi.org/10.1124/dmd.109.028555
- Delmonte P, Perry J, Rader JI. Determination of isoflavones in dietary supplements containing soy, Red Clover and kudzu: Extraction followed by basic or acid hydrolysis. *J Chromatogr A*. 2006;1107(1–2):59–69. Available at: http://doi.org/10.1016/j.chroma.2005.11.060
- 50. Guo N, Fang Z, Zang Q, et al. Spatially resolved metabolomics combined with bioactivity analyses to evaluate the pharmacological properties of two Radix Puerariae species. *J Ethnopharmacol.* 2023;313:116546. Available at: http://doi.org/10.1016/j.jep.2023.116546
- 51. Lim H-S, Kim YJ, Kim B-Y, Park G, Jeong S-J. The Anti-neuroinflammatory Activity of Tectorigenin Pretreatment via Downregulated NF-κB and ERK/JNK Pathways in BV-2 Microglial and Microglia Inactivation in Mice With Lipopolysaccharide. *Front Pharmacol.* 2018;9:462. Available at: http://doi.org/10.3389/fphar.2018.00462
- 52. Wang L, Liu N, Hu S, et al. Application of instantaneous nebulization dispersive liquid-phase microextraction combined with HPLC for the determination of chalcone and isoflavone in traditional Chinese medicines. *J Sep Sci.* 2023;46(19):e2300326. Available at:

http://doi.org/10.1002/jssc.202300326

- 53. Xie G-Y, Zhu Y, Shu P, et al. Phenolic metabolite profiles and antioxidants assay of three Iridaceae medicinal plants for traditional Chinese medicine "She-gan" by on-line HPLC–DAD coupled with chemiluminescence (CL) and ESI-Q-TOF-MS/MS. J Pharm Biomed Anal. 2014;98:40–51. Available at: http://doi.org/10.1016/j.jpba.2014.05.008
- Sohn SI, Pandian S, Oh YJ, Kang HJ, Cho WS, Cho YS. Metabolic Engineering of Isoflavones: An Updated Overview. *Front Plant Sci.* 2021;12:670103. Available at: http://doi.org/10.3389/fpls.2021.670103
- 55. Liu X, Huang R, Wan J. Puerarin: a potential natural neuroprotective agent for neurological disorders. *Biomed Pharmacother*. 2023;162:114581. Available at: http://doi.org/10.1016/j.biopha.2023.114581
- 56. Wagle A, Seong SH, Jung HA, Choi JS. Identifying an isoflavone from the root of Pueraria lobata as a potent tyrosinase inhibitor. *Food Chem.* 2019;276:383–389. Available at: http://doi.org/10.1016/j.foodchem.2018.10.008
- 57. Deyou T, Marco M, Heydenreich M, et al. Isoflavones and Rotenoids from the Leaves of Millettia oblata ssp. teitensis. *J Nat Prod.* 2017;80(7):2060–2066. Available at: https://doi.org/10.1021/acs.jnatprod.7b00255
- Zaidi KU, Ali SA, Ali A, Naaz I. Natural Tyrosinase Inhibitors: Role of Herbals in the Treatment of Hyperpigmentary Disorders. *Mini Rev Med Chem.* 2019;19(10):796–808. Available at: http://doi.org/10.2174/1389557519666190116101039

- 59. Guo B, Xu J, Xiao M, Ding M, Duan L. Puerarin reduces ischemia/reperfusion-induced myocardial injury in diabetic rats via upregulation of vascular endothelial growth factor A/angiotensin-1 and suppression of apoptosis. *Mol Med Report.* 2018;17(5):7421–7427. Available at: http://doi.org/10.3892/mmr.2018.8754
- Kwon JE, Lee JW, Park Y, et al. Biotransformation of Pueraria lobata Extract with Lactobacillus rhamnosus vitaP1 Enhances Anti-Melanogenic Activity. J Microbiol Biotechnol. 2018;28(1):22–31. Available at: http://doi.org/10.4014/jmb.1705.05087
- Thapa P, Kim HM, Hong J-P, et al. Absolute Quantification of Isoflavones in the Flowers of Pueraria lobata by qHNMR. *Plants*. 2022;11(4):548. Available at: http://doi.org/10.3390/plants11040548
- 62. Xi H, Zhu Y, Sun W, et al. Comparative Transcriptome Analysis of Pueraria lobata Provides Candidate Genes Involved in Puerarin Biosynthesis and Its Regulation. *Biomolecules*. 2023;13(1):170. Available at: http://doi.org/10.3390/biom13010170
- 63. He M, Yao Y, Li Y, et al. Comprehensive transcriptome analysis reveals genes potentially involved in isoflavone biosynthesis in Pueraria thomsonii Benth. *PLoS One.* 2019;14(6):e0217593. Available at:

https://doi.org/10.1371/journal.pone.0217593

- 64. Cheng H, Huang X, Wu S, et al. Chromosome-Level Genome Assembly and Multi-Omics Dataset Provide Insights into Isoflavone and Puerarin Biosynthesis in *Pueraria lobata* (Wild.) Ohwi. *Biomolecules*. 2022;12(12):1731. Available at: https://doi.org/10.3390/biom12121731
- 65. Shang X, Yi X, Xiao L, et al. Chromosomal-level genome and multi-omics dataset of Pueraria lobata var. thomsonii provide new insights into legume family and the isoflavone and puerarin biosynthesis pathways. *Hortic Res.* 2022;9:uhab035. Available at:

https://doi.org/10.1093/hr/uhab035

- 66. Zhu T, Zhu M, Qiu Y, et al. Puerarin Alleviates Vascular Cognitive Impairment in Vascular Dementia Rats. Front Behav Neurosci. 2021;15:717008. Available at: http://doi.org/10.3389/fnbeh.2021.717008
- 67. Zhou S, Li Y, Hong Y, Zhong Z, Zhao M. Puerarin protects against sepsis-associated encephalopathy by inhibiting NLRP3/Caspase-1/GSDMD pyroptosis pathway and reducing blood-brain barrier damage. *Eur J Pharmacol.* 2023;945:175616. Available at:

http://doi.org/10.1016/j.ejphar.2023.175616

- Zhang N, Guo P, Zhao Y, et al. Pharmacological mechanisms of puerarin in the treatment of Parkinson's disease: An overview. *Biomed Pharmacother*. 2024;177:117101. Available at: http://doi.org/10.1016/j.biopha.2024.117101
- 69. Tantipongpiradet A, Monthakantirat O, Vipatpakpaiboon O, et al. Effects of Puerarin on the Ovariectomy-Induced Depressive-Like Behavior in ICR Mice and Its Possible Mechanism of Action. *Molecules*. 2019;24(24):4569. Available at:

http://doi.org/10.3390/molecules24244569

70. Maciejewska-Turska M, Pecio Ł, Zgórka G. Isolation of Mirificin and Other Bioactive Isoflavone Glycosides from the Kudzu Root Lyophilisate Using Centrifugal Partition and Flash Chromatographic Techniques. *Molecules*. 2022;27(19):6227. Available at:

http://doi.org/10.3390/molecules27196227

- Chauhan P, Wadhwa K, Mishra R, et al. Investigating the Potential Therapeutic Mechanisms of Puerarin in Neurological Diseases. *Mol Neurobiol.* 2024. Available at: http://doi.org/10.1007/s12035-024-04222-4
- Huang Y, Wu H, Hu Y, et al. Puerarin Attenuates Oxidative Stress and Ferroptosis via AMPK/PGC1α/Nrf2 Pathway after Subarachnoid Hemorrhage in Rats. Antioxidants.

2022;11(7):1259. Available at: http://doi.org/10.3390/antiox11071259

73. Rivera P, Pérez-Martín M, Pavón FJ, et al. Pharmacological administration of the isoflavone daidzein enhances cell proliferation and reduces high fat diet-induced apoptosis and gliosis in the rat hippocampus. *PLoS One.* 2013;8(5):e64750. Available at:

https://doi.org/10.1371/journal.pone.0064750

74. Johnson SL, Park HY, Vattem DA, Grammas P, Ma H, Seeram NP. Equol, a Blood–Brain Barrier Permeable Gut Microbial Metabolite of Dietary Isoflavone Daidzein, Exhibits Neuroprotective Effects against Neurotoxins Induced Toxicity in Human Neuroblastoma SH-SY5Y Cells and Caenorhabditis elegans. *Plant Foods Hum Nutr.* 2020;75(4):512–517. Available at:

http://doi.org/10.1007/s11130-020-00840-0

- 75. Alò R, Fazzari G, Zizza M, et al. Emotional and Spontaneous Locomotor Behaviors Related to cerebellar Daidzein-dependent TrkB Expression Changes in Obese Hamsters. *Cerebellum*. 2022;22(4):698–707. Available at: http://doi.org/10.1007/s12311-022-01432-1
- 76. Yu L, Zhang Y, Chen Q, et al. Formononetin protects against inflammation associated with cerebral ischemia-reperfusion injury in rats by targeting the JAK2/STAT3 signaling pathway. *Biomed Pharmacother*. 2022;149:112836. Available at: http://doi.org/10.1016/j.biopha.2022.112836
- 77. Li Z, Wang Y, Zeng G, et al. Increased miR-155 and heme oxygenase-1 expression is involved in the protective effects of formononetin in traumatic brain injury in rats. *Am J Transl Res.* 2017;9(12):5653–5661. Available at: https://pubmed.ncbi.nlm.nih.gov/29312517/
- Wang X, Guan S, Liu A, et al. Anxiolytic effects of Formononetin in an inflammatory pain mouse model. *Mol Brain*. 2019;12(1):36. Available at: http://doi.org/10.1186/s13041-019-0453-4
- 79. Huang Y-H, Zhang Q-H. Genistein reduced the neural apoptosis in the brain of ovariectomised rats by modulating mitochondrial oxidative stress. *Br J Nutr.* 2010;104(9):1297–1303. Available at:

http://doi.org/10.1017/s0007114510002291

 Kazmi Z, Zeeshan S, Khan A, et al. Anti-epileptic activity of daidzin in PTZ-induced mice model by targeting oxidative stress and BDNF/VEGF signaling. *Neurotoxicology*. 2020;79:150–163. Available at:

http://doi.org/10.1016/j.neuro.2020.05.005

- Rong J, Fu F, Han C, Wu Y, Xia Q, Du D. Tectorigenin: A Review of Its Sources, Pharmacology, Toxicity, and Pharmacokinetics. *Molecules*. 2023;28(15):5904. Available at: http://doi.org/10.3390/molecules28155904
- 82. Park J, Jung J, Jeong Y, et al. Antioxidant mechanism of isoflavone metabolites in hydrogen peroxide-stimulated rat primary astrocytes: critical role of hemeoxygenase-1 and NQO1 expression. J Neurochem. 2011;119(5):909–919. Available at: http://doi.org/10.1111/j.1471-4159.2011.07395.x
- Zhou B, Zhang J, Chen Y, et al. Puerarin protects against sepsis-induced myocardial injury through AMPK-mediated ferroptosis signaling. *Aging (Milano)*. 2022;14(8):3617–3632. Available at:

http://doi.org/10.18632/aging.204033

- 84. He L, Wang T, Chen B, Lu F, Xu J. Puerarin inhibits apoptosis and inflammation in myocardial cells via PPARα expression in rats with chronic heart failure. *Exp Ther Med.* 2019;18(5):3347–3356. Available at: http://doi.org/10.3892/etm.2019.7984
- 85. Pan G, Cui B, Han M, et al. Puerarin inhibits NHE1 activity by interfering with the p38 pathway and attenuates mitochondrial damage induced by myocardial calcium overload in heart failure rats. *Acta Biochim Biophys Sin.* 2024;56(2):270–279. Available at:

http://doi.org/10.3724/abbs.2023269

- 86. Roghani M, Vaez Mahdavi M, Jalali-Nadoushan M, et al. Chronic Administration of Daidzein, a Soybean Isoflavone, Improves Endothelial Dysfunction and Attenuates Oxidative Stress in Streptozotocin-induced Diabetic Rats. *Phytother Res.* 2012;27(1):112–117. Available at: http://doi.org/10.1002/ptr.4699
- Li H, Zhang M, Wang Y, et al. Daidzein alleviates doxorubicin-induced heart failure via the SIRT3/FOXO3a signaling pathway. *Food Funct.* 2022;13(18):9576–9588. Available at: http://doi.org/10.1039/d2fo00772j
- Shu J, Hu L, Wu Y, et al. Daidzein suppresses TGF-β1-induced cardiac fibroblast activation via the TGF-β1/SMAD2/3 signaling pathway. *Eur J Pharmacol.* 2022;919:174805. Available at: http://doi.org/10.1016/j.ejphar.2022.174805
- 89. Ma C, Xia R, Yang S, et al. Formononetin attenuates atherosclerosis via regulating interaction between KLF4 and SRA in apoE-/- mice. *Theranostics*. 2020;10(3):1090–1106. Available at:

http://doi.org/10.7150/thno.38115

- 90. SUN T, LIU R, CAO Y. Vasorelaxant and antihypertensive effects of formononetin through endothelium-dependent and -independent mechanisms. *Acta Pharmacol Sin.* 2011;32(8):1009–1018. Available at: http://doi.org/10.1038/aps.2011.51
- Liang C, Zhou A, Sui C, Huang Z. The effect of formononetin on the proliferation and migration of human umbilical vein endothelial cells and its mechanism. *Biomed Pharmacother*. 2019;111:86–90. Available at: http://doi.org/10.1016/j.biopha.2018.12.049
- 92. Matori H, Umar S, Nadadur RD, et al. Genistein, a Soy Phytoestrogen, Reverses Severe Pulmonary Hypertension and Prevents Right Heart Failure in Rats. *Hypertension*. 2012;60(2):425–430. Available at:

http://doi.org/10.1161/hypertensionaha.112.191445

- 93. Yang R, Jia Q, Liu X, Ma S. Effect of genistein on myocardial fibrosis in diabetic rats and its mechanism. *Mol Med Rep.* 2018;17(2):2929–2936. Available at: http://doi.org/10.3892/mmr.2017.8268
- 94. Gu X, Fang T, Kang P, et al. Effect of ALDH2 on High Glucose-Induced Cardiac Fibroblast Oxidative Stress, Apoptosis, and Fibrosis. Oxid Med Cell Longev. 2017;2017:9257967. Available at: https://doi.org/10.1155/2017/9257967
- Chen X, Zhang W, Sun L, Lian Y. Tectorigenin protect HUVECs from H2O2-induced oxidative stress injury by regulating PI3K/Akt pathway. *Tissue Cell*. 2021;68:101475. Available at: http://doi.org/10.1016/j.tice.2020.101475
- 96. Ma Y-L, Xu M, Cen X-F, Qiu H-L, Guo Y-Y, Tang Q-Z. Tectorigenin protects against cardiac fibrosis in diabetic mice heart via activating the adiponectin receptor 1-mediated AMPK pathway. *Biomed Pharmacother*. 2024;174:116589. Available at: http://doi.org/10.1016/j.biopha.2024.116589
- Jeon Y-D, Lee J-H, Lee Y-M, Kim D-K. Puerarin inhibits inflammation and oxidative stress in dextran sulfate sodium-induced colitis mice model. *Biomed Pharmacother*. 2020;124:109847. Available at: http://doi.org/10.1016/j.biopha.2020.109847
- 98. Wu Y, Li Y, Ruan Z, et al. Puerarin Rebuilding the Mucus Layer and Regulating Mucin-Utilizing Bacteria to Relieve Ulcerative Colitis. J Agric Food Chem. 2020;68(41):11402–11411. Available at:

http://doi.org/10.1021/acs.jafc.0c04119

99. Zeng M-S, Yu W-D, Wang H-X, Xu P-P, Liu J-Y. Puerarin reduces impairment of intestinal and adipose immune responses to influenza virus infection in mice. Arch Virol. 2021;166(9):2387–2397. Available at: http://doi.org/10.1007/s00705-021-05112-z 100. Zhang B, Wei X, Ding M, Luo Z, Tan X, Zheng Z. Daidzein Protects Caco-2 Cells against Lipopolysaccharide-Induced Intestinal Epithelial Barrier Injury by Suppressing PI3K/AKT and P38 Pathways. *Molecules*. 2022;27(24):8928. Available at: http://doi.org/10.3390/molecules27248928

- 101. Durgun C, Aşır F. Daidzein alleviated the pathologies in intestinal tissue against ischemia-reperfusion. Eur Rev Med Pharmacol Sci. 2023;27(4):1487–1493. Available at: http://doi.org/10.26355/eurrev_202302_31389
- 102. Salama AAA, Allam RM. Promising targets of chrysin and daidzein in colorectal cancer: Amphiregulin, CXCL1, and MMP-9. *Eur J Pharmacol.* 2021;892:173763. Available at: http://doi.org/10.1016/j.ejphar.2020.173763
- 103. Yi L, Lu Y, Yu S, Cheng Q, Yi L. Formononetin inhibits inflammation and promotes gastric mucosal angiogenesis in gastric ulcer rats through regulating NF-κB signaling pathway. J Recept Signal Transduct. 2020;42(1):16–22. Available at: http://doi.org/10.1080/10799893.2020.1837873
- 104. Mendonça MAA, Ribeiro ARS, Lima AK, et al. Red Propolis and Its Dyslipidemic Regulator Formononetin: Evaluation of Antioxidant Activity and Gastroprotective Effects in Rat Model of Gastric Ulcer. *Nutrients*. 2020;12(10):2951. Available at: https://doi.org/10.3390/nu12102951
- 105. Wang WS, Zhao CS. Formononetin exhibits anticancer activity in gastric carcinoma cell and regulating miR-542-5p. *Kaohsiung J Med Sci.* 2021;37(3):215–225. Available at: http://doi.org/10.1002/kjm2.12322
- 106. Song S, Cheng D, Wei S, et al. Preventive effect of genistein on AOM/DSS-induced colonic neoplasm by modulating the PI3K/AKT/FOXO3 signaling pathway in mice fed a high-fat diet. *J Funct Foods.* 2018;46:237–242. Available at: http://doi.org/10.1016/j.jff.2018.05.006
- 107. Zhang L-X. Resveratrol and genistein inhibition of rat isolated gastrointestinal contractions and related mechanisms. World J Gastroenterol. 2014;20(41):15335. Available at: http://doi.org/10.3748/wjg.v20.i41.15335
- 108. Elhefnawy EA, Zaki HF, El Maraghy NN, Ahmed KA, Abd El-Haleim EA. Genistein and/or sulfasalazine ameliorate acetic acid-induced ulcerative colitis in rats via modulating INF-γ/JAK1/STAT1/IRF-1, TLR-4/NF-κB/IL-6, and JAK2/STAT3/COX-2 crosstalk. *Biochem Pharmacol.* 2023;214:115673. Available at: http://doi.org/10.1016/j.bcp.2023.115673
- 109. Kang G, Lee S, Jang S, Han MJ, Kim D. Irisolidone attenuates ethanol-induced gastric injury in mice by inhibiting the infiltration of neutrophils. *Mol Nutr Food Res.* 2016;61(2):10.1002/mnfr.201600517. Available at: http://doi.org/10.1002/mnfr.201600517
- 110. Wang L, Wu Y, Zhuang L, et al. Puerarin prevents high-fat diet-induced obesity by enriching Akkermansia muciniphila in the gut microbiota of mice. *PLoS One*. 2019;14(6):e0218490. Available at: https://doi.org/10.1371/journal.pone.0218490
- 111. Li J, Zhang L, Li Y, et al. Puerarin improves intestinal barrier function through enhancing goblet cells and mucus barrier. J Funct Foods. 2020;75:104246. Available at: http://doi.org/10.1016/j.jff.2020.104246
- 112. Song X, Wang W, Ding S, Liu X, Wang Y, Ma H. Puerarin ameliorates depression-like behaviors of with chronic unpredictable mild stress mice by remodeling their gut microbiota. *J Affect Disord*. 2021;290:353–363. Available at: http://doi.org/10.1016/j.jad.2021.04.037
- 113. Choo M-K, Park E-K, Yoon H-K, Kim D-H. Antithrombotic and Antiallergic Activities of Daidzein, a Metabolite of Puerarin and Daidzin Produced by Human Intestinal Microflora. *Biol Pharm Bull.* 2002;25(10):1328–1332. Available at: http://doi.org/10.1248/bpb.25.1328
- 114. Park E-K, Shin J, Bae E-A, Lee Y-C, Kim D-H. Intestinal Bacteria Activate Estrogenic Effect of Main Constituents Puerarin and

Daidzin of Pueraria thunbergiana. *Biol Pharm Bull.* 2006;29(12):2432–2435. Available at: http://doi.org/10.1248/bpb.29.2432

115. Ou W, Hu H, Yang P, et al. Dietary daidzein improved intestinal health of juvenile turbot in terms of intestinal mucosal barrier function and intestinal microbiota. *Fish Shellfish Immunol.* 2019;94:132–141. Available at: http://doi.org/10.1016/j.fsi.2019.08.059

116. Lin B, Zhao F, Liu Y, et al. Randomized Clinical Trial: Probiotics Alleviated Oral-Gut Microbiota Dysbiosis and Thyroid Hormone Withdrawal-Related Complications in Thyroid Cancer Patients Before Radioiodine Therapy Following Thyroidectomy. Front Endocrinol. 2022;13:834674. Available at:

http://doi.org/10.3389/fendo.2022.834674
117. Gilbert ElizabethR, Liu D. Anti-diabetic functions of soy isoflavone genistein: mechanisms underlying its effects on pancreatic β-cell function. *Food Funct.* 2013;4(2):200–212. Available at:

http://doi.org/10.1039/c2fo30199g

- 118. Jheng H, Hayashi K, Matsumura Y, et al. Anti-Inflammatory and Antioxidative Properties of Isoflavones Provide Renal Protective Effects Distinct from Those of Dietary Soy Proteins against Diabetic Nephropathy. *Mol Nutr Food Res.* 2020;64(10):e2000015. Available at: http://doi.org/10.1002/mnfr.202000015
- 119. Lertpatipanpong P, Janpaijit S, Park E-Y, Kim C-T, Baek SJ. Potential Anti-Diabetic Activity of Pueraria lobata Flower (Flos Puerariae) Extracts. *Molecules*. 2020;25(17):3970. Available at: http://doi.org/10.3390/molecules25173970
- 120. Zhang X, Zheng J, Jiang N, et al. Modulation of gut microbiota and intestinal metabolites by lactulose improves loperamide-induced constipation in mice. *Eur J Pharm Sci.* 2021;158:105676. Available at: http://doi.org/10.1016/j.ejps.2020.105676
- 121. Feng R, Chen J-H, Liu C-H, et al. A combination of Pueraria lobata and Silybum marianum protects against alcoholic liver disease in mice. *Phytomedicine*. 2019;58:152824. Available at: http://doi.org/10.1016/j.phymed.2019.152824
- 122. Kang NR, Pyun B-J, Jung DH, et al. Pueraria lobata Extract Protects Hydrogen Peroxide-Induced Human Retinal Pigment Epithelial Cells Death and Membrane Permeability. *Evid Based Complement Alternat Med.* 2019;2019:1–10. Available at: http://doi.org/10.1155/2019/5710289
- 123. Ma T, Wen Y, Song X, et al. Puerarin inhibits the development of osteoarthritis through antiinflammatory and antimatrix-degrading pathways in osteoarthritis-induced rat model. *Phytother Res.* 2020;35(5):2579–2593. Available at: http://doi.org/10.1002/ptr.6988

- 124. Zhang G, Ji J, Sun M, Ji Y, Ji H. Comparative Pharmacokinetic Profiles of Puerarin in Rat Plasma by UHPLC-MS/MS after Oral Administration of Pueraria lobata Extract and Pure Puerarin. J Anal Methods Chem. 2020;2020:1–8. Available at: http://doi.org/10.1155/2020/4258156
- 125. Lippert JA, Rimmer CA, Phillips MM, et al. Development of Kudzu (Pueraria Montana var. lobata) Reference Materials for the Determination of Isoflavones and Toxic Elements. J AOAC Int. 2022;105(4):1162–1174. Available at: https://doi.org/10.1093/jaoacint/qsac023
- 126. Wang X, Li C, Zhou Z, Zhang Y. Identification of Three (Iso)flavonoid Glucosyltransferases From Pueraria lobata. Front Plant Sci. 2019;10:28. Available at: http://doi.org/10.3389/fpls.2019.00028
- 127. Chen R, Wu P, Cai Z, et al. The combination of Puerariae Lobatae Radix and Chuanxiong Rhizoma enhanced the absorption and pharmacokinetics of puerarin by modulating the intestinal barrier and influenced gut microbiota. *J Funct Foods*. 2018;47:72–82. Available at: http://doi.org/10.1016/j.jff.2018.05.043
- 128. Anukunwithaya T, Poo P, Hunsakunachai N, Rodsiri R, Malaivijitnond S, Khemawoot P. Absolute oral bioavailability and disposition kinetics of puerarin in female rats. BMC Pharmacol Toxicol. 2018;19(1):25. Available at: http://doi.org/10.1186/s40360-018-0216-3
- 129. Jiang Z, Cui X, Qu P, Shang C, Xiang M, Wang J. Roles and mechanisms of puerarin on cardiovascular disease: A review. *Biochem Pharmacol.* 2022;147:112655. Available at: http://doi.org/10.1016/j.biopha.2022.112655
- 130. Virojchaiwong P, Suvithayasiri V, Itharat A. Comparison of Pueraria mirifica 25 and 50 mg for menopausal symptoms. *Arch Gynecol Obstet.* 2010;284(2):411–419. Available at: http://doi.org/10.1007/s00404-010-1689-5
- 131. Cao L, Pu J, Cao Q-R, Chen B-W, Lee B-J, Cui J-H. Pharmacokinetics of puerarin in pregnant rats at different stages of gestation after oral administration. *Fitoterapia*. 2013;86:202–207. Available at: http://doi.org/10.1016/j.fitote.2013.03.004
- 132. Xiao B-X, Feng L, Cao F-R, et al. Pharmacokinetic profiles of the five isoflavonoids from Pueraria lobata roots in the CSF and plasma of rats. *J Ethnopharmacol.* 2016;184:22–29. Available at: http://doi.org/10.1016/j.jep.2016.02.027
- 133. Guerra MC, Speroni E, Broccoli M, et al. Comparison between Chinese medical herb Pueraria lobata crude extract and its main isoflavone puerarin. *Life Sci.* 2000;67(24):2997–3006. Available at:

http://doi.org/10.1016/s0024-3205(00)00885-7